

1 **Ceftazidime-Avibactam Susceptibility Breakpoints Against**
2 ***Enterobacteriaceae* and *Pseudomonas aeruginosa***

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8 **Running header:** Ceftazidime-avibactam MIC breakpoints

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20 **ABSTRACT**

21 Clinical susceptibility breakpoints against *Enterobacteriaceae* and *Pseudomonas*
22 *aeruginosa* for the ceftazidime-avibactam dosage regimen of 2000-500 mg every 8
23 hours (q8h) by 2-h intravenous infusion (adjusted for renal function) have been
24 established by the FDA, CLSI and EUCAST as susceptible, MIC \leq 8 mg/L, and resistant,
25 MIC >8 mg/L. The key supportive data from PK/PD analyses, *in vitro* surveillance
26 including molecular understanding of relevant resistance mechanisms, and efficacy in
27 regulatory clinical trials, are collated and analyzed here.

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29 **MANUSCRIPT**

30 Ceftazidime-avibactam is active *in vitro* against ESBL-, AmpC- and serine-
31 carbapenemase- (e.g. KPC-) producing *Enterobacteriaceae* and *Pseudomonas*
32 *aeruginosa*, but not metallo β -lactamase (MBL) producers (1-6). Ceftazidime-avibactam
33 clinical breakpoints of susceptible/resistant, MIC \leq / $>$ 8 mg/L (tested with a fixed
34 avibactam concentration of 4 mg/L (7)) have been assigned to both *Enterobacteriaceae*
35 and *P. aeruginosa* by the United States (US) Food and Drug Administration (FDA),
36 Clinical and Laboratory Standards Institute (CLSI), and European Committee on
37 Antimicrobial Susceptibility Testing (EUCAST) for ceftazidime-avibactam 2000-500 mg
38 q8h (8-10) based on three key data sources (11-13): probabilities of
39 pharmacokinetic/pharmacodynamic (PK/PD) target attainment (PTA) analyses;
40 multinational surveillance; and clinical trials.

41 PK/PD targets were derived from non-clinical studies for avibactam, and from non-
42 clinical and clinical studies for ceftazidime. An established target for ceftazidime, used
43 previously to support ceftazidime breakpoint determinations, is 50% $fT > MIC$ per dosing
44 interval (13-18). For avibactam to render bacteria functionally β -lactamase-negative
45 (19), it must maintain a critical threshold concentration (C_T) for 50% of the dosing
46 interval (20). Conservative C_T values for avibactam in combination with ceftazidime
47 considered to correlate with clinical efficacy have been determined as 0.5 mg/L for up to
48 3 \log_{10} CFU reduction in an *Enterobacteriaceae* hollow-fiber model and 1 mg/L for
49 bacteriostasis in a *P. aeruginosa* neutropenic mouse thigh infection model and 2 \log_{10}
50 killing in a *P. aeruginosa* neutropenic mouse lung infection model (20-22). PTA
51 analyses for both *Enterobacteriaceae* and *P. aeruginosa* used 'joint' PK/PD targets,

52 defined as simultaneous attainment of 50% $fT > MIC$ for ceftazidime and 50% $C_T > 1$ mg/L
53 for avibactam in each patient (20).

54 Population PK models for ceftazidime and avibactam were developed using PK data
55 from Phase I, II and III trials (23-25). As both drugs are excreted predominantly via the
56 kidney, the primary covariate affecting exposure is creatinine clearance (CrCL),
57 necessitating dosage adjustments for patients with CrCL < 50 mL/min (8, 9). Exposure
58 simulations for each compound in 5000 paired patients per indication (complicated intra-
59 abdominal infections [cIAI], complicated urinary tract infections [cUTI], and nosocomial
60 pneumonia [NP] including ventilator-associated pneumonia [VAP]) and renal function
61 group, incorporated Phase III patient covariate distributions appropriate to each patient
62 population and between-subject variability; exposure for both ceftazidime and
63 avibactam was simulated in the same (virtual) patients to evaluate joint PTA (25, 26).
64 Representative PTA curves in cIAI patients (the most conservative indication for PTA)
65 with normal renal function (Figures 1A and 1B) were overlaid with MIC distributions from
66 the International Network for Optimal Resistance Monitoring (INFORM) surveillance
67 program. The simulations yielded PTA $> 94\%$ against bacteria with ceftazidime-
68 avibactam MICs ≤ 8 mg/L; lower PTA values were associated with MICs of 16 mg/L or
69 ≥ 32 mg/L. Sensitivity analyses for higher PK-PD targets produced PTAs $> 90\%$ at joint
70 exposure targets up to 60% $fT > MIC$ (for ceftazidime-avibactam MICs ≤ 8 mg/L) and 60%
71 $C_T > 1$ mg/L (Figure 1C). Ceftazidime-avibactam dosage adjustments for varying degrees
72 of renal impairment also demonstrated high ($> 98\%$) PTA at MICs ≤ 8 mg/L (27).
73 Individual predicted exposures in Phase III patients showed no clinically relevant impact
74 on joint target attainment associated with disease severity, obesity, advanced age, or

75 CrCL >150 mL/min (25, 26). Hence, a susceptible breakpoint of ≤ 8 mg/L is consistent
76 with PTA values yielded by the recommended dosage regimens.

77 A key consideration in setting the clinical breakpoint for an antibacterial agent tested
78 against a particular species or group of species is where the putative breakpoint is
79 located on the MIC frequency distribution. The breakpoint should encompass the great
80 majority of the MICs of the drug against contemporary isolates (11) and should not fall
81 on a “peak” in the MIC distribution (13). The clinical breakpoint of ≤ 8 mg/L for
82 ceftazidime-avibactam vs *P. aeruginosa* straightforwardly fit these criteria as follows.
83 Against 7,062 *P. aeruginosa* isolates collected globally (ex-US) in INFORM 2012–14
84 (Figure 1B), 92.0% were susceptible to ceftazidime-avibactam (MIC₉₀ 8 mg/L) (5); more
85 recent analyses, including from the US, have reported equivalent susceptibility rates
86 (28-34). Of note, 8 mg/L is at the upper end of the ceftazidime-avibactam MIC
87 distribution, which (as stated above) is an important attribute for the clinical breakpoint
88 (12, 35).

89 In the case of *Enterobacteriaceae*, the analysis was not as straightforward, because the
90 breakpoint of ≤ 8 mg/L supported by PK/PD target attainment analyses was higher than
91 the MIC₉₀ (0.5 mg/L) by several doubling dilutions. The global (excluding the US)
92 INFORM program analyzed 34,062 *Enterobacteriaceae* isolates collected during 2012–
93 14 (Figure 1A); 99.5% were inhibited by ≤ 8 mg/L ceftazidime-avibactam (MIC₉₀ 0.5
94 mg/L) (3), with equivalent susceptibility rates reported from recent analyses, including
95 the US (29, 31-34, 36). The argument might be made therefore that a breakpoint of ≤ 0.5
96 or ≤ 1 mg/L at the upper end of the mode of MICs would be suitable for the
97 *Enterobacteriaceae*. However, the following analyses of ceftazidime-avibactam MICs

98 against genotypically- and phenotypically-characterized antibiotic-resistant sub-
99 populations among the *Enterobacteriaceae* countered that idea. Figure 1A includes
100 meropenem-nonsusceptible isolates (3), and multi-drug-resistant (MDR: resistant to ≥ 3
101 classes of antibacterial agent) isolates (6), including 816 MBL-negative meropenem-
102 nonsusceptible isolates. The 90th percentile ceftazidime-avibactam MIC for these
103 isolates was 4 mg/L, with 97.7% inhibited by ≤ 8 mg/L (3), and the MIC distribution was
104 right-shifted compared to the whole distribution, with an upper cut-off of 4–8 mg/L i.e.,
105 the susceptible breakpoint was at the upper end of, and did not divide, the MIC
106 distribution against this critical phenotypically- and genotypically-defined sub-population.

107 The 34,062 *Enterobacteriaceae* isolates (Figure 1A) also included 2,739 MDR *Klebsiella*
108 *pneumoniae* and 82 MDR *Klebsiella oxytoca*. The ceftazidime-avibactam MIC was ≤ 2
109 mg/L against 90% of these isolates, and ≤ 8 mg/L against 96.6% (6); again, the MIC
110 distribution was right-shifted compared with the overall distribution, and the susceptible
111 breakpoint was at the upper end of, but did not divide, that distribution. From these
112 analyses, it is clear that a breakpoint of ≤ 8 mg/L is necessary to encompass important
113 antibiotic resistant sub-populations such as carbapenem-resistant or MDR strains.

114 Phenotypical/genotypical sub-population analyses of *P. aeruginosa* were less helpful
115 than analyses of *Enterobacteriaceae* sub-populations, possibly because in
116 approximately 30% of ceftazidime-non-susceptible *P. aeruginosa* the ceftazidime-
117 resistance was not β -lactamase-mediated, not being reversed by combination with
118 avibactam (5).

119 Ceftazidime-avibactam MIC distributions against *Enterobacteriaceae* and *P. aeruginosa*
120 isolates from clinical trials in cIAI, cUTI or NP patients (Figure 2) were consistent with
121 global INFORM data, apart from a greater proportion of ceftazidime-avibactam-resistant
122 *P. aeruginosa*, possibly because a relatively high proportion of trial patients were in
123 Eastern Europe, where MBL-producing *P. aeruginosa* are comparatively common (37,
124 38). Across the trials, clinical and microbiological response rates were generally
125 comparable, and similar for ceftazidime-avibactam and comparator treatments. Per-
126 pathogen responses were generally similar across indications (39-44); against *P.*
127 *aeruginosa*, clinical cure (but not favorable microbiological response) rates were notably
128 lower for ceftazidime-avibactam vs meropenem in the NP trial (44). Among patients who
129 received ceftazidime-avibactam, favorable microbiological response rates were
130 generally high for infections by *Enterobacteriaceae* and more variable for *P. aeruginosa*
131 with ceftazidime-avibactam MICs ≤ 8 mg/L, including ceftazidime non-susceptible
132 isolates (Tables 1 and 2). However, consistent with other investigations (45, 46),
133 response rates by MIC did not reveal any trends, possibly because few clinical trial
134 isolates had ceftazidime-avibactam MICs > 8 mg/L, and because MIC:outcome
135 correlations may be complicated in cIAI through surgical intervention, and in cUTI
136 because of the concentration of some drugs (including ceftazidime and avibactam) in
137 urine.

138 The low rate of clinical failures is a key limitation in interpreting the PK/PD targets used
139 for PTA analyses; however, the overall high clinical/microbiological success rates are
140 broadly consistent with the PK/PD analyses using joint target attainment criteria in
141 supporting the assigned ceftazidime-avibactam susceptible breakpoint (≤ 8 mg/L)

142 against both *Enterobacteriaceae* and *P. aeruginosa*. Moreover, surveillance data
143 confirm that the MIC cutoff of ≤ 8 mg/L separates ceftazidime-avibactam resistant MBL-
144 carrying isolates from those without known ceftazidime-avibactam resistance
145 mechanisms (3, 47, 48). These breakpoints define $\geq 90\%$ of *Enterobacteriaceae* and *P.*
146 *aeruginosa* from contemporary global surveillance, including key antibiotic resistant sub-
147 populations, as susceptible to ceftazidime-avibactam (3-6, 28-34).

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163 **POTENTIAL CONFLICTS OF INTEREST**

164 W.W.N. is a former employee of AstraZeneca and current shareholder in AstraZeneca.
165 G.G.S. is a former employee of and shareholder in AstraZeneca, and current employee
166 of Pfizer. P.N., H.B., A.W. and S.D. are former employees of and current shareholders
167 in AstraZeneca. K.Y. is a former contractor for AstraZeneca. M.M. is a former employee
168 of Wright Dose Ltd, Altrincham, UK, which received funding from AstraZeneca for
169 support and assistance with the population PK analyses; she is also a shareholder in

170 AstraZeneca. T.R. is an employee of and shareholder in Allergan. I.C. is a former
171 employee of and current shareholder in Allergan.

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378 **Table 1. Patients with favorable per-pathogen microbiological response[†] at test of**
 379 **cure pooled across one Phase II and five Phase III prospective clinical trials,**
 380 **analyzed by ceftazidime-avibactam MIC**

Patients with favorable response, n/N (%)					
MIC, mg/L	<i>Citrobacter freundii</i>	<i>Enterobacter cloacae</i>	<i>Escherichia coli</i>	<i>Klebsiella pneumoniae</i>	<i>Pseudomonas aeruginosa</i>
≤0.03	1/1 (100.0)	1/1 (100.0)	66/73 (90.4)	6/6 (100.0)	-
0.06	8/8 (100.0)	2/2 (100.0)	234/257 (91.1)	28/32 (87.5)	-
0.12	8/8 (100.0)	10/12 (83.3)	163/191 (85.3)	50/58 (86.2)	-
0.25	4/6 (66.7)	17/19 (89.5)	53/59 (89.8)	19/22 (86.4)	-
0.5	5/6 (83.3)	3/5 (60.0)	16/17 (94.1)	28/35 (80.0)	2/2 (100.0)
1	1/1 (100.0)	7/7 (100.0)	3/4 (75.0)	27/29 (93.1)	10/15 (66.7)
2	-	1/1 (100.0)	8/9 (88.9)	6/6 (100.0)	34/51 (66.7)
4	-	1/3 (33.3)	1/1 (100.0)	0/2 (0)	14/20 (70.0)
8	-	-	4/4 (100.0)	-	10/15 (66.7)
16	-	-	-	-	1/3 (33.3)
32	-	-	-	-	3/3 (100.0)
>32	-	0/1 (0)	-	0/1 (0)	6/9 (66.7)

381 † Patients could have >1 pathogen. Microbiological outcomes were categorized as eradication or presumed
 382 eradication of the baseline pathogen (i.e. favorable response); persistence or persistence with increasing MIC (i.e.
 383 unfavorable response); or indeterminate.

384 Data pooled from the ceftazidime-avibactam arms of the microbiologically evaluable (ME) population of the Phase II
 385 trial in patients with cIAI (NCT00752219) (39) and the extended ME (eME) populations of the Phase III trials in
 386 patients with cIAI (RECLAIM 1&2; NCT01499290, NCT01500239, and RECLAIM 3; NCT01726023) (40, 43), cUTI
 387 (RECAPTURE 1&2; NCT01595438, NCT01599806) (41), cIAI or cUTI caused by ceftazidime non-susceptible
 388 pathogens (REPRISE; NCT01644643) (42) and NP including VAP (REPROVE; NCT01808092) (44). Intra-abdominal
 389 cultures require an invasive procedure, and were obtained only if clinically indicated; therefore, microbiological

390 responses for patients with cIAI were presumed based on clinical outcomes. n: number of favorable responses; N:
391 total number of patients for whom MIC data were available. The dashed line shows the approved ceftazidime-
392 avibactam susceptible clinical breakpoint of MIC \leq 8 mg/L applied to both *Enterobacteriaceae* and *P. aeruginosa* (8,
393 9).

394 **Table 2. Patients with favorable per-pathogen microbiological response[†] at test of**
 395 **cure for ceftazidime non-susceptible pathogens pooled across one Phase II and**
 396 **five Phase III prospective clinical trials, analyzed by ceftazidime-avibactam MIC**

Patients with favorable response, n/N (%)					
MIC, mg/L	<i>Citrobacter freundii</i>	<i>Enterobacter cloacae</i>	<i>Escherichia coli</i>	<i>Klebsiella pneumoniae</i>	<i>Pseudomonas aeruginosa</i>
≤0.03	-	-	3/4 (75.0)	1/1 (100.0)	-
0.06	-	-	8/8 (100.0)	0/1 (0.0)	-
0.12	1/1 (100.0)	-	40/48 (83.3)	15/17 (88.2)	-
0.25	1/1 (100.0)	6/6 (100.0)	27/30 (90.0)	11/12 (91.7)	-
0.5	5/6 (83.3)	2/4 (50.0)	12/12 (100.0)	25/30 (83.3)	-
1	1/1 (100.0)	7/7 (100.0)	3/4 (75.0)	27/29 (93.1)	-
2	-	1/1 (100.0)	5/5 (100.0)	6/6 (100.0)	-
4	-	1/3 (33.3)	1/1 (100.0)	0/2 (0.0)	1/4 (25.0)
8	-	-	4/4 (100.0)	-	7/9 (77.8)
16	-	-	-	-	1/3 (33.3)
32	-	-	-	-	3/3 (100.0)
>32	-	0/1 (0.0)	-	0/1 (0.0)	6/9 (66.7)

397 † Patients could have >1 pathogen. Microbiological outcomes were categorized as eradication or presumed
 398 eradication of the baseline pathogen (i.e. favorable response); persistence or persistence with increasing MIC (i.e.
 399 unfavorable response); or indeterminate.

400 Data pooled from the ceftazidime-avibactam arms of the microbiologically evaluable (ME) population of the Phase II
 401 trial in patients with cIAI (NCT00752219) (39) and the extended ME (eME) populations of the Phase III trials in
 402 patients with cIAI (RECLAIM 1&2; NCT01499290, NCT01500239, and RECLAIM 3; NCT01726023) (40, 43), cUTI
 403 (RECAPTURE 1&2; NCT01595438, NCT01599806) (41), cIAI or cUTI caused by ceftazidime non-susceptible
 404 pathogens (REPRISE; NCT01644643) (42) and NP including VAP (REPROVE; NCT01808092) (44). Intra-abdominal
 405 cultures require an invasive procedure, and were obtained only if clinically indicated; therefore, microbiological

406 responses for patients with cIAI were presumed based on clinical outcomes. n: number of favorable responses; N:
407 total number of patients for whom MIC data were available. The dashed line shows the approved ceftazidime-
408 avibactam susceptible clinical breakpoint of MIC \leq 8 mg/L applied to both *Enterobacteriaceae* and *P. aeruginosa* (8,
409 9).

410 **FIGURE LEGENDS**

411 **Figure 1. Joint PTA[†] for patients with cIAI and normal renal function receiving**
412 **ceftazidime-avibactam 2000-500 mg q8h plotted as a function of ceftazidime-**
413 **avibactam MIC (A) overlaid with the ceftazidime-avibactam MIC distributions**
414 **against *Enterobacteriaceae* (n=34,062) from the INFORM global surveillance**
415 **program, 2012–2014; (B) overlaid with the ceftazidime-avibactam MIC**
416 **distributions against *Pseudomonas aeruginosa* (n=7,062) from the INFORM global**
417 **surveillance program, 2012–2014; (C) sensitivity analysis of PTA at different joint**
418 **PK-PD targets**

419 † Defined as simultaneous attainment of 50% $fT > MIC$ of ceftazidime-avibactam for ceftazidime and 50% $fT > C_T$ of 1
420 mg/L for avibactam, with both targets having to be achieved for a simulated patient to be categorized as achieving the
421 joint target. Ceftazidime-avibactam MICs were evaluated with avibactam tested at a fixed concentration of 4 mg/L.
422 PTA was evaluated for ceftazidime-avibactam MIC values of 0.125, 0.25, 0.5, 1, 2, 4, 8, 16, 32, 64, and 128 mg/L.
423 The values above the bars are the numbers of isolates tested at each MIC. The arrows show the position of the
424 approved ceftazidime-avibactam susceptible clinical breakpoint of MIC ≤ 8 mg/L (8, 9). This set of isolates of
425 *Enterobacteriaceae* was also the source of analyses of phenotypically- and genotypically-defined resistant sub-
426 populations (3, 4, 6) as discussed in the text. The isolates of *P. aeruginosa* have been presented and analyzed in
427 detail elsewhere (5).

428 **Figure 2. Distributions of ceftazidime-avibactam MICs[†] against (A)**
429 ***Enterobacteriaceae* (n=2615) and (B) *Pseudomonas aeruginosa* (n=276) across**
430 **one Phase II and five Phase III prospective clinical trials**

431 † The ranges of MICs tested were up to 32 mg/L in the Phase II trial, and up to 256 mg/L in the Phase III trials. The
432 upper limit plotted here was >128 mg/L, for comparability with Figure 1. Three isolates of *Enterobacteriaceae* and one
433 isolate of *P. aeruginosa* from the Phase II trial tested with ceftazidime-avibactam MIC >32 mg/L and are excluded
434 from these frequency distributions.

435 Data pooled from the microbiological modified intent-to-treat (mMITT) populations of the following trials: Phase II cIAI
436 (NCT00752219) (39), Phase III cIAI (RECLAIM 1&2; NCT01499290, NCT01500239, and RECLAIM 3;
437 NCT01726023) (40, 43), Phase III cUTI (RECAPTURE 1&2; NCT01595438, NCT01599806) (41), Phase III cIAI and
438 cUTI caused by ceftazidime non-susceptible pathogens (REPRISE; NCT01644643) (42) and NP including VAP
439 (REPROVE; NCT01808092) (44). Ceftazidime-avibactam MICs were evaluated with avibactam tested at a fixed
440 concentration of 4 mg/L. The values above the bars are the numbers of isolates tested at each MIC. The arrows show
441 the position of the approved ceftazidime-avibactam susceptible clinical breakpoint of MIC ≤8 mg/L (8, 9).



