

1 **Shrinking body sizes in response to warming: explanations for the** 2 **temperature-size rule with special emphasis on the role of oxygen**

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18 19 **ABSTRACT**

20 Body size is central to ecology at levels ranging from organismal fecundity to the
21 functioning of communities and ecosystems. Understanding temperature-induced variations in
22 body size is therefore of fundamental and applied interest, yet thermal responses of body size
23 remain poorly understood. Temperature–size (T–S) responses tend to be negative (e.g. smaller
24 body size at maturity when reared under warmer conditions), which has been termed the
25 temperature–size rule (TSR). Explanations emphasize either physiological mechanisms (e.g.
26 limitation of oxygen or other resources and temperature-dependent resource allocation) or the
27 adaptive value of either a large body size (e.g. to increase fecundity) or a short development
28 time (e.g. in response to increased mortality in warm conditions). Oxygen limitation could act
29 as a proximate factor, but we suggest it more likely constitutes a selective pressure to reduce
30 body size in the warm: risks of oxygen limitation will be reduced as a consequence of
31 evolution eliminating genotypes more prone to oxygen limitation. Thus, T–S responses can be
32 explained by the ‘Ghost of Oxygen-limitation Past’, whereby the resulting (evolved) T–S
33 responses safeguard sufficient oxygen provisioning under warmer conditions, reflecting the
34 balance between oxygen supply and demands experienced by ancestors.

35 T–S responses vary considerably across species, but some of this variation is
36 predictable. Body-size reductions with warming are stronger in aquatic taxa than in terrestrial
37 taxa. We discuss whether larger aquatic taxa may especially face greater risks of oxygen
38 limitation as they grow, which may be manifested at the cellular level, the level of the gills
39 and the whole-organism level. In contrast to aquatic species, terrestrial ectotherms may be less
40 prone to oxygen limitation and prioritize early maturity over large size, likely because
41 overwintering is more challenging, with concomitant stronger end-of season time constraints.

42 Mechanisms related to time constraints and oxygen limitation are not mutually
43 exclusive explanations for the TSR. Rather, these and other mechanisms may operate in

44 tandem. But their relative importance may vary depending on the ecology and physiology of
45 the species in question, explaining not only the general tendency of negative T–S responses
46 but also variation in T–S responses among animals differing in mode of respiration (e.g. water
47 breathers *versus* air breathers), genome size, voltinism and thermally associated behaviour
48 (e.g. heliotherms).

49

50 *Key words*: Bergmann’s rule, cell size, climate warming, gigantism, growth trajectory,
51 hypoxia, life-history trade-off, phenotypic plasticity, temperature–size rule, thermal reaction
52 norms.

53

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83 **I. INTRODUCTION: THE IMPORTANCE OF TEMPERATURE–SIZE** 84 **RELATIONSHIPS**

85 Body size is central to ecology at multiple scales, from organismal fecundity to the
86 functioning of communities and ecosystems (Hildrew, Raffaelli & Edmonds-Brown, 2007).

87 Larger individuals can potentially produce more offspring, live longer, may be superior

88 competitors and be better at avoiding predators. These advantages favour growing to a large
89 size (Brown & Sibly, 2006). The drawbacks to becoming large are varied; for example,
90 growing larger takes more time, and during this time period, organisms may die or the
91 environment may become unfavourable (Blanckenhorn, 2000). Larger individuals also
92 commonly require more resources per unit time. Consequently, there is an optimal size and
93 age to reproduce, which depends on the environmental conditions that enable growth and, for
94 example, influence juvenile and adult mortality risks (Stearns, 1992).

95 Over 80% of ectothermic species examined follow the temperature–size rule (TSR), i.e. they
96 mature at a smaller size when reared in warmer conditions, despite initially growing faster
97 (Atkinson, 1994; Fig. 1A). Despite the generality of this empirical pattern (Berrigan &
98 Charnov, 1994), explaining it from life-history theory is not straightforward (Atkinson &
99 Sibly, 1997*b*; Day & Rowe, 2002). In fact, life-history optimality models commonly predict
100 that faster growth would favour animals growing to a larger size, and this is also generally
101 observed when growth rates are experimentally manipulated by altering food quantity or
102 quality (Kindlmann, Dixon & Dostalkov, 2001; Yasuda *et al.*, 2016; Diamond & Kingsolver,
103 2010). However, warming-induced reductions in body size are pervasive (Daufresne,
104 Lengfellner & Sommer, 2009) and have been termed the third universal response to warming
105 (Gardner *et al.*, 2011); the first and second universal responses to warming being directed
106 dispersal in space (range shifts) and in time (phenological shifts). Clines in body size are
107 observed across thermal geographic gradients (e.g. latitude or altitude), where small body size
108 is typically associated with warmer conditions (low latitude or altitude) and such clines are
109 referred to as Bergmann’s rule for differences among closely related species, and as James’
110 rule for differences among populations of the same species. The TSR is restricted to
111 phenotypically plastic effects that arise during ontogeny, setting it apart from James’ and
112 Bergmann’s rules, which can include ecological and evolutionary body-size responses to
113 temperature and associated climatic factors over longer timescales [see Watt, Mitchell &
114 Salewski (2010) and Pincheira-Donoso (2010) for in-depth discussions on James’ rule and
115 Bergmann’s rule and their applicability to ectotherms].

116 There is great interest in solving the life-history puzzle of the TSR, not least because more
117 than 99.9% of all species are ectotherms. Previous research on the TSR has focussed on
118 whether there is a general mechanism to explain the TSR and whether the TSR is adaptive.
119 Although the idea of a general explanation makes intuitive sense when confronted with a
120 pattern that is so pervasive, a simple, general explanation has not yet emerged. The finding
121 that size reductions with warming can be achieved at different levels of organization and
122 stages of ontogeny, and by different mechanisms (e.g. thermal responses in cell size, offspring
123 size, differences in thermal sensitivity of growth rate and development rate), has in itself been
124 used to argue that the TSR is adaptive (Atkinson, 1994; Forster & Hirst, 2012). In addition,
125 similar directions of plastic and evolved thermal responses (e.g. both becoming smaller in the
126 warm), and of latitudinal *versus* plastic responses, suggest that the TSR is likely to be
127 adaptive (Partridge *et al.*, 1994; Kingsolver & Huey, 2008; Horne, Hirst & Atkinson, 2015).

128 To understand the complex nature of thermal adaptation and the TSR better, Angilletta &
129 Dunham (2003) advocated a multivariate approach with greater emphasis on the ecological
130 context in which life histories evolve within physiological constraints set by their body plan.
131 Similarly to a recent review we highlight the role of oxygen (Audzijonyte *et al.*, 2019), but we

132 here emphasize not just temperature–size (T–S) responses induced by oxygen limitation but
133 also how T–S responses can have evolved to avoid such limitation. Additionally, we adopt a
134 broader focus beyond aquatic ectotherms to include terrestrial ectotherms. We first describe
135 what constitutes the TSR. Next, we summarize the observed variation in the strength of the T–
136 S response across groups of organisms. We then proceed to discuss how T–S responses can
137 arise from thermal influences on growth and development rates, and the adaptive value of
138 maturing at a certain size and age (Table 1). Past reviews have focussed on whether a species
139 follows the rule or not (Shelomi, 2012; see also Blanckenhorn & Demont, 2004), but we
140 consider that quantifying differences in the strength of the T–S response across groups of
141 organisms will more likely reveal the relative contributions of different explanations for T–S
142 responses. Understanding the causes of variation in the magnitude of T–S responses may lead
143 to a more complete explanation of why a reduction in body size with warming (the TSR) is
144 especially prevalent. We conclude this review by suggesting research that would best advance
145 our knowledge of temperature effects on body size.

146

147 **II. THE NATURE OF THE TSR**

148 The TSR in its simplest form describes how ectotherms develop to a smaller size for a given
149 stage, especially late in ontogeny (e.g. size at maturity) when reared under warmer conditions.
150 Size-at-stage results from the interplay between the rate of growth and the length of the period
151 spent growing, and therefore a faster growth to a smaller size in the warm (i.e. the TSR) arises
152 logically from warming stimulating development rate more than growth rate. Body-size
153 responses to temperature vary both in strength and sign (i.e. increases or decreases) across
154 species. Consequently, the field has moved to a more quantitative approach examining the
155 magnitude and direction of size responses to temperature (e.g. Forster, Hirst & Atkinson,
156 2012) rather than adopting a binary classification of whether a species is smaller or larger at a
157 given ontogenetic stage when reared in warmer conditions. In addition, most of the literature
158 focusses on size at maturity, but for organisms with indeterminate growth, T–S responses can
159 differ between size at maturity and asymptotic size, suggesting that different mechanisms are
160 involved (Hoefnagel *et al.*, 2018). T–S responses of eggs are also somewhat different
161 (weaker) than those for size at maturity (Atkinson *et al.*, 2001). The T–S response can change
162 as animals proceed through ontogeny, but in a discontinuous fashion, being more pronounced
163 in certain larval instars than others (Forster, Hirst & Atkinson, 2011a; Forster & Hirst, 2012;
164 Horne *et al.*, 2019). TSR patterns may arise not only during ontogeny, but also across
165 sequential generations, which develop at different temperatures in seasonal environments (e.g.
166 summer and winter generations in the field) (Horne, Hirst & Atkinson, 2017). Moreover, such
167 T–S responses may also be observed across populations of a species, with latitudinal clines in
168 adult body size also broadly matching plastic body-size responses to rearing temperatures
169 (Horne *et al.*, 2015). Size reductions in response to warming are also evident across species
170 within whole communities (Daufresne *et al.*, 2009). Although the mechanisms generating T–S
171 patterns within and across species could be different, the overall trends do indicate a size-
172 based filtering that favours smaller species and/or younger ages, as has been observed along a
173 latitudinal thermal cline (Zeuss, Brunzel & Brandl, 2017) and along a thermal gradient
174 associated with urbanization (Merckx *et al.*, 2018). This review focuses on plastic body-size
175 responses to temperature. However, given the concordance between the TSR, James’ rule and

176 Table 1. Overview of the different mechanisms, grouped into proximate and ultimate explanations. See text for further details.
177

Explanations	Further reading
Mechanistic (proximate) explanations	
Animals grow faster but develop even faster in warm conditions	
– Different thermal sensitivity of DNA replication <i>versus</i> protein synthesis: DNA replication (limited by enzyme kinetics) is more sensitive to temperature than protein synthesis (limited by diffusion)	Section IV.1
– At high temperatures or low oxygen, animals may preferentially allocate resources towards development and away from growth	Section IV.5
– Thermal sensitivity of growth may be reduced to prevent oxygen limitation, whereas thermal sensitivity of development may depend on genome size	Sections IV.1, IV.3 and IV.7; Fig. 4
Larger requirements for resources (food, oxygen) in warmer conditions prevent animals from growing larger	
– Different thermal sensitivity of catabolism and anabolism: growth efficiency is lower in warmer conditions	Sections IV.3
– Different thermal sensitivity of size-dependent changes in catabolism and anabolism: decline in growth efficiency with size is amplified in warmer conditions, resulting in a lower growth efficiency in warmer conditions for large (but not small) individuals	Sections IV.3
– Insufficient capacity to extract oxygen constrains animals from growing larger, even more so under warm conditions	Sections IV.2
Animals consist of smaller cells in warm conditions	
– A large genome (resulting in a larger cell size) with multiple copies ensures sufficient enzyme activity in cold conditions	Section IV.7
– Smaller cells have more membrane surface area relative to their volume supporting a greater capacity for oxygen transport in warm conditions	Section IV.7
– The ratio between oxygen supply and demand may function as a threshold for cell growth, thus regulating cell size and possibly the critical size observed in insects	Sections IV.7

178
179
180

181 Table 1. Continued.

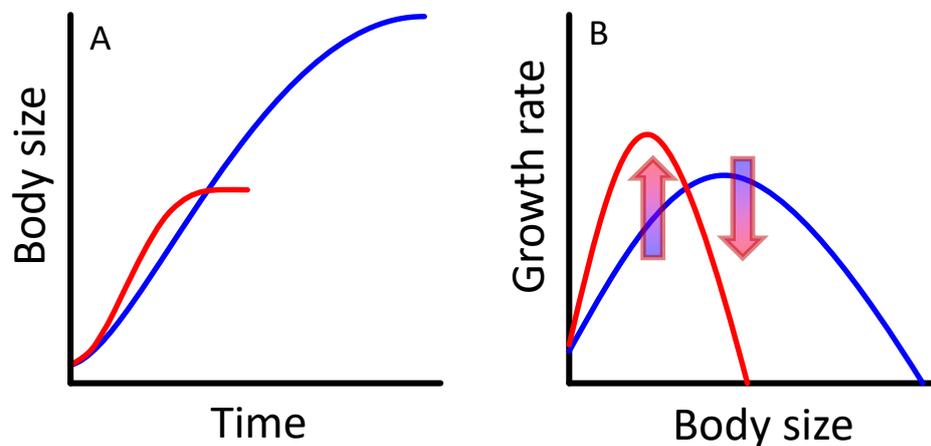
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Evolutionary (ultimate) explanations		
It becomes more advantageous to grow larger in cold conditions because of reduced mortality		
– Senescence and mortality are greater in warmer environments, favouring early maturation (at a smaller size)		Sections V.1 and V.3
It becomes more advantageous to grow larger in cold conditions because of gains in fecundity		
– Fecundity may increase more strongly with body size in cold conditions, favouring large size		Sections V.1 and V.3
It becomes more advantageous to grow larger in cold conditions because of resource limitations		
– Selection for starvation resistance typical for larger animals is stronger in cool conditions		Sections V.2 and V.3
It becomes more advantageous to produce an additional generation rather than growing to a larger size in growing populations		
– Faster maturity (at a smaller size) allows for completion of an additional generation in multivoltine species		Section VI.1; Fig. 5.
The ‘Ghost of Oxygen-limitation Past’ has led to the evolution of thermal reaction norms for adult size that are anticipatory to temperature and oxygen conditions experienced by ancestors		
– Past occurrences of oxygen limitation have selected for a canalized response with smaller sizes under warmer conditions as a compensatory response to safeguard sufficient oxygen provisioning		Sections IV.2, VI.3 and VI.4

183

184 Bergmann's rule, we also discuss explanations with an ecological and evolutionary basis,
 185 where temperature is involved only indirectly (e.g. as a cue for seasonal progression and for
 186 time remaining to complete development).

187 Finally, the TSR is only a puzzle when evaluated under benign conditions, including
 188 non-stressful temperatures and non-limiting resource supply (Atkinson 1994; Walczyńska,
 189 Kielbasa & Sobczyk, 2016). For example, when high temperatures impair growth, rather than
 190 stimulate it, life-history theory predicts animals to mature at a smaller size. Similarly, when
 191 warming alleviates cold, stressful temperatures, it may result in animals growing to a larger
 192 body size (Forster, Hirst & Woodward, 2011*b*). Ectotherms that follow the TSR, grow faster
 193 but to a smaller size in warmer conditions. Therefore, effects of temperature on growth differ
 194 throughout ontogeny: at earlier or smaller life stages temperature stimulates growth while at
 195 later or larger life stages temperature reduces growth (Fig. 1B). Thus, understanding the
 196 effects of temperature on size needs to incorporate interactions between time, temperature and
 197 body size.
 198



199 **Fig. 1.** Thermal responses in body size (A) and growth rate (B). Responses are indicated
 200 for warm (red lines) and cold (blue lines) conditions. Arrows in B indicate that effects of
 201 warming are contingent on body size (and hence on time during ontogeny), stimulating
 202 growth during small, early life stages (upward arrow), but reducing growth later in later,
 203 larger life stages (downward arrow). Note that this is a simplified schematic and in
 204 reality, the temperature–size rule (TSR) may progress irregularly over ontogeny (see
 205 Forster *et al.*, 2011*a*; Horne *et al.*, 2019).
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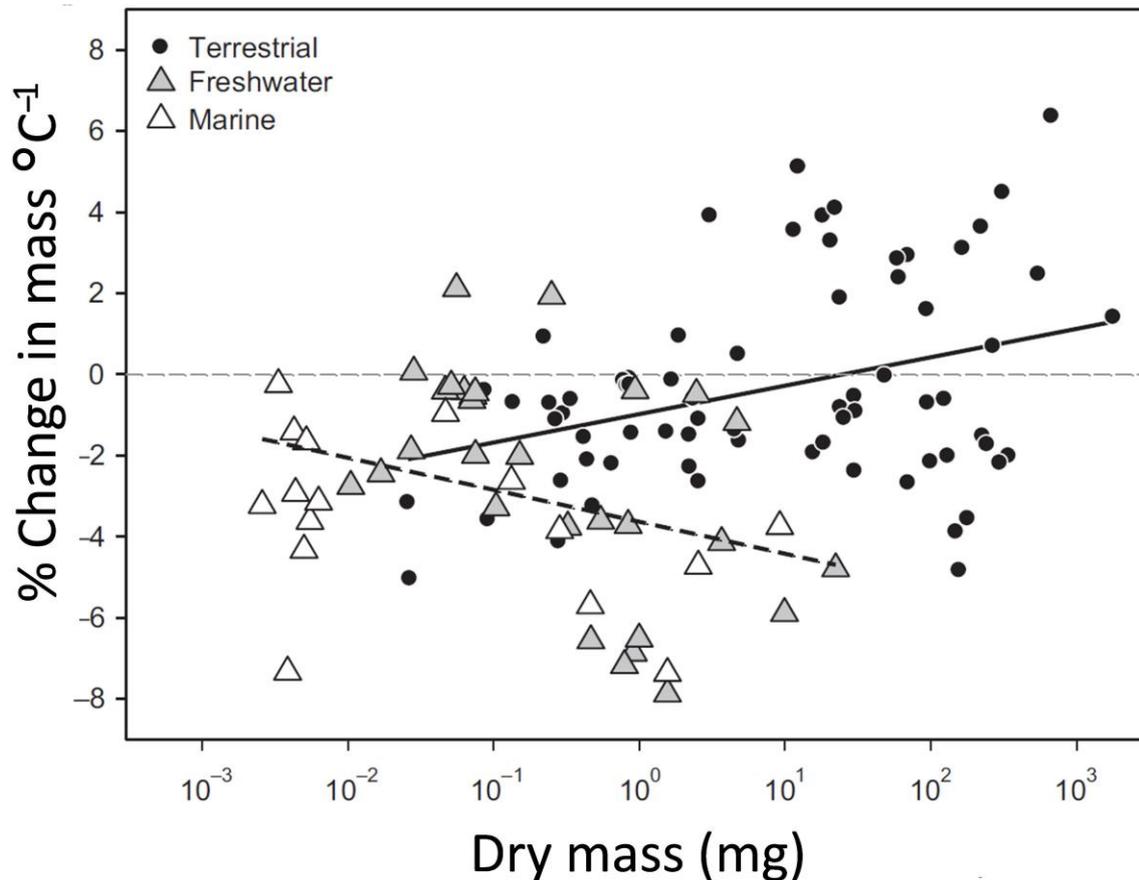
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208 III. PATTERNS IN T–S RESPONSES

209 (1) T–S responses due to phenotypic plasticity

210 Although adult body size is usually reduced under warmer rearing conditions (i.e. following
 211 the TSR), we will also describe the substantial variation in responses across different taxa and
 212 environments (terrestrial, aquatic). Taking a meta-analytical perspective, Horne *et al.* (2015)
 213 extended the work of Forster *et al.* (2012) and Klok & Harrison (2013), to find distinct
 214 patterns in the extent to which body size responds to temperature across taxonomic groups of
 215 arthropod species. A primary finding was that T–S responses became more negative (stronger
 216 TSR) in aquatic arthropods with increasing body size (Fig. 2). In terrestrial arthropods, this
 217 pattern with body size appeared to be reversed. However, body size and voltinism (i.e. the

218 number of generations of an organism in a year) tend to co-vary (univoltine species, with one
 219 generation per year, are typically larger than multivoltine species, which have more than one
 220 generation per year). In terrestrial arthropods, voltinism has been found to be a stronger
 221 predictor than body size, with univoltine species often displaying the reverse T–S response,
 222 such that they commonly mature at a larger size in the warm. Thus, within aquatic arthropods,
 223 the T–S response appears to become stronger with increasing body size, whereas within
 224 terrestrial arthropods the opposite pattern is found, with the T–S response weakening and
 225 eventually reversing with increasing body size (Fig. 2).
 226



227
 228 **Fig. 2.** Temperature–size (T–S) responses (% change in body mass per °C) for
 229 terrestrial (black circles), freshwater (grey triangles) and marine (white triangles)
 230 arthropod species, plotted against their dry mass (standardized to 20 °C) With increasing
 231 body mass, T–S responses became more negative in aquatic arthropods (dashed line;
 232 $F_{1,43} = 5.40$, $P = 0.02$, $r^2 = 0.09$), but in terrestrial arthropods they became more
 233 positive (solid line; $F_{1,69} = 9.28$, $P = 0.003$, $r^2 = 0.11$). Figure reprinted from Horne *et al.*
 234 (2015) with permission from John Wiley & Sons Ltd/CNRS.

235

236 (2) T–S responses across populations, species and communities

237 Horne *et al.* (2015) report a concordance between phenotypically plastic size responses to
 238 temperature (the TSR) and latitudinal clines in body size (i.e. James' rule). Obviously,
 239 latitudinal size clines could be related to various factors other than temperature, which also
 240 co-vary with latitude (e.g. duration of growth season, day length, food availability, potential
 241 evapotranspiration, and thermal fluctuations), and the mechanisms could likewise differ as

242 they apply to differences across populations. For example, dispersal could obscure spatial
243 relationships between environmental temperature and body size (Horne, Hirst & Atkinson,
244 2018), as has been suggested for altitudinal clines in body size within several species of
245 grasshoppers differing in dispersal potential (Levy & Nufio, 2015) and latitudinal clines
246 across dytiscid beetle species (Pallarés *et al.*, 2019). Still, the correspondence noted by Horne
247 *et al.* (2015) suggests that these body-size responses across individuals, populations and
248 species may share at least some of the same temperature-related drivers. This makes it
249 informative to compare T–S responses at the population and species level across aquatic and
250 terrestrial groups of different body size.

251 Makarieva, Gorshkov & Li (2005) showed that the largest terrestrial ectotherm species
252 tend to live in the warm tropics. By contrast, in a variety of animal groups, aquatic species of
253 gigantic proportions have been documented in cold, polar regions (Moran & Woods, 2012).
254 These contrasting geographical trends in maximum body size can be seen as a special case of
255 the more general pattern in which T–S responses across latitudinal clines become increasingly
256 negative in larger-bodied taxa in aquatic but not terrestrial habitats. Similarly, among aquatic
257 amphipod communities, stronger T–S responses were observed for the largest species (a
258 sixfold change), while changes in median body size were less pronounced (2.6-fold change)
259 (Chapelle & Peck, 2004). In summary, the pattern of intraspecific T–S responses becoming
260 stronger with increasing body size in water but not on land is also observed across species and
261 across communities. This concordance across ecological levels of organization could be a
262 coincidence or could reflect similar drivers and constraints.

263

264 **IV. THE DEPENDENCY OF T–S RESPONSES ON GROWTH AND** 265 **DEVELOPMENT**

266 **(1) Growth and development rates have different thermal sensitivities**

267 Differences in thermal sensitivity of growth and development rates give rise to T–S
268 responses (Forster *et al.*, 2011*b*; Banas & Campbell, 2016; Hoefnagel *et al.*, 2018; Fig. 3), and
269 many explanations therefore focus on explaining differences in the thermal sensitivity of
270 growth and development (Table 1). Instead of differences in their thermal dependency,
271 Walters & Hassall (2006) argued for a focus on differences between the minimum threshold
272 temperature for growth and that for development (i.e. the temperature below which growth
273 and development are arrested). Indeed, when growth and development rates change linearly
274 with temperature, a decrease in the ratio between growth rate and development rate with
275 warming is equivalent to a greater threshold temperature for development rate than for growth
276 rate. However, different threshold temperatures for growth and development are not a
277 necessary condition for T–S responses to arise when thermal dependencies are non-linear (e.g.
278 Forster *et al.*, 2011*b*; see also Kutcherov, Lopatina & Kipyatkov, 2011; Sweeney *et al.*, 2018).

279 Van der Have & de Jong (1996) suggested that protein synthesis, involved in both cell
280 and organism growth, is limited by temperature-insensitive diffusion of heavy ribosomal sub-
281 units, and that DNA replication – central to cell division, differentiation, and rate of
282 organismal development towards maturity – is instead limited by the more thermally sensitive
283 rates of DNA polymerase activity. Consequently, they argued that DNA replication (related to
284 differentiation) was more temperature sensitive than protein synthesis (related to growth),
285 thereby linking whole-organism growth and development to the kinetics of individual

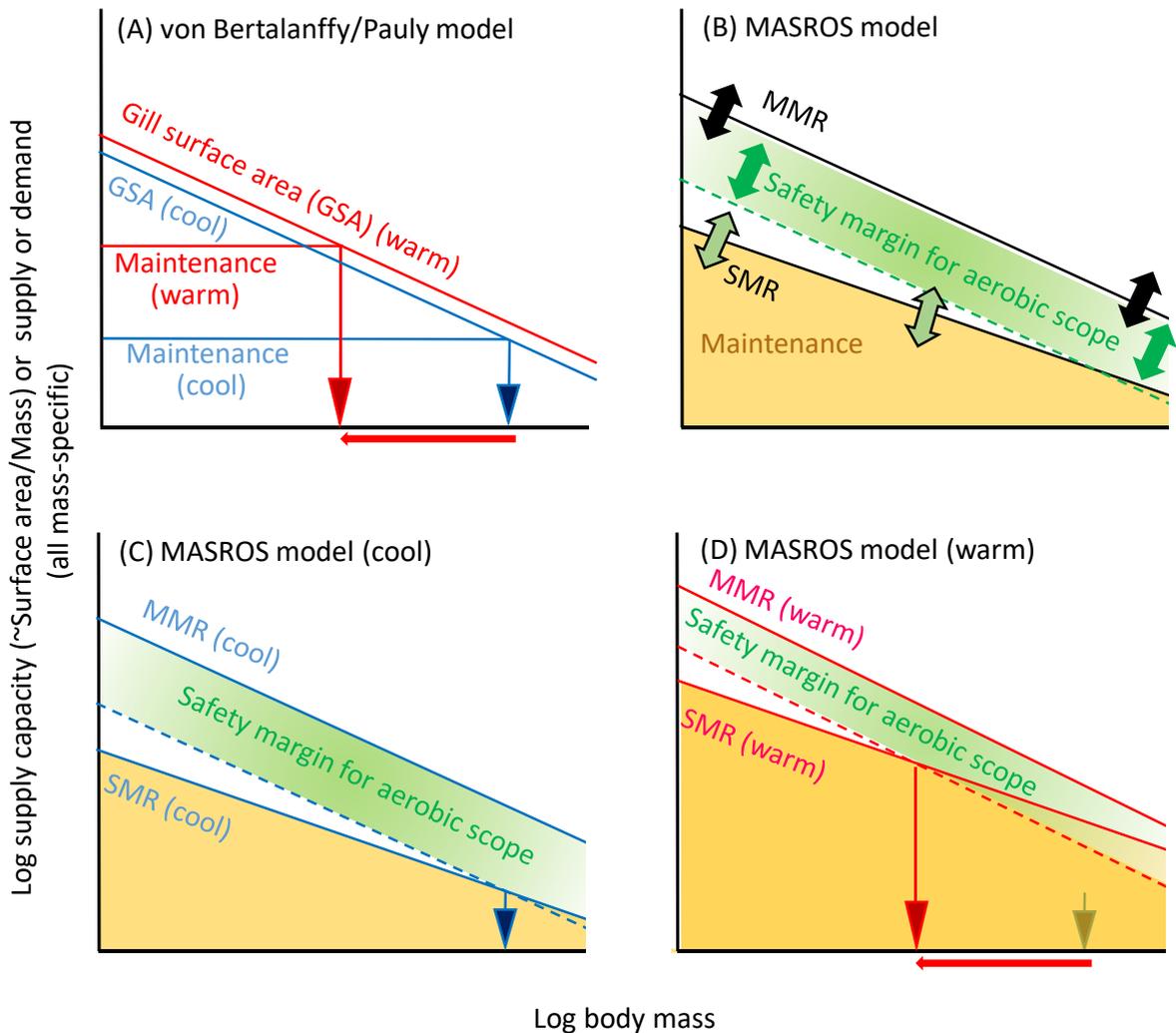
286 enzymes. This mechanism may contribute to producing negative T–S responses. However,
 287 given the extant variation in the strength of the T–S response across different groups of taxa
 288 and during ontogeny (see Section III) differences in thermal sensitivity can be modulated (see
 289 also Section IV.2). Modulation of the thermal sensitivity of growth and development is
 290 perhaps most obvious in unicells (with binary division). Since the offspring size is half that of
 291 the parent cell, the ratio between specific growth rates and development rates equals 2 or they
 292 would increase or decrease in body size *ad infinitum*. Hence, they can only achieve a T–S
 293 response if temperature shifts the ratio between growth rates and development rates away
 294 from 2 temporarily (Forster *et al.*, 2011a; Forster, Hirst & Esteban, 2013). Thus, a greater
 295 thermal sensitivity of development rate relative to growth rate may arise partly because of
 296 differences in kinetics of DNA replication and protein synthesis, but other additional
 297 explanations are required to explain the variation in T–S responses.

298

299 **(2) Are there insurmountable constraints on growth?**

300 Debates continue on whether or not growth rate is increasingly constrained during ontogeny,
 301 and if so, whether or not warmer temperature increases these constraints, leading to smaller
 302 size at maturity or final size (Pauly, 1998; Lefevre, McKenzie & Nilsson, 2017; Pauly &
 303 Cheung, 2018; Audzijonyte *et al.*, 2019). The debated constraint is geometric, based on
 304 reductions in the surface area to volume ratio as size increases, which has been called a
 305 ‘dimensional tension’ by Pauly & Cheung (2018). The diminishing ratio of surface area to
 306 volume has been argued to result in resource limitation – food limitation owing to insufficient
 307 area of the digestive tract, or oxygen limitation owing to insufficient area of respiratory
 308 surfaces (Kooijman, 2010; Pauly, 2010). Oxygen limitation has been emphasized in aquatic
 309 species, which can expend non-trivial proportions of their energy budget obtaining oxygen
 310 (von Bertalanffy, 1960; Pauly, 2019). This geometric constraint is used to explain growth
 311 deceleration during ontogeny up until maximum size where growth is no longer possible. At
 312 this point, the supply of resources available for growth and other routine metabolism [upper
 313 line at each temperature in Fig. 3A, corresponding to ‘anabolism’ of von Bertalanffy (1960)
 314 and assimilation of Kooijman (2010)] has converged with the line representing non-growth or
 315 ‘maintenance’ resource demand [lower line at each temperature in Fig. 3A, corresponding to
 316 ‘catabolism’ of von Bertalanffy (1960)]. Added to the dimensional tension is the idea that
 317 resource uptake has a rate-limiting step that is less sensitive to temperature (e.g. diffusion)
 318 than is resource demand (e.g. rate of enzymatic reactions). Consequently, maximum resource
 319 supply increases relatively little as temperatures rises (from blue to red), compared with the
 320 greater increase of maintenance costs. Oxygen diffusion in water is relatively temperature
 321 insensitive, accelerating only by about 10% with 10 °C warming (Verberk *et al.*, 2011),
 322 contrasting with the approximate doubling of metabolic rate (Seebacher, White & Franklin,
 323 2015). However, the uptake of food resources is less likely to be widely thermally insensitive,
 324 varying with feeding mode (Dell, Pawar & Savage, 2014) and how temperature affects food
 325 availability (i.e. the balance of food production to consumption).

326 Constraints on oxygen supply have been described as ‘insurmountable’ (Pauly, 1998;
 327 Lefevre *et al.*, 2017) or ‘uncircumventable’ (Pauly & Cheung, 2018). Yet they are not
 328 completely insurmountable. Organisms have evolved the capacity to: (i) increase surface area
 329 for resource uptake during growth, such as by changing body shape (Hirst, Glazier &



330
 331 **Fig. 3.** Role of constraints in the von Bertalanffy/Pauly model (A) and the maintain
 332 aerobic scope and regulate oxygen supply (MASROS) model (B–D). In A, constraints on
 333 growing to a larger size are considered to be insurmountable, arising from geometric
 334 constraints on gill surface area scaling, and growth ceases when maintenance
 335 metabolism converges to supply capacity. Maintenance is here considered to fuel
 336 essential processes such as maintenance of electrochemical gradients, protein synthesis,
 337 and repair. In the MASROS model, animals still have aerobic scope left when reaching
 338 maximum size, which is considered to be a safety margin when animals face demanding
 339 but transient conditions (e.g. disease, episodes of hypoxia, predator attack, and possibly
 340 part of reproduction). Aerobic scope not reserved for the safety margin (in white) can be
 341 used to fuel growth and other routine activities (e.g. activity, digestion and possibly part
 342 of reproduction). Evolution is thus assumed to have modified growth trajectories to avoid
 343 oxygen limitation. Growth trajectories can be modulated by adaptive changes in the
 344 scaling of standard metabolic rate (SMR), maximum metabolic rate (MMR) or the width of
 345 the safety margin for aerobic scope. Warm conditions (shown in red), may lead to growth
 346 to a smaller size if the thermal sensitivity of maintenance (SMR) is higher than that of
 347 supply (MMR). This size decrease could be partly compensated for by allowing a
 348 reduction in the safety margin (panel D). Note that the slopes of the lines (i.e. the scaling
 349 exponents) can also vary with temperature, but are here kept constant for reasons of
 350 clarity.
 351

352 Atkinson, 2014; Glazier, Hirst & Atkinson, 2015) or increasing the size of uptake organs
353 (Antoń *et al.*, 2020) or their surfaces (gill re-modelling; Nilsson, Dymowska & Stecyk, 2012);
354 (ii) increase rates at which they obtain and distribute resources (e.g. by increasing feeding
355 activity or by active ventilation and circulation; Woods & Moran, 2020); and (iii) reduce rates
356 of demand for resources (e.g. less locomotion, or lower mitochondrial density). These
357 adjustments to the rate of resource uptake and demand reduce the likelihood that the
358 constraints will be observed directly in controlled laboratory studies of the TSR, which
359 provide abundant food, levels of oxygen availability typical for the species, non-extreme
360 ('physiological range') temperatures and an absence of predators and disease. Even in the
361 field, such constraints may be observed only occasionally. On the other hand, overcoming or
362 avoiding these physical constraints is unlikely to be cost-free. Thus, organisms may have
363 adapted so that they are not 'panting for breath' during normal growth, but could nonetheless
364 experience resource limitation under more demanding conditions (e.g. when pursued by
365 predators or encountering pathogens). Although such demanding conditions may be rare, they
366 are also disproportionately detrimental. Organisms should therefore maintain a safety margin
367 (e.g. aerobic scope) to prevent resource limitation of growth and reduction in fitness, e.g. the
368 maintain aerobic scope and regulate oxygen supply (MASROS) model, in which size is
369 adjusted to maintain sufficient oxygen supply relative to demand (Atkinson, Morley &
370 Hughes, 2006; Fig. 3B). Selection to avoid resource limitation may favour adaptive
371 modulation of growth in response to temperature especially when temperature is a reliable cue
372 (i.e. temperature has correlated with fitness benefits from developing faster or maturing at a
373 smaller size during the population's evolutionary history). This adaptive response to *avoid*
374 resource limitation or other harm represents an important conceptual distinction. Instead of
375 direct constraints on growth, we here emphasize the evolution of adaptive reaction norms in
376 which growth responds to temperature as a cue to avoid harm.

377 Evolutionary adjustments (double-headed arrows in Fig. 3B) can be made to resource
378 uptake capacity (upper boundary of safety margin), to the size of the safety margin (height of
379 green shaded area at different body sizes), and to the amount of other non-growth investment,
380 sometimes referred to as 'maintenance' (height of orange shaded area at different body sizes).
381 All of these can shape the resulting growth trajectories. Thermal responses in growth
382 trajectories are the evolutionary outcome that temperature has had on these factors (Fig. 3C
383 and D). This adaptive perspective should also be applicable to other potential constraints
384 affecting the evolution of the TSR (e.g. temperature-dependent uptake of food resources,
385 whose safety margin is set by extra feeding and assimilation capacity and by the amount of
386 stored reserves; or by viscosity affecting oxygen supply; Verberk & Atkinson, 2013). In
387 summary, simultaneous adaptive modulation of growth, maintenance and a safety margin
388 reflects the evolutionary effects of past size- and temperature-dependent constraints on
389 resource availability and other selection pressures. Evolved plastic responses of growth to
390 temperature can mitigate current or predictable future resource limitations, thereby avoiding
391 constraints on whole-organism growth. Adaptive modulation of growth trajectories likely also
392 integrates other fitness-enhancing activities such as reproductive development or
393 reproduction, leading to a deceleration of growth with increasing body size (Kozłowski,
394 Czarnoński & Danko 2004; Kooijman, 2010; Marshall & White, 2019).

395

396 (3) Thermal responses in growth rate

397 In order to grow, organisms need resources such as food and oxygen, which together
398 shape the energy budget of an organism. Changes in the energy budgets and energy allocation
399 with temperature have been used to explain the TSR (e.g. Pauly, 2010; see Section IV.2).
400 Much of this work can be traced back to the work of von Bertalanffy (1960) and Pütter (1920)
401 who noted that somatic growth must be equal to the difference between anabolism and
402 catabolism, although part of the energy surplus must also be allocated to reproductive growth
403 (Kozłowski, Czarnołęski & Danko, 2004; Marshall & White, 2019). If catabolism increases
404 relative to anabolism with increasing body mass, a decrease in body size with warming could
405 then arise when temperature stimulates catabolism more than anabolism (von Bertalanffy,
406 1960) or when temperature stimulates resource demand more than supply (DeLong, 2012;
407 Fig. 3A). Angilletta & Dunham (2003) argued that while warming could increase absolute
408 growth rates, warming must also, according to von Bertalanffy's growth model, reduce net
409 growth efficiency (expressed as the percentage of biomass produced relative to total energy
410 absorbed), as relatively more energy is spent on catabolism with warming. However, their
411 analysis of published data on growth efficiency did not find the expected decrease in net
412 growth efficiency with warming. A potential resolution to this problem is that the thermal
413 dependency of net growth efficiency is itself size dependent. Consequently, the decline in
414 growth rates and growth efficiency observed with increasing size should be more pronounced
415 under high temperatures (e.g. Perrin, 1988; Panov & McQueen, 1998; Kozłowski *et al.*, 2004;
416 Hoefnagel *et al.*, 2018). It has been suggested that larger organisms have smaller net energy
417 balances in warm conditions because oxygen demand increases with temperature relative to
418 oxygen supply (Pedersen, 1987; Pörtner, 2001; Pauly, 2010; Verberk *et al.*, 2011; Verberk &
419 Atkinson, 2013). However, whether the decline in growth rates and growth efficiency with
420 increasing size is constrained by resource limitation is still debated (see Section IV.2). A role
421 for oxygen in generating the TSR may explain the stronger T–S responses observed in aquatic
422 taxa compared to terrestrial taxa (Forster *et al.*, 2012; Horne *et al.*, 2015; Rollinson & Rowe,
423 2018), owing to the greater challenges of breathing underwater (lower diffusion rates, larger
424 costs of ventilation) (Dejours, 1981; Verberk *et al.*, 2011; Verberk & Atkinson, 2013). Few
425 studies have tested interactive effects of oxygen and temperature on growth and size at stage,
426 but the few that have demonstrate that T–S responses depend on oxygen conditions in aquatic
427 isopods (Hoefnagel & Verberk, 2015), and in air-breathing fruit flies (Frazier, Woods &
428 Harrison, 2001). Size reductions with warming were more pronounced under hypoxia and less
429 pronounced – or reversed – under normoxia and hyperoxia. This suggests either a direct role
430 of oxygen in generating the TSR (i.e. the strongest T–S response is observed under conditions
431 where resource limitation is most likely), or that oxygen limitation has acted as a selection
432 pressure on growth trajectories, and animals use temperature and oxygen conditions as cues to
433 modulate growth. Given that effects of hyperoxia are much weaker (but usually opposite) to
434 those of hypoxia, oxygen limitation as a selection pressure on growth seems more likely.
435 Indeed, direct evidence that individuals become more prone to warming-induced oxygen
436 limitation as they grow larger is scarce, and may differ between aquatic ectotherms (e.g. fish)
437 and terrestrial ectotherms (e.g. insects), as the costs of increasing oxygen uptake are greater in
438 water than in air (Verberk & Bilton, 2013; Verberk & Atkinson, 2013).

439 Fish appear to adhere to the TSR (see Section IV.4), but size dependency of oxygen
440 supply capacity in fish has been debated (see Section IV.2). Since fish can dynamically alter
441 their gill surface area, it is unlikely that constraints are completely insurmountable, but gill
442 proliferation also carries costs, such as the cost of maintaining ion homeostasis and water
443 transport, increased exposure to toxic substances in the water, and increased risk of disease
444 and parasitism (Nilsson *et al.*, 2012; Audzijonyte *et al.*, 2019). Since excessive oxygen itself
445 is toxic, the act of balancing toxicity and asphyxiation risks may also directly reduce
446 performance of animals with an excess capacity for oxygen uptake (Verberk & Atkinson,
447 2013). To explain the TSR from an oxygen-limitation perspective, these costs and benefits of
448 altering capacity for oxygen uptake must be size and temperature dependent. Most studies
449 focus on two-way interactions, rather than the three-way interaction between size, temperature
450 and oxygen (Woods & Moran, 2020). Boundary layers at the gill surface affect uptake
451 capacity in such a size- and temperature-dependent manner; they result from viscosity and
452 impede oxygen diffusion, especially in colder, more viscous water, and smaller animals are
453 disproportionately affected (Verberk & Atkinson, 2013). Consequently, larger individual fish
454 in warmer waters could have a lower aerobic scope or a higher sensitivity to oxygen limitation
455 (Rubalcaba, J.G., Verberk, W.C.E.P., Hendriks, A.J., Saris, B. & Woods, H.A., in
456 preparation). There is also evidence that larger individuals are more prone to oxygen
457 limitation in some fish species (Reid *et al.*, 2013; Bursleson, Wilhelm & Smatresk, 2001; Robb
458 & Abrahams, 2003), but it is difficult to generalize this to all fish, given the many different
459 strategies for coping with hypoxia (Chapman & McKenzie, 2009). Indeed, fish may deal with
460 hypoxic stress in a size-dependent manner, with larger animals relying more on anaerobic
461 metabolism (Goolish, 1989; Urbina & Glover, 2013; Lv *et al.*, 2018). On the relatively short
462 timescales typical for hypoxia-tolerance assays, larger fish could supplement their energy
463 needs with anaerobic metabolism; on longer timescales of growth and development, a lower
464 aerobic scope of larger fish in warm waters could reduce growth. This is an area in need of
465 more empirical data.

466 In terrestrial ectotherms such as many insect species, evidence that risks of oxygen
467 limitation increase as they grow larger is scarce, possibly because animals can compensate in
468 a range of ways (e.g. by increasing capacity for ventilation and circulation; see Harrison,
469 Greenlee & Verberk, 2018). In the grasshopper *Schistocerca americana*, hypoxia sensitivity
470 (used here as a proxy for risks of oxygen limitation) was highest in the youngest instars which
471 lack air sacs and rely more on diffusive gas exchange (Greenlee & Harrison, 2004). Larger
472 individuals tend to employ convective gas exchange, which could explain their lower
473 sensitivity to hypoxia. Also across species, there is little evidence for size dependency of
474 hypoxia sensitivity. For example, Harrison, Klok & Waters (2014) found the critical oxygen
475 partial pressure (pO_2) for metabolism to be independent of adult body size across a range of
476 insect species. Larger species likely prevent progressive oxygen limitation with increasing
477 body size by having greatly increased tracheal dimensions and these do appear to set upper
478 limits to the size that insects may attain (Kaiser *et al.*, 2007). In cases where oxygen limitation
479 is less of a constraint on growth, patterns of larger species at higher temperatures have been
480 explained by the need to maintain metabolism (expressed per gram of body tissue) within an
481 optimal range, as increasing body size reduces metabolism, counteracting the increased
482 metabolism associated with higher temperatures (Makarieva *et al.*, 2005). In summary, in

483 terrestrial ectotherms, several reasons may explain why sensitivity to low oxygen is decoupled
484 from size, although upper size limits may still be set by limits to tracheal expansion.

485 If larger aquatic species are indeed more challenged to provision their tissues with
486 adequate oxygen to maintain sufficient aerobic scope (see also Section IV.2), this would
487 provide an explanation for stronger T–S responses with increasing body size in these taxa
488 (Fig. 2; see also Section IV.4). Similarly, if larger terrestrial species are less challenged by
489 oxygen limitation because of increased reliance on convective transport, this could also
490 explain why T–S responses weaken and then reverse with increasing species body size in
491 terrestrial arthropods (Fig. 2; Klok & Harrison, 2013).

492

493 **(4) Fish and the temperature–size rule**

494 Fish have been documented to adhere to the TSR (e.g. Trexler, Travis & Trexler, 1990;
495 Dhillon & Fox, 2004; Loisel, Isla & Daufresne, 2019), and thermal clines in the field are often
496 related to size clines (Daufresne *et al.*, 2009; Baudron *et al.*, 2014; Van Rijn *et al.*, 2017;
497 Moffett *et al.*, 2018; but see Belk & Houston, 2002). Latitudinal clines in fish body size have
498 also been documented, but other factors may play a role here. For instance, larger fish may
499 have a greater capacity to disperse to higher latitudes (Weber *et al.*, 2015), increased mortality
500 in warmer areas may select for individuals to mature faster at a smaller size (Heibo,
501 Magnhagen & Vøllestad, 2005), and warming may produce opposite effects in species with
502 contrasting thermal niches (Rypel, 2014). Moreover, fishing pressure may greatly affect size
503 distributions in the field, confounding, blurring or strengthening patterns in body size related
504 to temperature (Tu, Chen & Hsieh, 2018; Cheung *et al.*, 2013).

505 Clearly, the existence of large species living in warm tropical waters indicate
506 evolutionary capacity to overcome constraints on growth to a large size. Hence, these
507 constraints are not insurmountable from an evolutionary perspective (see Section IV.2).
508 Instead, adaptive evolution can enhance the capacity to supply oxygen depending on the
509 lifestyle (Seibel & Deutsch, 2020). Different adaptations can enhance oxygen supply such as
510 planktonic feeding with greatly enlarged gills (e.g. whale shark), ram ventilation (e.g. tuna,
511 marlin) or adopting a sluggish lifestyle as adults (groupers). While large fish species such as
512 those mentioned above can clearly live in warmer waters (an interspecific pattern), it is
513 unknown whether they will show a stronger T–S response (an intraspecific pattern). They
514 could grow larger still when reared under colder conditions, but such experiments would be
515 logistically challenging: consequently, T–S responses for size at maturity are recorded only up
516 to the size of small fish or large insects. A major issue is therefore predicting the extent to
517 which strengthening T–S responses with species body size in aquatic ectotherms will extend
518 further to include commercial fish and aquaculture species. A recent study indicated that
519 responses in mean fish size to temperature were weakening and reversing towards larger sizes
520 (Audzijonyte *et al.*, 2020), although it is unclear how results on mean size relate to size at
521 stage (e.g. maturity or maximum). Concordant with a role for oxygen, Van Rijn *et al.* (2017)
522 focussing on maximum size in the field, found greater T–S responses in more active fish
523 species. It is challenging, however, to isolate effects of temperature on body size in field data
524 where responses could also reflect (size-dependent) species interactions, dispersal, differences
525 in productivity and length of the growing season. Therefore, rearing experiments under

526 controlled conditions should help us understand physiological mechanisms better (Ohlberger,
527 2013; Edeline *et al.*, 2013; Knouft, 2014).

528

529 **(5) Thermal responses in development rate**

530 Effects of oxygen and food are not limited to growth rate, but may also act on
531 development (Table 1). Callier & Nijhout (2011) showed that in growing caterpillars of
532 *Manduca sexta*, the decision to moult or pupate is size and oxygen dependent. As animals
533 increase their body mass, their demand for oxygen also increases, but since the tracheal
534 system can only be enlarged upon moulting they cannot correspondingly increase the capacity
535 for oxygen supply. Such low capacity for oxygen supply relative to demand triggers the
536 endocrine cascade that advances development (see also Callier *et al.*, 2013; Kivelä *et al.*,
537 2018). Greenberg & Ar (1996) found that the mealworm beetle (*Tenebrio molitor*) developed
538 into smaller adults when reared under hypoxia, but more than doubled the number of moults
539 to get there compared to normoxia. Under hyperoxia there were fewer moults, supporting the
540 idea that oxygen availability directly influences developmental processes.

541 External resource conditions, such as environmental hypoxia or food conditions,
542 appear to affect development less compared to their effects on growth rates. Development rate
543 is generally more sensitive to temperature than growth rate, and this temperature dependence
544 also appears to vary less across ontogeny (e.g. Horne *et al.*, 2019). In addition, stimulating
545 effects of temperature on growth seemed to level off with increasing temperature (De Block
546 & Stoks, 2003) and increasing body size during ontogeny (Forster *et al.*, 2012), which
547 suggested resource limitation (or responses to avoid it; see Section IV.2) under these
548 conditions. By contrast, thermal effects on development did not suggest resource limitation or
549 its avoidance. Changes in development rates across populations occupying different positions
550 along a latitudinal or altitudinal cline suggest adaptive modulations of development.
551 Substantial counter-gradient variation in development rate has been reported across latitudinal
552 and altitudinal clines (e.g. faster development of high-latitude populations), likely as an
553 adaptation to the shorter growing season at high altitudes and latitudes (Dingle & Mousseau,
554 1994; Chown & Klok, 2003; Berner & Blanckenhorn, 2006; Blanckenhorn & Demont, 2004;
555 Ayres & Scriber, 1994; Kivelä *et al.*, 2011; Parson & Joern, 2014; Buckley *et al.*, 2015). Co-
556 gradient variation in development rate has also been reported, but again to resolve time
557 limitations (i.e. faster development in warmer, but ephemeral habitats) (Dittrich *et al.*, 2016).
558 Heliotherms prefer and reach high operative body temperatures *via* basking. This could select
559 for a reduced development rate, or a lower thermal sensitivity of development which avoids
560 leaving insufficient time for completing growth in terms of mass, thus explaining converse
561 TSR in heliotherms (see Section IV.6). In summary, responses of development rate to
562 temperature appear to be adapted to duration of the growing season. Compared to growth,
563 development is relatively insensitive to availability of environmental resources.

564

565 **(6) Are grasshoppers an exception to the temperature–size rule?**

566 A notable exception to the near-universal pattern of size reductions with warming are the
567 grasshoppers. Grasshoppers could be less inclined to follow the TSR for several reasons.

568 First, oxygen may be less limiting in larger terrestrial arthropods such as grasshoppers,
569 because gas exchange in their tracheal network relies more on convection (Greenlee &

570 Harrison, 2004). This could at least partially explain why they differ from aquatic
571 counterparts, but is unlikely to be the complete reason, as plenty of large tracheated
572 arthropods do follow the TSR.

573 Second, grasshoppers are heliotherms and have a high preferred body temperature,
574 sometimes as high as 38 °C (Miller *et al.*, 2009). Heliotherms will likely also experience
575 larger variations in body temperature than other ectotherms. Under widely fluctuating
576 temperatures, the realized thermal performance curve for growth is different from the thermal
577 performance curve under constant temperatures due to Jensen's inequality (Denny, 2017),
578 reaching peak performance at a lower temperature. To compensate, heliotherms likely have a
579 thermal performance curve with a peak shifted to higher temperatures and since most TSR
580 rearing experiments employ constant temperatures, it is unlikely that the higher rearing
581 temperatures will coincide with limitations for resource supply and thus strongly stimulate
582 growth. In addition to growth being highly responsive to temperature, development may be
583 less responsive to temperature in heliotherms: the operative body temperatures of heliotherms
584 may have frequent excursions into the warmer ranges of their thermal window, and a low
585 thermal sensitivity for development may be required to prevent development from proceeding
586 too rapidly, which would leave little time for the animal to grow. As argued in Section IV.7, a
587 low thermal sensitivity of development rate appears to be associated with larger genomes, and
588 grasshoppers indeed have the largest genome among insects (Alfsnes, Leinaas & Hessen,
589 2017). The combination of a high thermal sensitivity for growth rate (at the rearing
590 temperatures employed) and a reduced thermal sensitivity for development rate will make a
591 positive T–S response more likely in grasshoppers and other heliotherms (e.g. lizards).

592 Grasshoppers may adaptively reverse the TSR for other reasons. First, sun-basking
593 grasshoppers will gain heat rapidly, but heat loss will be equally rapid as they are too small to
594 conserve heat in any significant amount. According to the heat balance model by Olalla-
595 Tárraga & Rodríguez (2007), it could be adaptive to be smaller in colder environments:
596 during the periods of sunshine they can then heat up more rapidly and spend less time in
597 absolute terms on heating up and more time on foraging.

598 Second, grasshoppers are commonly univoltine owing to an obligatory diapause in
599 their egg stage (Van Wingerden, Musters & Maaskamp, 1991), making it more profitable to
600 grow larger, as completing an additional generation may not be an option. Larger adult
601 grasshoppers produce proportionately larger egg pods, conferring a fitness advantage to
602 growing larger (Walters & Hassall, 2006). In warmer conditions, avoidance of excessive
603 developmental acceleration would leave sufficient time for the animal to grow to a large and
604 fecund body size (Berner & Blanckenhorn, 2006). In line with this reasoning, Buckley *et al.*
605 (2015) documented that grasshoppers inhabiting high elevations increased their development
606 time over the course of 50 years of climate warming.

607

608 **(7) Effects of cell and genome size on thermal responses**

609 Changes in body size mostly result from changes in either cell number, cell size, or a
610 combination of these (Calboli, Gilchrist & Partridge, 2003). As a result, thermal plasticity in
611 body size could reflect changes in cell size (Hessen, Daufresne & Leinaas, 2013; Table 1).
612 Clearly, changes in cell size mirror changes in body size in eutelic animals, whose number of
613 cells upon reaching adulthood is fixed (e.g. rotifers, most nematodes and some copepods; see

614 McLaren & Marcogliese, 1983; Ruppert, Fox & Barnes, 2004). However, also in non-eutelic
615 animals, changes in cell size can correlate strongly with T–S responses (Partridge *et al.*, 1994;
616 Van Voorhies, 1996; Arendt, 2007; Leinaas *et al.*, 2016; Hermaniuk, Rybacki & Taylor,
617 2016). Strikingly, while food availability generally affects cell number, temperature appears
618 to act mainly *via* changing cell size (Arendt, 2007; Czarnołęski *et al.*, 2013), although the
619 effects of food and temperature are not completely independent (Padmanabha *et al.*, 2011).
620 Thus T–S responses at the cellular level are also consistent with the contrasting effects of
621 rearing temperature and food conditions on whole-organism size (Berrigan & Charnov, 1994).

622 Across species or degrees of cell ploidy, cell size appears to be linked to the size of the
623 nucleus, which in turn is linked to genome size, although the causality and its direction are not
624 completely resolved (Gregory, 2001; Cavalier-Smith, 2005; Hessen *et al.*, 2013). Indeed,
625 artificially inducing triploidy in zebrafish (*Danio rerio*) resulted in a 50% increase in cell size,
626 resembling the 50% increase in genome size (van der Pol, Flik & Verberk, 2020). Studies
627 have found that plastic thermal responses in body size were accompanied by dynamic
628 adjustments in both cell size and nucleus size (by adjusting chromatin packaging) and thus
629 there is scope for cell size also to generate or parallel the TSR during ontogeny (Leinaas *et al.*,
630 2016; Hermaniuk *et al.*, 2016).

631 The consequences of cell size are temperature dependent (Szarski, 1983). Protein
632 synthesis rates are naturally slowed down in the cold. Boosted expression of key enzymes to
633 maintain adequate protein synthesis rates at the cellular level may be facilitated by a large
634 genome with multiple gene copies due to gene duplication, or by having uncondensed DNA
635 (Xia, 1995; Hessen *et al.*, 2013). Another potential advantage of larger cells in the cold is to
636 mitigate developmental noise. Finite numbers of molecules [proteins or messenger RNA
637 (mRNA)] introduce stochasticity in developmental pathways whose regulation arising from
638 random interactions of molecules becomes increasingly unpredictable and variable with
639 reduced absolute numbers of molecules (see Woods, 2014). Such stochasticity increases if the
640 number of molecules that participate in a reaction are lower or if the reaction rates are slower.
641 Thus, having larger cells with higher absolute numbers of molecules mitigates the effect of
642 slower reaction rates in the cold. Differences in cell size could also be mechanistically linked
643 to oxygen supply (Woods, 1999; Makarieva *et al.*, 2005; Atkinson *et al.*, 2006). Since
644 diffusion rates of oxygen are greater in lipids, membranes may act as preferential diffusion
645 pathways for oxygen (Subczynski, Hyde & Kusumi, 1989). Small cells also have more
646 surface area relative to volume, conferring a greater capacity for uptake of oxygen and other
647 resources. Finally, diffusion distances from the cell membrane to the mitochondria in the
648 cytosol are smaller in small cells. A reduced cell size in warm conditions may thus be part of
649 an adaptive response to improve oxygen provisioning and modulate growth and development.
650 Reductions in cell size might be achieved *via* oxygen sensing and activation of the HIF
651 (Hypoxia-Inducible Factor) and mTOR (mammalian Target Of Rapamycin) regulatory
652 pathways (e.g. Guzy & Schumacker, 2006). In the nematode *Caenorhabditis elegans*, a
653 mutation in a single gene appeared to control whether animals conformed to the TSR or not
654 (Kammenga *et al.*, 2007). The gene involved encoded a calpain-like protease, which has a
655 high homology with mammalian calpains known to regulate cell size and which can be
656 induced by hypoxia (Cui *et al.*, 2015). An oxygen perspective may therefore apply not just to
657 organism size, but also to a lower, cellular, level of biological organization, whereby a

658 reduced capacity for oxygen uptake may impact the energy budgets of larger cells (Atkinson
659 *et al.*, 2006; Table 1).

660 The strength and direction of T–S responses could also possibly be related to cell size.
661 If (temperature-induced) risks of oxygen limitation are more likely to arise in tissues made up
662 of large cells, animals with larger cells may be more likely to reduce cell size plastically to
663 improve oxygen provisioning. If variation in cell number is small, such changes in cell size
664 will be reflected in stronger T–S responses in body size. Triploid tadpoles of the frog
665 *Pelophylax esculentus* were indeed shown to exhibit a stronger T–S response upon
666 metamorphosis compared to diploid tadpoles (Hermaniuk *et al.*, 2016) and also a comparison
667 of fruit flies differing in genome size revealed stronger T–S responses in flies with larger
668 genomes (Ellis *et al.*, 2014). Wyngaard *et al.* (2005) reported differences in T–S responses
669 across five species of copepods whereby the strongest T–S responses were observed in the
670 species with the largest genome [*M. latipes*: ~ C-value (the amount, in picograms, of DNA
671 contained within a haploid nucleus) of 4 pg or 3.91×10^9 base pairs], and the weakest T–S
672 responses were observed in the species with the smallest genome (*T. crassus*: ~ 0.8 pg).
673 Horne *et al.* (2016) report differences in T–S responses in which copepods of the order
674 Calanoida (~4.5 pg) showed a stronger TSR than those of the order Cyclopoida (~ 1 pg).
675 These orders exhibit a significant difference in genome size (and hence probably cell size) [*t*-
676 test: $P = 1.005 \times 10^{-7}$; calculated from data in Gregory (2018); see also Wyngaard & Rasch,
677 2000]. Similarly, aquatic species living in cold environments tend to have larger genomes
678 (Dufresne & Jeffery, 2011; Lorch *et al.*, 2016; Alfsnes *et al.*, 2017; Jeffery, Yampolsky &
679 Gregory, 2017), suggesting a cell-size parallel with Bergmann’s rule, at least for aquatic
680 animals. In general, animals increase body size mainly through cell proliferation during early
681 development, but by cell growth in later life (Kammenga *et al.*, 2007; Czarnołęski *et al.*,
682 2008, but see Aguilar-Alberola & Mesquita-Joanes, 2014; Horne *et al.*, 2019) and this fits
683 with the TSR being less pronounced for egg size, manifesting itself in later life stages (Forster
684 *et al.*, 2011a; Forster & Hirst, 2012). In summary, there are clear patterns between genome
685 size and the strength of the T–S response, with stronger T–S responses being found in animals
686 with larger cell sizes. Such patterns suggest a link between cell size and the strength of the
687 TSR.

688 Genome size is also linked to development rate, with large genomes being associated
689 with slower development in fruit flies (e.g. Gregory & Johnston, 2008), copepods (McLaren,
690 Sevigny & Corkett, 1988) and anurans (van der Have, 2008). Genome size and development
691 rate are mechanistically linked as DNA replication takes proportionally longer with larger
692 genomes (Van ‘t Hof & Sparrow, 1963), although slow replication of large genomes can be
693 compensated for by increased ribosomal DNA (rDNA) copy number (Prokopowich, Gregory
694 & Crease, 2003; White & McLaren, 2000). These genomic effects on development rates may
695 be temperature specific (e.g. Ellis *et al.*, 2014), and a lower thermal sensitivity of
696 development has been reported in copepod species with a larger genome (Wyngaard *et al.*,
697 2005), and in both triploid froglets (Hermaniuk *et al.*, 2016) and polyploid cladocerans (Van
698 Geest *et al.*, 2010; Dufresne & Hebert, 1998) when compared to their diploid counterparts.
699 Whereas a lower thermal sensitivity of development may be beneficial in certain
700 environments (see Section IV.6), several copepods exhibit chromatin diminution during early
701 embryogenesis, possibly as a way to increase development rate by removing the burden of

702 lengthy replication cycles from large genomes. Chromatin diminution results in substantial
703 decreases in nuclear DNA content of the somatic cells (Wyngaard & Rasch, 2000) due to
704 chromosomal fragmentation and excision of large portions of DNA in the presomatic line.
705 Such diminution has also been reported in other eukaryotes (Parfrey, Lahr & Katz, 2008). In
706 summary, there is evidence that a large genome size reduces development rates and possibly
707 also their thermal sensitivity.

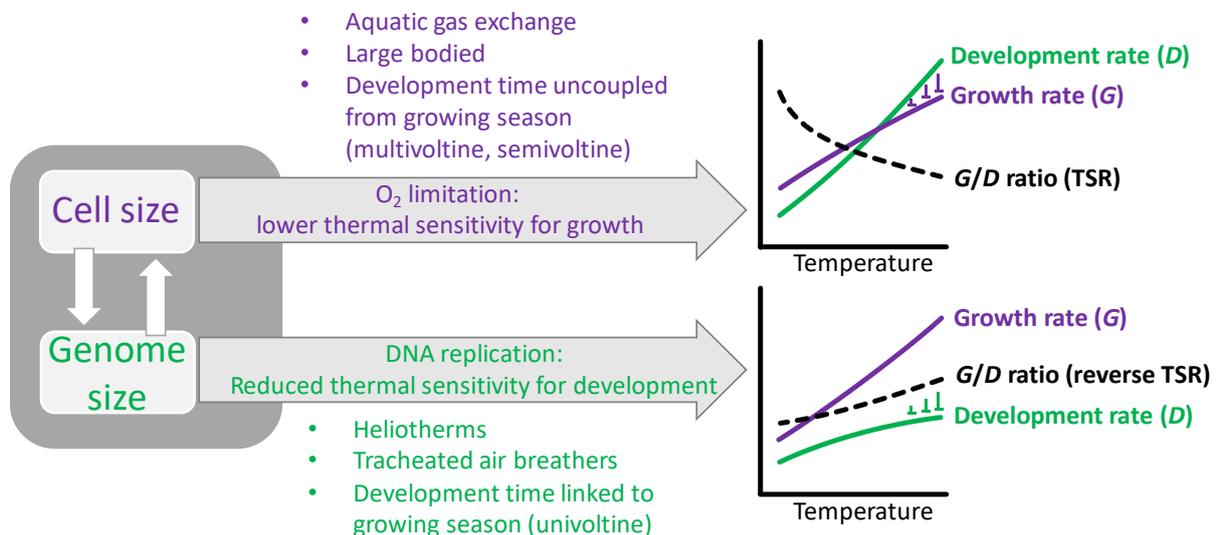
708 Walczyńska *et al.* (2015a,b), building on the work of Stelzer (2002), showed that in
709 eutelic rotifers, thermal responses in cell size (and thus body size) followed the TSR. In
710 addition to the absolute temperature, the direction in which temperature changed mattered.
711 When reared at a common temperature of 20 °C, differences in cell size depended on the
712 temperatures the mothers had previously experienced during egg development: offspring from
713 mothers that had experienced warmer conditions attained a larger cell size, compared to
714 offspring from mothers that had experienced cooler conditions. Egg size in fruit flies (Crill,
715 Huey & Gilchrist, 1996) and butterflies (Fisher *et al.*, 2003) was also found to vary in
716 response to the temperature that parents experienced. The importance of parental temperature
717 and the direction of temperature change suggest that parents convey information to their
718 offspring on when to arrest cell growth. Thus, it is unlikely that oxygen or another resource
719 sets absolute or insurmountable limits to the size that a cell can attain (see also Section IV.2);
720 cells will not keep growing to the point at which they will become energy limited due to
721 insufficient resource provisioning. Since conditions experienced by the parents can reasonably
722 be anticipated to resemble the conditions that the offspring will face as well, providing such
723 information may be adaptive. If offspring developed at temperatures cooler than those
724 experienced by the parents, offspring had larger cells and *vice versa* when temperatures are
725 warmer than parental temperatures (Walczyńska *et al.*, 2015b). Documented responses in
726 body size to temperature and oxygen combinations coincided with higher fecundity,
727 suggesting that they are adaptive (Walczyńska *et al.*, 2015a). Thus, the information bestowed
728 upon the offspring may arrest cell growth when the ratio between oxygen supply and demand
729 falls below a certain threshold. The ratio between oxygen supply and demand is reduced both
730 by an increase in cell size (by reducing oxygen supply) and an increase in temperature (by
731 increasing oxygen demand). Such a threshold ratio may safeguard sufficient oxygen
732 provisioning under warmer conditions (Walczyńska *et al.*, 2015a,b). A similar threshold may
733 govern the critical size in insect development; critical size decreases both under warming
734 (Ghosh, Testa & Shingleton, 2013) and hypoxia (Callier *et al.*, 2013), while there is also
735 evidence for increased critical size under hyperoxia (Kivelä *et al.*, 2018). Also, when medaka
736 fish (*Oryzias latipes*) were reared for multiple generations, the smallest size was observed in
737 fish reared under temperatures that were warmer than they had previously experienced (Loisel
738 *et al.*, 2019). A tentative conclusion from the studies reviewed above is that oxygen limitation
739 may take the form of an ultimate driver, whereby animals have evolved plastic, canalized
740 responses geared to avoid oxygen limitation, limiting cell growth to a point with sufficient
741 capacity for oxygen provisioning, a threshold calibrated against the temperatures experienced
742 by adults [see also Harrison *et al.* (2018) and Section VI.3].

743

744

745 **(8) Explaining the TSR as the balance between growth and development**

746 A low thermal sensitivity of development rate relative to growth rate weakens (or reverses)
 747 the TSR, while the TSR is strengthened if development rates increase with temperature more
 748 than do growth rates. We have seen why warming-induced acceleration is more likely
 749 curtailed for growth: although limited oxygen or food is unlikely to constrain growth rates
 750 directly when provided *ad libitum*, growth rates could be adaptively modulated as a result of
 751 warming exacerbating resource limitation during the species' evolutionary history (see
 752 Section IV.2). Risks of oxygen limitation are more likely in ectotherms that rely on
 753 underwater gas exchange, but are less likely in air-breathing, tracheated arthropods that
 754 employ convective ventilation. Since the TSR is expressed at the level of the whole organism
 755 (size at maturity, asymptotic size), it integrates the effects that strengthen or weaken the TSR
 756 at each level of biological organization. A large genome can either weaken the TSR if it
 757 predominantly decelerates developmental rate, or strengthen the TSR when the concomitant
 758 larger cell size results in oxygen limitation effects on growth (Fig 4). With oxygen limitation
 759



760
 761

762 **Fig. 4.** Overview of influences on growth rate (G) and development rate (D) responses to
 763 temperature, and hence their ratio and the temperature–size response. TSR,
 764 temperature–size rule. Temperature stimulates both growth rate and development rate,
 765 but the relative increase may be modulated by effects of cell size, genome size, body
 766 size, life cycle, thermoregulatory behaviour and mode of respiration. Oxygen limitation is
 767 more likely in large aquatic ectotherms with large cells, and could constrain the
 768 stimulating effects of temperature on growth rate. Consequently, animal development
 769 outpaces growth under warmer conditions, resulting in a decrease in body size (purple
 770 pathway). A large genome size may be associated with a lower thermal sensitivity of
 771 development. Consequently, development does not outpace growth under warmer
 772 conditions and the faster growth results in larger body sizes (green pathway). Due to the
 773 strong linkage between genome size and cell size, both mechanisms will operate in
 774 tandem, but the relative importance of these mechanisms may differ among animals,
 775 depending on their characteristics.

776

777 likely having less of an influence in air-breathers, the effect of genome size on development
 778 rate could be dominant here. The slowing down of development rates with increases in
 779 genome size could explain why latitudinal clines of genome size are predominantly negative
 780 for (terrestrial) insects (see Alfsnes *et al.*, 2017): at higher latitudes the shorter seasons would
 781 require smaller genomes to enable more rapid development. By contrast, for aquatic
 782 arthropods cell-size effects on oxygen limitation may be more important under warm
 783 conditions. This could help explain the divergent T–S response between these two groups.

784

785 **V. WHY ARE NEGATIVE T–S RESPONSES SO PREVALENT?**

786 Assuming that the TSR is adaptive, resources are allocated in a temperature- and size-
 787 dependent way to enhance fitness. Compared to cool conditions, warmer conditions are
 788 associated with faster growth, increased mortality and faster maturity at a smaller size,
 789 mirroring major patterns in life histories of animals (Pianka, 1970). The trade-off between
 790 adult size and development time (Abrams *et al.*, 1996) places animals on a continuum from
 791 early maturation at a small size (allocating resources preferentially to development,
 792 prioritizing time), to maturing later at a large size (allocating resources preferentially to
 793 growth, prioritizing size). Although there are clear benefits to both being large and being fast,
 794 we here focus on how warm temperatures may tip the balance in favour of growing faster to a
 795 smaller size and *vice versa* (Table 1).

796

797 **(1) Mortality and reproduction are temperature- and size-dependent**

798 Prioritizing time may be favoured in warm conditions, as warming may reduce
 799 lifespan *via* increased competition, predation or resource scarcity (food or oxygen). Such
 800 increased mortality risks may be associated with thermal acceleration of physiological rates
 801 (e.g. growth, development and reproduction), whose resultant energetic costs are known to
 802 impair immune function (De Block & Stoks, 2008) and reduce lifespan (Lee, Monaghan &
 803 Metcalfe, 2013; Lind *et al.*, 2017), likely *via* oxidative stress and cellular senescence
 804 (Hemmer-Brepson *et al.*, 2014). Increased mortality typically favours adaptively reducing the
 805 duration of the life stage at increased risk. In addition, if warm conditions during juvenile
 806 growth incur costs that increase the risk of (reproductive) senescence or reduced lifespan, it
 807 should pay to reproduce sooner (with consequently smaller size) to reduce these risks, which
 808 thus can provide an adaptive explanation for the TSR at maturity (Sibly & Atkinson, 1994;
 809 Kindlman *et al.*, 2001; Kozłowski *et al.*, 2004). Thermal effects on survivorship reported from
 810 laboratory studies do not generally constitute a sufficiently strong selection pressure to
 811 account fully for the TSR (Myers & Runge, 1983; Angilletta, Steury & Sears, 2004).
 812 However, the TSR may still be explained adaptively from increased mortality at higher
 813 temperatures if thermal effects in the field are larger than those reported from laboratory
 814 studies, for example when higher temperature increases predator-induced mortality (Hirst &
 815 Kiorboe, 2002). Increased mortality risks in the warm may also be size dependent (e.g. related
 816 to predator escape). If mortality increases with warming especially in larger individuals as has
 817 been found for *Daphnia magna* (Bruijning, ten Berge & Jongjans, 2018), it could be
 818 beneficial to mature at a smaller size. Leiva, Calosi & Verberk (2019) also found survival of
 819 heat stress to be dependent on body size.

820 Prioritizing size may be favoured in cold conditions because of gains in fecundity.
821 Larger mothers typically produce more offspring. In fish, fecundity, egg size and egg energy
822 content all increased with body size, such that larger mothers had disproportionately higher
823 reproductive energy output (Barneche *et al.*, 2018). Such gains in fecundity in larger
824 individuals were magnified under cold conditions in freshwater snails of the genus *Physa*
825 (Arendt, 2015) and in *Daphnia* cladocerans (Weetman & Atkinson, 2004), but not in the
826 water strider *Aquarius remiges* (Arendt & Fairbairn, 2012). In summary, temperature may
827 evolve as a cue such that warm conditions accelerate juvenile development rate, because of a
828 predictable association between warm conditions and increased mortality risks in the field
829 during a species' evolutionary past or because the size–fecundity relationship changes with
830 temperature.

831

832 **(2) Resource limitation is temperature- and size-dependent**

833 At higher temperatures, ectotherms require more resources to fuel their enhanced
834 activity rates. Although this increased demand may not constrain growth (see Section IV.2) it
835 could increase the risk that resources (e.g. oxygen or food) become limiting under resource-
836 demanding conditions. Therefore, negative T–S responses could have evolved to avoid
837 resource limitation. This hypothesis can be considered the selective effect of 'resource-
838 limitation past' (see Section VI.3 for this principle applied to oxygen). Larger individuals
839 have a higher *per capita* resource demand and both their aerobic scope and their ability to
840 obtain sufficient food or oxygen may be less capable of matching warming-enhanced demand
841 compared with that of smaller individuals (Atkinson *et al.*, 2006; Neubauer & Andersen,
842 2019; Rubalcaba, J.G., Verberk, W.C.E.P., Hendriks, A.J., Saris, B. & Woods, H.A., in
843 preparation). Although larger individuals can have greater tolerance to lack of food
844 (Cushman, Lawton & Manly, 1993; Arnett & Gotelli, 2003; Scharf, Galkin & Halle, 2015),
845 starvation is more likely outside the growing season (i.e. winter). Moreover, studies
846 demonstrating that animals evolve larger body sizes when reared under cool conditions and
847 fed *ad libitum* (e.g. Partridge *et al.*, 1994) indicate that starvation is not a necessary condition
848 for the evolution of the observed T–S reaction norms. In summary, temperature may evolve as
849 a cue such that cool conditions favour growing to a larger size, because of a predictable
850 association from the species' evolutionary past, between cool conditions and lower risks of
851 resources becoming limiting.

852

853 **(3) Mortality and fecundity are often resource dependent**

854 Evolutionary effects of mortality, fecundity (see Section V.1), and resource limitation (see
855 Section V.2) are not independent: resource shortage can impair fecundity and increase
856 mortality risks. Hence, seemingly disparate causes of the TSR can be unified. Specifically,
857 although some warming-enhanced mortality risks (e.g. predation, pond drying) may kill
858 irrespective of resource availability to the organism, it is clear that mortality and resource
859 limitation (related to an organism's aerobic scope, feeding capacity or energy reserves) are
860 often interdependent. On the one hand, increased resource limitation may limit the ability to
861 mount a sufficient defence against threats such as predators or drought, and hence lead to
862 increased mortality from these threats. This increased mortality could favour the evolution of
863 a reaction norm that accelerates juvenile development (maturation) in the warm, thereby

864 reducing this resource-dependent mortality (see Section V.1). On the other hand, selection to
 865 actively avoid increased predation risk or unfavourable environmental conditions in the warm
 866 may increase selection for a greater resource-supply safety margin, thereby favouring the
 867 evolution of a thermal reaction norm producing individuals with a larger safety margin (e.g.
 868 aerobic scope), which may be achieved by growing to a smaller size (see Section IV.2). In
 869 both cases, adults would be smaller, and would mature earlier in the warm. Thermal effects on
 870 fecundity are also likely to be highly dependent on the resource-supply safety margin for
 871 adults of different sizes. For instance, in the (aquatic) rotifer *Lecane inermis*, gains in
 872 fecundity with body size were dependent not only on temperature, but also on oxygen levels
 873 (Walczyńska *et al.*, 2015a), and in mosquitofish (*Gambusia affinis*) populations, the
 874 reproductive advantage of larger body size decreased with increasing site temperature within
 875 100 years (Fryxell *et al.*, 2020).

876 In summary, the hypotheses discussed in *Sections V(1) and (2)* are not independent.
 877 Indeed, to explain the TSR, it is not necessary for resource limitation, mortality and fecundity
 878 all to be size- and temperature-dependent. If one of these is, and their effects are amplified by
 879 any of the others, a TSR response can evolve from a variety of mechanisms, potentially
 880 explaining why it is so widespread among ectotherms.

881

882 **VI. WHAT EXPLAINS MOST OF THE VARIATION IN T–S RESPONSES?**

883 Although adult body size is usually reduced under warmer rearing conditions, understanding
 884 the causes of variation in the magnitude and direction of T–S responses is likely to lead to a
 885 more complete explanation of why body size changes with temperature. The most striking
 886 differences to be explained occur between aquatic and terrestrial species (Forster *et al.*, 2012;
 887 Horne *et al.*, 2015). Here we focus on the greater thermal seasonality in terrestrial
 888 environments, stronger selection to avoid oxygen limitation in aquatic environments, and
 889 temperature being a more reliable cue in aquatic environments.

890

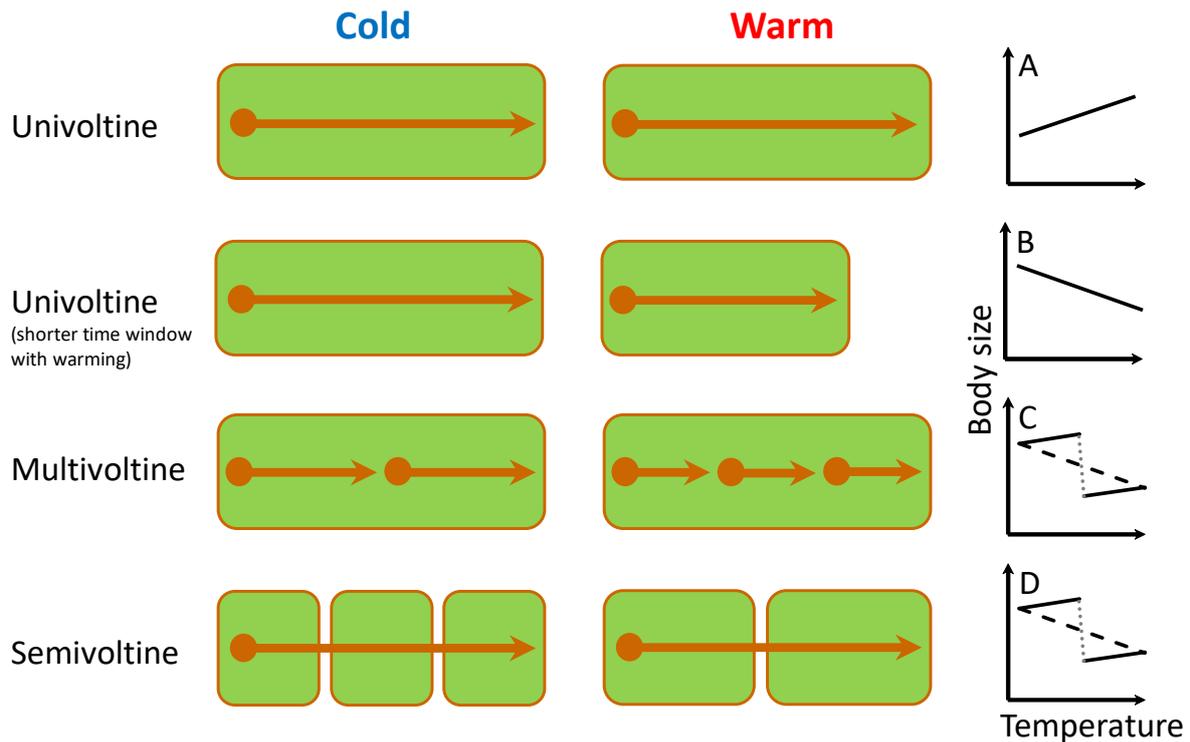
891 **(1) Seasonality and voltinism**

892 Maturing earlier may become feasible under warmer conditions (e.g. due to enhanced
 893 growth and food consumption). The benefits of early breeding or even completion of an extra
 894 generation may be substantial (Cole, 1954), and can outweigh the disadvantages associated
 895 with a smaller body size. These benefits of early maturation are greater in growing
 896 populations, since the earlier an organism breeds the greater the proportional contribution
 897 each offspring makes to the population. This increase in parental fitness is analogous to the
 898 increased financial gains from investing money early in a bank account that gives high
 899 compound interest (Calow, 1981; Kozłowski, 1992; Atkinson, 1994; Fisher & Fiedler, 2002;
 900 Kingsolver & Huey, 2008). Although population growth rate is variable and not consistently
 901 linked to temperature, in seasonal habitats, the number of generations in a year may influence
 902 T–S responses. Univoltine species, completing a single generation every year, do not have the
 903 opportunity to increase fitness by speeding up development to increase the number of
 904 generations per year, especially when univoltinism is enforced by an obligatory diapause.
 905 Instead, obligate univoltine species should use all the time available to them, and their faster
 906 growth in warm conditions therefore will weaken the TSR or even give rise to a converse
 907 TSR (Fig. 5; overcompensation *sensu* Blanckenhorn & Demont, 2004). Indeed, Fisher &

908 Fiedler (2002) demonstrated a weaker TSR in univoltine populations of the butterfly *Lycaena*
909 *hippotoe* than in multivoltine populations. Similarly, perceived time available can alter T–S
910 responses (see Section VI.2). Moreover, Sniegula, Golab & Johansson (2016) reported a
911 smaller size at maturity in cold-latitude populations of the obligate univoltine damselfly
912 *Lestes sponsa* (i.e. a converse James' cline). Grasshoppers have a diapause at the egg stage,
913 enforcing univoltinism, which may explain why grasshoppers generally exhibit converse T–S
914 responses (see Section IV.6). Some univoltine species may be obligatorily univoltine not
915 because of a limited duration of the warm season *per se*, but because host plants are available
916 for a specific period only, such as is the case for the butterfly *Anthocharis cardamines*
917 (Posledovich *et al.*, 2014). When host plants are available for a shorter period due to warming,
918 this may still result in faster development to a smaller size despite being univoltine (Fig. 5).
919 Semivoltine species, like multivoltine species, can decrease the time spent per generation (in
920 their case by decreasing the number of years per generation): benefits from increasing
921 numbers of generations per unit time can therefore accrue to species from both groups, but not
922 to univoltine species (Fig. 5). In summary, options to accrue fitness by speeding up
923 development are limited in univoltine species and these are therefore less likely to adhere to
924 the TSR.

925 Seasonality interacts with voltinism as strong end-of season constraints will result in
926 high mortality, selecting against semivoltine species taking multiple years to complete a
927 generation (Ejsmond *et al.*, 2018; Walczyńska, Dańko & Kozłowski, 2010). Chown & Klok
928 (2003) found clear differences in altitudinal clines in body size between two regions that
929 differed substantially in seasonality. Species in the region with strong end-of season
930 constraints displayed discrete generations and converse size clines (larger individuals at
931 lower, warmer altitudes), whereas those in the region without clear seasonality displayed
932 overlapping generations and achieved the largest body size in colder, higher altitudes.
933 Minards *et al.* (2014) also report differences between low- and high-altitude populations of
934 *Hemideina* orthopterans in their T–S responses established by rearing animals in the
935 laboratory: low-altitude populations followed the TSR and high-altitude populations showed a
936 converse T–S response, suggesting that thermal responses may be adaptively shaped by
937 selection due to differences in season length. The importance of end-of-season constraints and
938 voltinism is also suggested in a study of community-level body-size gradients by Zeuss *et al.*
939 (2017): aquatic odonates showed Bergmann clines, whereas terrestrial lepidopterans showed
940 converse Bergmann clines. The different responses of lepidopterans and odonates could at
941 least partly be explained *via* effects on voltinism: many odonate species extend juvenile
942 development over multiple years, whereas most butterflies were univoltine and hence more
943 likely to have seasonal time limitation on body size, which would be relaxed at lower latitudes
944 with longer annual growth periods. In summary, the risks of not completing juvenile
945 development in time before the onset of winter has much more severe consequences in
946 terrestrial environments than in aquatic environments (e.g. Van Dyck *et al.*, 2015; Forrest,
947 Regan Cross & CaraDonna, 2019): this may explain why time constraints did not generally
948 reverse T–S responses of aquatic ectotherms (Cabanita & Atkinson, 2006), and may
949 contribute to the difference between aquatic and terrestrial T–S responses (Forster *et al.*,
950 2012; Horne *et al.*, 2015; Rollinson & Rowe, 2018).

951



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954 **Fig. 5.** Schematic overview of different temperature-size (T-S) responses in relation to
 955 voltinism. T-S responses may depend on the interaction between the length of the
 956 growing season (green box) and the development time (brown arrow), especially in
 957 (terrestrial) organisms living in habitats with strong end-of season constraints. For
 958 univoltine species, warming may allow animals to grow faster during their (fixed)
 959 development time, resulting in animals reaching a larger size (A), unless time for
 960 development is also reduced under warmer conditions (B). Warming may also allow
 961 animals to fit more generations into a certain amount of time, either by increasing the
 962 number of generations (C; multivoltine species) or by decreasing the number of years
 963 needed for completion (D; semivoltine species). A faster development can result in
 964 animals growing to a smaller size under warmer conditions when viewed across the
 965 whole thermal gradient (dashed black line). However, shifts in voltinism, may result in a
 966 sawtooth pattern, with animals growing to a larger size with warming (solid black line),
 967 until there is an increase in voltinism at which point animals reach a smaller size (due to
 968 less time available for growth in a given generation, dotted grey line). Such shifts in
 969 voltinism and the resulting sawtooth patterns are most readily seen in latitudinal clines.
 970

971 (2) Effects of day length on T-S responses

972 To understand seasonality effects in the field better, laboratory experiments have shown how
 973 daylength modulates the effects of temperature on size, changing the allocation of resources
 974 to growth, development, reproduction and maintenance (lifespan) (Ernsting & Isaaks, 2000;
 975 Camus & Zeng, 2008). Consequently, day length may affect the strength of the TSR in both
 976 terrestrial and aquatic taxa (e.g. Kutcherov *et al.*, 2011; Martínez-Jerónimo, 2012; Kollberg *et al.*,
 977 2013; De Block & Stoks, 2003; but see Cabanita & Atkinson, 2006). In multivoltine
 978 tephritid flies, warming results in early maturation at a smaller size under an early-season
 979 photoperiod, whereas maturation was delayed under a late-season photoperiod, resulting in

980 larger adults with presumably better chances of surviving winter conditions (Xi *et al.*, 2016).
981 Such modulation of T–S responses and of juvenile development period by photoperiod
982 suggests that these animals use day length as a cue to gauge time in the year, and hence the
983 availability of future favourable conditions. Animals tend to speed up development when less
984 time is available, but they can also increase growth rates, which can buffer changes in adult
985 size (Abrams *et al.*, 1996; Blanckenhorn & Demont, 2004; Kivelä *et al.*, 2011; Buckley *et al.*,
986 2015). This complexity between voltinism, body size and temperature can make it difficult to
987 disentangle the different influences (Cabanita & Atkinson, 2006) and males and females may
988 prioritize development time and body size differently (De Block & Stoks, 2003). What is
989 clear though is that cold, high-latitude environments present stronger time limitations, which
990 may be overcome by extending development over multiple years (decreasing voltinism).
991 Changes in voltinism across populations may be accompanied by changes in allele frequency
992 of clock genes linked to post-diapause development time such that complete life cycles can be
993 ‘fitted’ into latitudinally varying growing seasons (Levy *et al.*, 2015). Shifts in voltinism
994 across thermal clines can result in changes in strength of seasonal time constraints on the
995 populations. Consequently, thermal responses in body size along thermal clines (e.g.
996 associated with altitude and latitude) can be discontinuous, following a saw-tooth pattern,
997 which arises from alternate intensification and relaxation of time constraints as both voltinism
998 and season length vary (Roff, 1980). Within species of aquatic arthropods, gradients of body
999 size across latitudes are non-linear, indicating that there is more to latitudinal clines in body
1000 size than just temperature (Johansson, 2003; Hassall, 2013). In summary, day length may alter
1001 the thermal reaction norms and the resulting size and age at maturity in ways that depend on:
1002 (i) the voltinism of the population from which the individuals originated; (ii) the physiological
1003 state of the individual; and (iii) the temperature and light regime of its environment (Honěk,
1004 1996; Gotthard, Nylin & Wiklund, 2000; Lopatina *et al.*, 2011; Martínez-Jerónimo, 2012;
1005 Clemmensen & Hahn, 2015). All of these provide information about availability of time for
1006 development (Roff, 1980; Lee, Monaghan & Metcalfe, 2010).

1007

1008 **(3) The ‘Ghost of Oxygen-limitation Past’**

1009 The proximate mechanisms by which a shortage of oxygen puts large individuals in the warm
1010 at a disadvantage, especially in aquatic environments (see Section IV.3) can also act on
1011 evolutionary timescales to eliminate phenotypes that produce large adults under warm
1012 conditions. Importantly, powerful selective events in the past need not be frequent to affect
1013 the phenotypes of descendants (Grant *et al.*, 2017). According to this evolutionary
1014 perspective, at warmer temperatures oxygen may be limiting only infrequently, even for
1015 aquatic species, because past selection on T–S responses has eliminated genotypes more
1016 prone to oxygen limitation. This selection could favour genotypes with enhanced supply
1017 capacity, and indeed, arthropods have evolved a suite of plastic responses geared to avoid
1018 oxygen limitation (see Harrison *et al.*, 2018). For aquatic ectotherms, selection may also have
1019 favoured reaction norms that increase body size in cool conditions, as being large may be
1020 helpful in overcoming the viscosity of cold water: larger animals can generate higher flow
1021 speeds of water, which increases the energy efficiency of gill surface irrigation and of body
1022 propulsion (Verberk & Atkinson, 2013). Experiments that investigate T–S responses typically
1023 include normoxic laboratory settings with abundant food and without natural enemies. Under

1024 these favourable conditions, individuals would be unlikely to experience resource limitation.
1025 Instead, these individuals would have evolved a canalized growth response that safeguards
1026 sufficient oxygen provisioning (e.g. a safety margin, such as aerobic scope) under warmer
1027 conditions. In the same way that Connell's 'Ghost of Competition Past' could explain how
1028 avoidance of competition had evolved (Connell, 1980), we can invoke a 'Ghost of Oxygen-
1029 limitation Past' potentially to explain the evolution of a strong TSR in aquatic species, even if
1030 oxygen limitation is not evident under favourable conditions. This idea can explain why
1031 responses to hyperoxia are usually small (Frazier *et al.*, 2001): the 'Ghost of Oxygen-
1032 limitation Past' may have selected against animals that would grow bigger in the presence of
1033 additional oxygen if their ancestors did not benefit from an increased body size at hyperoxia.
1034 Indeed, fruit fly body-size responses to varying levels of oxygen do evolve (Henry &
1035 Harrison, 2004). T-S responses also evolve as shown by multi-generation experiments on
1036 medaka fish reared under different temperatures (Loisel *et al.*, 2019). Fish reared under warm
1037 conditions grew to a smaller asymptotic size compared with those reared under cool
1038 conditions. However, fish reared under warm conditions for a single generation grew smaller
1039 but also produced fewer offspring than those reared for multiple generations under these
1040 conditions (Loisel *et al.*, 2019). Thus, evolution can modulate growth trajectories. If the
1041 juvenile growth temperatures carry information on whether resource limitation is likely to
1042 constrain fitness once these juveniles mature, thermal reaction norms for size at maturity
1043 could reflect the balance of oxygen demand to supply under T-S combinations experienced
1044 by ancestors (Atkinson & Sibly, 1997a; Atkinson *et al.*, 2006). In this way, temperatures
1045 experienced early in ontogeny may act as a cue to adjust growth trajectories and the resulting
1046 adult or final size. Such cues may also be experienced by the mother, and this information can
1047 then be passed on to the offspring as has been argued for adaptive changes in cell size [see
1048 Walczyńska *et al.* (2015b) and Section IV.7).

1049

1050 **(4) Temperature and oxygen as an information cue**

1051 The 'Ghost of Oxygen-limitation Past' views T-S responses as adaptive responses to
1052 maintain a safety margin for aerobic scope (Fig. 3) that have evolved in response to
1053 temperature and oxygen conditions experienced by ancestors. These reaction norms
1054 incorporate temperature and oxygen levels (or physiological correlates of oxygen levels) as
1055 cues and the information content of such cues is likely to differ between terrestrial and aquatic
1056 environments. In aquatic environments, temperature fluctuates less and varies more
1057 predictably than on land, especially for larger bodies of water, making water temperature a
1058 reliable cue. Moreover, terrestrial animals can exploit the greater thermal heterogeneity on
1059 land *via* behavioural thermoregulation, reducing the information content of air temperature.
1060 Hypoxia is also more common in aquatic environments, especially during warm periods. The
1061 'Ghost of Oxygen-limitation Past' is therefore more likely to operate in aquatic ectotherms,
1062 favouring genotypes that grow to a smaller size in warm water, and consequently avoiding
1063 oxygen constraints on growth. This prediction is consistent with the stronger TSR in aquatic
1064 than in terrestrial species (Fig. 2; Forster *et al.*, 2012; Horne *et al.*, 2015). In summary, on
1065 land, temperature is spatially more heterogeneous and temporally more variable. Coupled to
1066 the ability to thermoregulate, this may weaken any selection pressure on thermal reaction
1067 norms for size and age at maturity in terrestrial species.

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VII. QUESTIONS AND DIRECTIONS FOR FUTURE RESEARCH

We are beginning to understand the distinct patterns in T–S responses across animal groups. Still, many important questions remain, and here we propose some questions and directions for future research:

(1) Elucidate the role of cell size and genome size in generating the TSR. Does the thermal dependency of growth and development differ with cell size or genome size in both terrestrial and aquatic ectotherms? How do cell proliferation and cellular enlargement contribute to whole-organism growth during ontogeny, and do T–S responses become stronger during periods where cellular enlargement contributes most? Is the decision to arrest cell growth governed by a threshold ratio between supply and demand of oxygen to safeguard sufficient oxygen delivery, which is calibrated against the temperatures experienced by parents? How does this threshold ratio relate to the critical size of an insect?

(2) Elucidate the role of ecological factors in shaping T–S responses. How do T–S responses change with photoperiod, voltinism, predator characteristics (endotherm, ectotherm, sit-and-wait, active hunter, etc.) and food conditions? How do T–S responses affect community assembly by affecting geographic range shifts, predator–prey relationships and phenology?

(3) Extend the scope of data on T–S responses. How does developmental temperature affect body size in larger-bodied species, both aquatic and terrestrial? How does size in aquatic species change across altitudinal clines? How do T–S responses change over multiple generations when long-term adaptation also starts to play a role? Do multiple-generation T–S responses differ if temperature is decoupled from proposed selective factors (e.g. oxygen supply, time constraints, mortality schedules)?

(4) Create a database for T–S responses, facilitating future (meta) analyses.

(5) Explain the adaptive nature of T–S responses by integrating growth trajectories of juveniles to fitness consequences of adults (e.g. by modelling of energy budgets or conducting multigenerational studies into the evolution of T–S reaction norms, where putative selective agents (e.g. oxygen limitation, mortality risks) are decoupled from temperature. Are larger individuals more susceptible to oxygen limitation under warmer conditions and does such (incipient) oxygen limitation proximately or ultimately limit growth?

VIII. CONCLUSIONS

(1) The many explanations proposed (Table 1) for observed phenotypically plastic body-size responses to temperature (the TSR) differ in domain (focussing on physiological mechanisms that bring about T–S responses or their adaptive nature) and apply to different biological levels of organization (activity rates of enzymes, cells, organisms, populations, communities). The TSR is not universal, but the strength of the TSR varies in predictable ways (being stronger in larger, aquatic ectotherms and being weaker or reversed in larger terrestrial ectotherms).

(2) Effects of temperature are pervasive, affecting biological levels of organization ranging from whole-organism growth performance down to activities of individual proteins, which makes it unlikely that a single proximate mechanism underlies the TSR. An oxygen perspective may help to explain the effects of temperature on size, especially in large aquatic ectotherms, which are arguably most susceptible to risks of oxygen limitation.

1112 (3) Warming may exacerbate risks of oxygen limitation or reduce the safety margin of aerobic
 1113 scope. For air breathers such as terrestrial insects, problems with insufficient oxygen may be
 1114 less likely and time constraints take centre stage. Season length may constrain developmental
 1115 period, forcing them to prioritize time over size as overwintering is more challenging. Day
 1116 length and temperature may together provide information on how long conditions will remain
 1117 favourable for development, explaining why thermal responses are modulated by photoperiod.
 1118 (4) Time constraints, mortality risks, and resource limitation are not mutually exclusive
 1119 explanations for the TSR. Rather, they may operate in tandem but their relative importance
 1120 may vary depending on the ecology and physiology of the species in question (Fig. 4). At the
 1121 level of cells, effects of cell size on oxygen provisioning may be more relevant for aquatic
 1122 species, whereas effects of genome size on development time may be more relevant for
 1123 terrestrial species. Similarly, at the level of the whole organism, capacity for oxygen
 1124 provisioning differs with mode of respiration and habitat use, while end-of-season constraints
 1125 likely differ between aquatic and terrestrial species. Thus, multiple pathways operating at
 1126 different levels of organization show T–S responses that broadly differ across the aquatic–
 1127 terrestrial divide. T–S responses may be viewed as being canalized – producing the same
 1128 adaptive response by a range of mechanisms, since the resultant response has proven its
 1129 adaptive worth both for safeguarding energy status (e.g. *via* oxygen provisioning) and for
 1130 safeguarding completion of development (*via* time sensing). As such, oxygen supply can be
 1131 both a proximate mechanism and an ultimate driver (the ‘Ghost of Oxygen-limitation Past’).
 1132

1133 IX. ACKNOWLEDGEMENTS

1134 We thank Asta Audzijonyte and Aleksandra Walczyńska for providing thoughtful and
 1135 constructive feedback, which was very helpful in improving our manuscript. W.C.E.P.V.
 1136 gratefully acknowledges support from the Netherlands Organization for Scientific Research
 1137 (NWO-VIDI Grant 016.161.321). D.A., A.G.H. and C.R.H. were supported by NERC grant
 1138 NE/P012183/2.
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