Evidence of exposure to SARS-CoV-2 in cats and dogs from households in Italy

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Summary: SARS-CoV-2 emerged from animals and is now easily transmitted between people. Sporadic detection of natural cases in animals alongside successful experimental infections of pets, such as cats, ferrets and dogs, raises questions about the susceptibility of animals under natural conditions of pet ownership. Here we report a large-scale study to assess SARS-CoV-2 infection in 919 companion animals living in northern Italy, sampled at a time of frequent human infection. No animals tested PCR positive. However, 3.3% of dogs and 5.8% of cats had measurable SARS-CoV-2 neutralizing antibody titers, with dogs from COVID-19 positive households being significantly more likely to test positive than those from COVID-19 negative households. Understanding risk factors associated with this and their potential to infect other species requires urgent investigation.

**Introduction**

Cases of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection were detected in late December 2019 in Wuhan, Hubei province, China1, possibly as a spillover from bats to humans2, and rapidly spread worldwide becoming a pandemic3. Although the virus is believed to spread almost exclusively by human-to-human transmission, there are concerns that some animal species may contribute to the ongoing SARS-CoV-2 pandemic4. To date, sporadic cases of SARS-CoV-2 infection have been reported in dogs and cats. These include detection of SARS-CoV-2 RNA in respiratory and/or fecal specimens of dogs and cats with or without clinical signs5-7, as well as of specific antibodies in sera from pets from coronavirus disease 2019 (COVID-19) affected areas7,8. In addition, experimental infection of various animal species has demonstrated that while dogs appear poorly susceptible to SARS-CoV-2 infection, developing asymptomatic infections and shedding low-titer or no virus, cats develop respiratory pathology and shed high titers of SARS-CoV-2, even being able to infect in-contact animals9,10. Wide scale testing of susceptible species is needed to assess the extent of animal infection under more natural conditions of husbandry. Here, we conducted an extensive epidemiological survey from March to May 2020 in cats and dogs living in Italy, either in SARS-CoV-2 positive households or living in geographic areas that were severely affected by COVID-19. To our knowledge, this is the largest study to investigate SARS-CoV-2 in companion animals to date.

**Results**

All animals were sampled by their private veterinary surgeon during routine healthcare visits between March 15 and May 11, 2020 (Source data S1). Sampling of animals for this study was approved by the Ethics Committee of the Department of Veterinary Medicine, University of Bari, Italy (approval number 15/2020). A total of 603 dogs and 316 cats were sampled from different Italian regions, mostly Lombardy (476 dogs, 187 cats). Animals were sampled either from regions severely affected by COVID-19 outbreaks in humans or from those that offered convenient access to samples. Oropharyngeal (303 dogs, 173 cats), nasal (183 dogs, 78 cats), and/or rectal (66 dogs, 30 cats) swabs were collected from a total of 494 pets using synthetic fiber swabs (Table 1). For 340 dogs and 188 cats, full signalment and clinical history were collected, including breed, sex, age, exposure to COVID-19 infected humans in the previous 2 weeks (COVID-19 positive household, suspected COVID-19 positive household but not confirmed by specific assay, and COVID-19 negative household), presence of respiratory signs (cough, sneezing, conjunctivitis, nasal and/or ocular discharge). Pets living with SARS-CoV-2 infected patients included 47 dogs and 22 cats, with a single animal being sampled from each COVID-19 positive household.

Sera were available for 188 dogs and 63 cats for which complete signalment, history and location were available (Fig. 1). Additional sera were collected from diagnostic laboratories for 263 dogs and 128 cats from the affected areas, but which lacked further historical information.

Detection of SARS-CoV-2 RNA used two real-time RT-PCR assays targeting nucleoprotein and envelope protein genes11. Plaque reduction neutralization tests (PRNT)8 were performed with SARS-CoV-2/human/Liverpool/REMRQ0001/2020 isolate9. PRNT80 was determined by the highest dilution with ≥80% reduction in plaques compared to the control.

All 494 animals from which at least a swab was available tested negative for SARS-CoV-2 RNA, including 38 cats and 38 dogs that showed respiratory disease at the time of sampling, suggesting absence of active SARS-CoV-2 infection in the tested animals. In addition, 64 of these dogs and 57 of the cats that tested negative were living in households previously confirmed as having had COVID-19 and 14 animals (11 cats and 3 dogs) from COVID-19 households were displaying respiratory signs at the time of sampling.

SARS-CoV-2 neutralizing antibodies were detected in 15 dogs (3.33%, 15/451) and 11 cats (5.76%, 11/191), with titers ranging from 1:20 to 1:160 and from 1:20 to 1:1280 in dogs and cats, respectively. Of samples from households with known COVID-19 status, neutralizing antibodies were detected in 6 of 47 dogs (12.8%) and 1 of 22 cats (4.5%) from COVID-19 positive households, 1 of 7 dogs (14.3%) and 0 of 3 cats (0%) from suspected COVID-19 positive households and 2 of 133 dogs (1.5%) and 1 of 38 cats (2.6%) from COVID-19 negative households (Table 2). For those 423 animals where an age was recorded, 0 of 30 aged less than 1 year (0%), 6 of 92 aged 1-3 years (6.5%), 3 of 102 aged 4-7 years (2.9%) and 6 of 199 aged 8 and over (3.0%) tested positive. None of the animals with neutralizing antibodies displayed respiratory signs at the time of sampling.

Reference sera or ascitic fluids from animals previously shown to be positive for canine enteric coronavirus14 (3 sera), canine respiratory coronavirus15 (3 sera) and feline coronavirus16 (3 ascitic fluid) tested negative by the PRNT assay for SARS-CoV-2, confirming the specificity of the obtained results8.

Dogs were significantly more likely to test positive for SARS-CoV-2 neutralizing antibodies if they came from a known COVID-19 positive household (Fisher’s exact test, p=0.004) or were male (Fisher’s exact test, p=0.045), while there was insufficient data to assess any correlation with the neuter status, since this was reported only for 1 seropositive bitch. For provinces in the Lombardy region where at least 10 samples were available, there was a positive trend between the proportion of dogs that tested positive and the recorded burden of human disease (Spearman’s r = 0.771, p = 0.103) (Fig. 2) (Source data S2). A similar association was observed for cats (Spearman’s r = 0.696, p = 0.125).

**Discussion**

Following its original probable transmission to humans from animals, SARS-CoV-2 has spread globally within the human population with devastating health and economic impacts. To date, SARS-CoV-2 has been sporadically detected in naturally infected dogs and cats, most of which were living in close contact with infected humans. Most studies of companion animals are small in nature, likely because of an inevitable research focus on human disease. Our results from this extensive study of SARS-CoV-2 infection in owned cats and dogs living in areas where viral transmission was active in the human population confirms field observations that both cats and dogs can seroconvert under the normal conditions of pet ownership.

The link between SARS-CoV-2 household infection and a pet’s seropositivity was only apparent for dogs, possibly suggesting greater interaction between positive people and their household dogs as compared to cats. This contrasts experimental studies where dogs were less susceptible to infection9. However, this finding could be also reflective of the small numbers of seropositive cats identified. In addition, a higher proportion of male dogs were seropositive compared to female dogs, but also in this case the results could have been biased by the small size of positive animals. In fact, neuter status was unknown for the majority of seropositive animals, which prevented a full comparison with the situation in humans. Future studies in animals and humans should investigate whether this phenomenon is based in physiological or behavioral differences between males and females. Although there are clear gender differences in outcomes in human COVID-19 infections, with males at higher risk of severe disease, there seems to be no evidence for a difference in infection risk17. None of the 30 juvenile animals, less than one year-of-age, tested positive. Our findings are consistent with reports of other seropositive naturally exposed cats and dogs which were all adult6, 7, and in experimentally infected adult cats and dogs where clinical signs were not displayed18. However, in cats aged less than 1 year a higher susceptibility to SARS-CoV-2 infection was observed at 70-100 days than at 8 months of age9.

In contrast to the serology results, all animals tested negative by PCR, including those animals living in households with confirmed COVID-19 human infection and those with and without respiratory signs. This suggests that whilst pet animals can seroconvert, they may shed virus for relatively short periods of time. In experimental studies, cats stopped shedding virus by 10 days post infection (dpi) and developed neutralizing antibody responses by 7-13 dpi9, 18. Similar results were reported in experimental infection of dogs, in which virus was detected in faeces up to 6 dpi, but not in oropharyngeal swabs6. However, in a naturally infected Pomeranian dog SARS-CoV-2 RNA was detected from nasal swabs by quantitative RT-PCR for at least 13 days at low titer, whilst the virus was not detected in faecal/rectal samples7, suggesting that virus shedding patterns may vary in some animals. Half of the challenged dogs had detectable antibodies by 14 dpi. These studies and our own highlight similar challenges in detecting SARS-CoV-2 infection that exist for both humans and animals19. It is not possible with our field data set to estimate the time of infection in animals that were seropositive, and restrictions on human and animal movement during the pandemic may have delayed visits to veterinary practitioners where sampling occurred. We advocate the inclusion of pets in ongoing assessments of community and household shedding to improve detection of active infection.

In this extensive epidemiological survey of SARS-CoV-2, we found that companion animals living in areas of high human infection have been exposed to SARS-CoV-2, thus confirming previous reports of natural infections of dogs and cats with the novel coronavirus. Our results suggest that dogs warrant further investigation regarding SARS-CoV-2 susceptibility, although higher antibody titers were detected in cats, which is in agreement with previous studies suggesting that these animals are most susceptible to SARS-CoV-2 infection9, 18. We also observed seropositivity rates in animals comparable to those of humans via community sampling at a similar time in European countries20-22. This suggests that infection in companion animals is not unusual. Based on current knowledge, it is unlikely that infected pets play an active role in SARS-CoV-2 transmission to humans. However, animal-to-human transmission may be more likely under certain environmental conditions, such as the high animal population densities encountered on infected mink farms23. As and when human transmission becomes rarer and contact tracing becomes more accessible, serological surveillance of pets may be advocated to develop a holistic picture of community disease dynamics and ensure that all transmission opportunities are terminated.

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Table 1. Sample type distribution for detection of SARS-CoV-2 RNA

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Sample type | Dogs | Cats | Total |  |
| OP only | 131 | 98 | 229 |
| N only | 11 | 3 | 14 |
| R only | 0 | 0 | 0 |
| OP+N | 106 | 49 | 155 |
| OP+R | 0 | 4 | 4 |
| N+R | 0 | 0 | 0 |
| O+N+R | 66 | 26 | 92 |
| Total | 314 | 180 | 494 |

OP, oropharyngeal swab; N, nasal swab; R, rectal swab.

Table 2. Seropositivity among dogs and cats, split into risk factor groupings where data was availablea.

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | Dogs | |  |  | | Cats | |  | |  |
| Risk factor | No. + (total) | % | | *p* | No. + (total) | | % | | *p* | |
| Household |  |  | | 0.004 |  | |  | | >0.999 | |
| COVID+ | 6 (47) | 12.8% | |  | 1 (22) | | 4.5% | |  | |
| COVID- | 2 (133) | 1.5% | |  | 1 (38) | | 2.6% | |  | |
| Suspected COVID+ | 1 (7) | 14.3% | |  | 0 (3) | | 0.0% | |  | |
|  |  |  | |  |  | |  | |  | |
| Sex |  |  | | 0.045 |  | |  | | 0.492 | |
| Male | 7 (83) | 8.4% | |  | 2 (31) | | 6.5% | |  | |
| Female | 2 (105) | 1.9% | |  | 0 (30) | | 0.0% | |  | |
|  |  |  | |  |  | |  | |  | |
| Age (years) |  |  | | na |  | |  | | na | |
| < 1 | 0 (20) | 0.0% | |  | 0 (9) | | 0.0% | |  | |
| 1-3 | 5 (70) | 7.1% | |  | 1 (22) | | 4.5% | |  | |
| 4-7 | 2 (83) | 2.4% | |  | 1 (19) | | 5.3% | |  | |
| 8+ | 4 (137) | 2.9% | |  | 2 (62) | | 3.2% | |  | |
| Unknown | 4 (141) | 2.8% | |  | 7 (78) | | 9.0% | |  | |
|  |  |  | |  |  | |  | |  | |

aFor household and sex, p value determined by Fisher’s exact test. Household COVID + defined as one or more members of a household with a confirmed positive COVID-19 test. All the information was not available for all the animals.

**Figure legends.**

**Fig. 1.** Distribution of dog and cat samples assayed for neutralizing antibody titer across Italy and the region of Lombardy. Data on human COVID-19 cases from the Italian Department of Civil Protection as of May 31, 2020 and population data from the Italian National Institute of Statistics (ISTAT), January 2019. Source data are provided as a Source Data file

**Fig. 2.** Correlation of percentage of seropositive animals per province and human COVID-19 infection density in Lombardy. Data points were taken from provinces with at least 10 samples. Spearman's correlation was used to assess association. Source data are provided as a Source Data file.

Methods

*Samples*

All animals were sampled by their private veterinary surgeon during a healthcare visit for other reasons. A total of 603 dogs and 316 cats were sampled from different Italian regions, mostly Lombardy (476 dogs, 187 cats). Animals were sampled either from regions severely affected by COVID-19 outbreaks in humans or from those that offered convenient access to samples. Oropharyngeal (303 dogs, 173 cats), nasal (183 dogs, 78 cats), and/or rectal (66 dogs, 30 cats) swabs were collected from a total of 494 pets using synthetic fiber swabs (Table 1).

For 340 dogs and 188 cats, full signalment and clinical history were available, including breed, sex, age, exposure to COVID-19 infected humans (COVID-19 positive household, suspected COVID-19 positive household but not confirmed by specific assay, and COVID-19 negative household), presence of respiratory signs (cough, sneezing, conjunctivitis, nasal and/or ocular discharge).

Sera were available for 188 dogs and 63 cats for which complete signalment, history and location were available (Fig. 1). Additional sera were collected from diagnostic laboratories for 263 dogs and 128 cats from the affected areas, but which lacked further historical information.

Sampling of animals for this study was approved by the Ethics Committee of the Department of Veterinary Medicine, University of Bari, Italy (approval number 15/2020).

*Polymerase chain reaction*

Sample preparation and RNA extraction were carried out in the biosafety level 3 containment laboratory at the Department of Veterinary Medicine, University of Bari, Italy. Detection of SARS-CoV-2 RNA used two real-time RT-PCR assays targeting nucleoprotein and envelope protein genes 11. Briefly, a one-step method was adopted using with the Superscript III one step RT-PCR system with Platinum Taq Polymerase (Invitrogen srl, Milan, Italy) and the following 50-µl mixture: 25 µl of master mix, 400 nM (E\_Sarbeco\_F, E\_Sarbeco\_R), 600 nM (N\_Sarbeco\_F) or 800 nM (N\_Sarbeco\_R) of primers, 200 nM of probes (E\_Sarbeco\_P1 or N\_Sarbeco\_P) (Supplementary Table) and 10 μl of template RNA. The thermal profile consisted of incubation at 55 °C for 10 min for reverse transcription, followed by 95 °C for 3 min and then 45 cycles of 95 °C for 15 s, 58 °C for 30 s.

*Plaque reduction neutralization test (PRNT)*

The SARS-CoV-2/human/Liverpool/REMRQ0001/2020 isolate was cultured in Vero E6 cells12. For PRNT13, sera were heat inactivated at 56°C for 1 hour and stored at -20°C until use. Dulbecco’s minimal essential medium (DMEM) containing 2% fetal bovine serum (FBS) and 0.05 mg/mL gentamicin was used for serial two-fold dilutions of serum. SARS-CoV-2 at 800 PFU/mL was added to an equal volume of diluted serum and incubated at 37°C for 1 hour. The virus-serum dilution was inoculated onto Vero E6 cells, incubated at 37°C for 1 hour, and overlaid as in standard plaque assays. Cells were incubated for 48 hours at 37°C and 5% CO2 then fixed with 10% formalin and stained with 0.05% crystal violet solution. PRNT80 was determined by the highest dilution with 80% reduction in plaques compared to the control. Samples with detectable neutralizing antibody titer were repeated as technical replicates for confirmation.

*Data analysis*

Fisher’s exact test was used to analyze differences in antibody detection from households with known COVID-19 infection status, and antibody detection from male and female animals. Spearman correlation was used to analyze the relationship between human COVID-19 case numbers and detection of antibodies in animals. All statistical analyses were performed in GraphPad Prism.

Data and materials availability: The authors declare that the data supporting the findings of this study are available within the article and its Supplementary Information files, or are available from the authors upon request. Source data are provided with this paper.

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Author contributions: EIP and ND designed the study and wrote the first draft of the manuscript; SLS, ADR, TP, GTP and GLH edited the manuscript, GE, AG and SP contributed to the study’s design, CD, SLS, ERA, TP, EL, MSL, and GL performed experiments; AG, MM, AS, SL, UB, VM, FSB, VRB, ADR, UA, GTP and GLH analyzed data.

Competing interests: Authors declare no competing interests.

Additional information

Supplementary Information is available for this paper.

Supplementary Table. Oligonucleotides used in the real-time RT-PCR assays for SARS-CoV-2.

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**Fig. 1.** Distribution of dog and cat samples assayed for neutralizing antibody titer across Italy and the region of Lombardy. Data on human COVID-19 cases from the Italian Department of Civil Protection as of May 31, 2020 and population data from the Italian National Institute of Statistics (ISTAT), January 2019.



**Fig. 2.** Correlation of percentage of seropositive animals per province and human COVID-19 infection density in Lombardy. Data points were taken from provinces with at least 10 samples. Spearman's correlation was used to assess association.