**Characteristics of *Salmonella* recovered from stools of children enrolled in the Global Enteric Multicenter Study**

Running Title: Characteristics of *Salmonella* in GEMS

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**KEYWORDS:** Moderate-to-severe-diarrhea (MSD); *Salmonella*; antibiotic susceptibility; serovars; gastroenteritis

**SUMMARY:** Our data shows that non-typhoidal *Salmonella* is prevalent in stool of children under the age of 5, albeit at low levels, in Africa and South Asia. Additionally, our findings suggest that *Salmonella* Typhimurium ST313 can be carried asymptomatically by humans in sub-Saharan Africa.

**ABSTRACT**

**BACKGROUND:** The Global Enteric Multicenter Study (GEMS) determined the etiologic agents of moderate-to-severe diarrhea (MSD) in children under 5 years old in Africa and Asia. Here, we describe the prevalence and antimicrobial susceptibility of non-typhoidal *Salmonella* (NTS) serovars in GEMS and examine the phylogenetics of *Salmonella* Typhimurium ST313 isolates.

**METHODS:** *Salmonella* isolated from children with MSD or diarrhea-free controls were identified by classical clinical microbiology and serotyped using antisera and/or whole genome sequence data. We evaluated antimicrobial susceptibility using the Kirby-Bauer disk diffusion method. *Salmonella* Typhimurium sequence types were determined using multi-locus sequence typing and whole genome sequencing was performed to assess the phylogeny of ST313.

**RESULTS:** We identified *Salmonella* spp. amongst 190 (51.4%) MSD cases and 180 (48.6%) diarrhea-free controls. The most frequent *Salmonella* serovars identified were *Salmonella* Typhimurium, serogroup O:8 (C2-C3), serogroup O:6,7 (C1), *Salmonella* Paratyphi B Java and serogroup O:4 (B). The prevalence of NTS was low but similar across sites, regardless of age, and was similar amongst both cases and controls except in Kenya where *Salmonella* Typhimurium was the dominant serovar amongst cases. Phylogenetic analysis showed that these *Salmonella* Typhimurium isolates, all ST313, were highly genetically related to isolates from controls. Generally, *Salmonella* isolates were resistant to ciprofloxacin and ceftriaxone in Asia but were resistant to these antibiotics in Africa.

**CONCLUSION:** Our data suggest that NTS is prevalent, albeit at low levels, in Africa and South Asia. Our findings provide the first evidence that multi-drug resistant *Salmonella* Typhimurium ST313 can be carried asymptomatically by humans in sub-Saharan Africa.

**INTRODUCTION**

*Salmonella enterica* subspecies *enterica* serovars Typhi (Typhi), Paratyphi A (Paratyphi A) and Paratyphi B *sensu stricto* (Paratyphi B) cause enteric fever, while non-typhoidal *Salmonella* (NTS) generally cause self-limited gastroenteritis in healthy individuals. However, in young infants, the elderly and immunocompromised hosts, NTS can lead to bacteremia resulting in hospitalization and death [1]. In some resource-limited countries, NTS is a recognized etiologic agent of diarrhea [2-5] and an important risk factor for diarrhea-related morbidity and mortality in children [6]. In 2015, an estimated 38,526 children died as a result of NTS gastroenteritis; two thirds of whom lived in sub-Saharan Africa and 20% resided in Southeast Asia and South Asia [7]. Serovars Typhimurium and Enteritidis are the most common NTS isolated from cases of gastroenteritis worldwide. Despite the capacity to isolate *Salmonella* by stool culture, little is known about the prevalence of NTS serovars that cause gastroenteritis in Africa and South Asia.

Invasive NTS (iNTS) causes bacteremia in sub-Saharan Africa, occurring predominantly in infants, toddlers, as well as in HIV-infected, malnourished and/or malaria-infected adults [8-10]. Interestingly, unique clades of serovars Typhimurium and Enteritidis are associated with bacteremia in this region [8, 11, 12]. Most of the Typhimurium strains isolated from blood in sub-Saharan Africa belong to multi-locus sequence type (ST) 313 [13]. In contrast, the most common genotype isolated worldwide is ST19 which is generally associated with gastroenteritis [14] but has recently been reported as a primary cause of invasive infections in a study in Uganda [15]. Both ST19 and ST313 genotypes have been isolated from patients with either gastroenteritis or bacteremia in Kenya, although the number of diarrhea cases was low [16].

Provision of antibiotics to treat uncomplicated NTS gastroenteritis in children is not recommended, except where progression to invasive disease is a risk [17, 18]. However, information about the antimicrobial susceptibility of NTS is useful as this knowledge contributes to our overall understanding of resistance markers that are circulating in specific geographic locations. In fact, NTS harboring antimicrobial resistance traits in the gastrointestinal tract could serve as a reservoir for iNTS [19]. Presently, countries with the highest burden of iNTS disease report 48-75% multidrug resistance (MDR) to commonly-used antibiotics; a major concern given that more effective 3rd generation cephalosporins or fluoroquinolones may be less available or more costly in these settings [9, 20].

During 2007–2010, the Global Enteric Multicenter Study (GEMS) determined the etiologic agents of moderate-to-severe diarrhea (MSD) in children 0-59 months old living in The Gambia, Mali, Mozambique, Kenya, India, Bangladesh and Pakistan [21]. This large, prospective, case-control study determined that NTS was significantly associated with MSD in infants (0-11 months) from the Bangladesh site and toddlers (12-23 m) and young children (24-59 m) from the Kenya site [21]. Here, we determined the prevalence of *Salmonella* serovars isolated in GEMS, evaluated antimicrobial susceptibility, identified Typhimurium sequence types, and examined the phylogenetic relatedness of Typhimurium ST313 isolates.

**METHODS**

**GEMS study participants**

The methods and main findings from GEMS have previously been described [21-23]. Briefly, GEMS participants were recruited from censused populations during 2007-2010 in The Gambia, Mali, Mozambique, Kenya, Bangladesh, India and Pakistan. Study participants included children aged 0-59 months of age with MSD who presented to a sentinel health facility (see Supplementary text for additional details). Children were recruited into 0-11-, 12-23- and 24-59-month age groups. For each child with MSD (case) enrolled, 1-3 children without diarrhea during the previous week (controls) were recruited. Scientific and ethical committees and Institutional Review Boards (IRBs) of participating institutions in each country as well as the coordinating institution, University of Maryland, Baltimore, approved the study protocol prior to implementation. Informed consent was obtained in the local dialect from all participating caretakers before recruitment of their children into the study.

**Detection of *Salmonella* spp.**

A panel of enteropathogens was identified from stool specimens, collected at the clinic from MSD cases or obtained at home by caregivers of children in the control group, as previously described [23]. *Salmonella* spp.were shipped to the Center for Vaccine Development and Global Health (CVD) at the University of Maryland School of Medicine for additional characterization.

**Characterization of *Salmonella* serovars from stools**

At CVD, *Salmonella* spp. were agglutinated using polyvalent O and O1 antisera followed by serogroups O:2 (A), O:4 (B), O:6,7 and O:7 (C1), O:6,8 and O:8 (C2-C3), O:9 (D1), O:9,46 (D2), O:3,10 (E1), O:11 (F), O:13 (G) antisera (Denka Seiken, Tokyo, Japan). Serovars Typhimurium, Typhi, Enteritidis and Paratyphi B were fully serotyped (using O and H typing antisera) and additionally confirmed by PCR [24, 25].

**Sequence typing of Typhimurium isolates**

Sequence types were determined for all 87 Typhimurium isolates using Multi-Locus Sequence Typing (MLST) by PCR and sequencing and/or by examining whole genome sequences. Sequence typing by MLST followed methodology described previously [14].

## Whole genome sequencing and phylogenetic analysis

The majority of the *Salmonella* isolates (355 out of 370) were subjected to whole genome sequencing (WGS). Following sequencing, 120 isolates were excluded from subsequent analyses as they did not meet the quality control criteria. Details of sequencing and phylogenetic analyses are described in the Supplementary text.

**Antimicrobial susceptibility testing**

The susceptibility of the 370 *Salmonella* isolates to chloramphenicol, ampicillin, ciprofloxacin, trimethoprim/sulfamethoxazole (TMP/SMX), gentamicin and ceftriaxone was determined using the Kirby-Bauer disk diffusion method and interpreted according to Clinical and Laboratory Standards Institute (CLSI) guidelines. Multidrug resistance (MDR) was defined as resistance to ampicillin, chloramphenicol and TMP/SMX.

**Statistical analysis**

To determine which individual *Salmonella* serovars were driving the association between *Salmonella* and MSD that was found in the original GEMS analyses [21], we used the same conditional logistic regression model as previously described [26]. Instead of including *Salmonella* species in the model we included variables for each *Salmonella* serogroup/serovar [18, 33]. The association of each serovar with MSD was adjusted for other co-pathogens (Supplementary Table 2). The rationale for this approach, generally, and in the unique context of GEMS, has been discussed previously [26]. Analyses were conducted using R version 3.3.2. P-values < 0.05 were considered statistically significant.

**RESULTS**

**Characteristics of MSD study participants**

Among the MSD cases, *Salmonella* spp. were isolated from stool of 86 (44.2%), 55 (28.9%), and 49 (25.8%) childrenin the 0-11 m, 12-23 m and 24-59 m age groups, respectively (Table 1). Cases experienced severe signs of MSD. Approximately 20% of infants with *Salmonella* spp. detected had bloody diarrhea, while 100% of children 12-to 59-months-old with *Salmonella* spp. produced watery diarrhea. Of note, a lower proportion of children with MSD who had *Salmonella* isolated tended to be female in all age groups.

**Geographical distribution and prevalence of *Salmonella* serovars**

The serovar distribution of the 370 *Salmonella* isolates (190 from cases and 180 from controls) collected from stools of study participants is shown in Figure 1. Of these, 361 were NTS. Additionally, we recovered eight Typhi from Asia and one Paratyphi A isolate from Bangladesh. The most frequent NTS serovars identified were Typhimurium, serogroup O:8 (C2-C3), serogroup O:6,7 (C1), Paratyphi B Java and serogroup O:4 (other than Typhimurium or Paratyphi B). Serovar Typhimurium predominated in Africa, whereas serogroup O:6,7 (C1) and O:8 (C2-C3) serovars were the most common in Asia.

**Prevalence of *Salmonella* serovars by site and age stratum**

The prevalence of the most abundant serovars isolated in GEMS, as well as serovar Enteritidis (due to its importance in iNTS disease in Africa), are shown in Table 2. The individual serovars of isolates are listed in Supplementary Table 1. In general, we found that the rates of NTS isolation were low (≤ 5.3%) in both cases and controls regardless of age groups, although some site-to-site variation was apparent. At the Kenya site, serovar Typhimurium, the most prevalent serovar, was recovered in stools of MSD cases at a rate of 3.3% for infants, 3.7% for toddlers and 4.3% for young children. In Bangladesh, Paratyphi B Java, the most prevalent serovar there, was recovered from 2.0% of infants with MSD. Serogroup O:8 (C2-C3) organisms were most prevalent in stools at the Pakistan site. The prevalence of NTS in cases and controls in The Gambia, Mali, Mozambique and India was less than 1.5%.

***Salmonella* serovars significantly associated with MSD**

Previously, 3.2% and 3.7% of MSD episodes in toddlers (12-23 m) and children (24-59 m) at the Kenya site, respectively, and 4.6% of MSD episodes in infants at the Bangladesh site were shown to be attributable to *Salmonella* [21]. We determined the serovars driving the associations by using a conditional logistic regression model (Supplementary Table 2). In Bangladesh, serogroup O:6,7 (C1) (Odd’s Ratio [OR] = 6.4, 95% CI = 1.84-22.58), O:8 (C2-C3) (OR = 6.0, 95% CI = 1.28-28.33) and serovar Paratyphi B Java (OR = 4.8, 95% CI = 1.87-12.29) were significantly associated with MSD. In Kenya, the association was driven by serovar Typhimurium among children aged 12-23 m (OR = 4.3, 95% CI = 1.86-9.93) and 24-59 m (OR = 4.9, 95% CI = 2.09-11.64). All other serovars occurred in too few cases and controls to produce significant results.

**Antimicrobial susceptibility at GEMS sites**

*Salmonella* isolates from the African and Asian sites differed in terms of their antimicrobial susceptibility (Figure 2). Isolates from Africa were susceptible to ciprofloxacin and ceftriaxone whereas resistance to these antibiotics was observed amongst Asian NTS isolates. We observed 36.7% of NTS from MSD cases in Kenya to be multidrug resistant (MDR) to ampicillin, trimethoprim/sulfamethoxazole (TMP/SMX) and chloramphenicol. However, isolation of non-susceptible NTS was less frequent at The Gambian site; only 2.9% of NTS from cases showed an MDR phenotype (Figure 2A). In Asia, Indian isolates showed more resistance to the antibiotics tested than isolates from the other two Asian sites (Figure 2B). We observed similar antimicrobial susceptibility profiles amongst NTS from controls as from cases at each site except for Kenya and India.

Serovars Typhimurium and Enteritidis from Africa and serogroup O:6,7 (C1) and serogroup O:13 (G) isolates from Asia showed the highest percentage of antimicrobial resistance (Figure 3). All Enteritidis and Paratyphi B Java isolates recovered from GEMS stools from Asia were pan-susceptible to antimicrobial agents, while the six serogroup O:13 (G) isolates from Africa were pan-susceptible. Five of the eight (62.5%) serovar Typhi from Asia (Pakistan and India) were MDR.

To assess whether the high resistance of NTS to antimicrobials was associated with antibiotic prescription rate, we determined the percentage of children with MSD (and *Salmonella* isolated in stools) who had been prescribed (but may or may not have been given) antimicrobial agents after visiting any of the sentinel health facilities that participated in GEMS (Table 3). In general, most MSD cases with *Salmonella* had been prescribed or given an antibiotic (except in Pakistan). TMP/SMX was the most commonly prescribed antibiotic in Africa whereas ciprofloxacin was the most common in Asia.

## Phylogenetic analysis of *Salmonella* Typhimurium

Since Typhimurium was the most important cause of iNTS disease at several GEMS sites and was the most frequent serovar isolated from stools, a phylogenetic analysis was performed. Of 87 Typhimurium isolates, 74 (85.0%) were from Africa (Kenya), while 13 (14.9%) were from Asia (Pakistan, India and Bangladesh). Table 4 shows the sequence types (ST) of these Typhimurium isolates listed by site of origin.

A phylogeny was constructed using whole genome sequences to determine the relationship between the Typhimurium ST313 isolates from MSD cases and controls (Figure 4). The African ST313 sequence type has been divided into the older lineage 1 isolates, and the more recent lineage 2 [12, 27]. Here, 50 of 55 study isolates analyzed (90.9%) clustered with the ST313 lineage 2 reference genome D23580, 49 of 55 (81%) of which showed the typical MDR phenotype associated with lineage 2, namely resistance to chloramphenicol, co-trimoxazole and ampicillin. The lineage 2 isolates from the MSD cases and diarrhea-free controls were closely related and could not be distinguished phylogenetically. A group of 5 isolates in stools of cases and controls, regardless of age, formed a small lineage 2 sub-cluster associated with susceptibility to chloramphenicol.

Four of the 55 isolates (7%) clustered with the ST313 lineage 1 reference genome A130, all of which were isolated from MSD cases in Kenya and were sensitive to chloramphenicol. Of note, the one study isolate (specimen 700477) that failed to cluster with lineage 1 or lineage 2 and demonstrated pan-susceptibility to antibiotics was isolated from a case of MSD from Pakistan.

**DISCUSSION**

*Salmonella* isolates were detected in stools of children with MSD and from diarrhea-free community controls at each of the GEMS sites. A primary finding of our analysis was that except for Typhimurium, the prevalence of most *Salmonella* serovars was similar in stools of cases and controls, regardless of age and across study sites. Because children enrolled as controls in GEMS only had to have been free of diarrhea for the previous seven days, we could not rule out asymptomatic carriage or shedding of *Salmonella* among controls due to persistent excretion or convalescence [28]. NTS are reportedly excreted for longer periods in children than adults, lasting from several weeks to months [17, 29]. We found that NTS was as prevalent in cases as in controls, which suggests that NTS is endemic at the seven GEMS sites [3, 30, 31].

In this study, we report the association of Typhimurium ST313 with acute diarrhea, showing that these bacteria cause diarrhea and are not just associated with invasive disease. This observation is supported by recent studies from Kenya, Central African Republic and Democratic Republic of Congo which also detected Typhimurium ST313 in stool [16, 32, 33]. Typhimurium ST313 was identified in both MSD cases and controls, suggesting that this important sequence type can be carried asymptomatically by humans. Phylogenetic analysis identified lineages 1 and 2, in accordance with previous findings [13]. We found that the same ST313 lineage (lineage 2) was prevalent in the stools of both MSD cases and controls. The fact that isolates from cases and controls are found in every part of the phylogeny suggests that the *Salmonellae* that cause MSD are closely related to those associated with asymptomatic carriage.

Several groups have attempted to identify the reservoir of iNTS isolates in Africa. Kariuki *et al* [19] were the first to suggest that iNTS are not acquired zoonotically, but are acquired by anthroponotic transmission. In this and other studies, NTS isolated from blood cultures of bacteremic index cases were highly similar to isolates from household contacts but different from NTS from animal or environmental sources taken from around the homes of index cases [19, 34]. Collectively, these prior studies suggest that the reservoir for Typhimurium ST313 is indeed humans. It remains possible the lack of detection of *Salmonella* spp. from animals and the environment reflects difficulties in culture from these specimen types. However, if the inference from the above-mentioned studies is correct, our data would support these findings by showing that Typhimurium strains isolated from stool of cases and controls in GEMS are highly genetically related to isolates from blood.

When we examined antimicrobial susceptibility of GEMS NTS isolates, we detected marked regional differences in resistance. We observed similar antimicrobial susceptibility patterns in stools of cases and asymptomatic controls at all GEMS sites except Kenya and India. Our data suggest that antibiotic resistant NTS are circulating in the GEMS communities. Non-susceptible NTS strains could serve as a reservoir from which antibiotic resistance determinants can spread horizontally to other microorganisms [35]. In Africa, the majority of Typhimurium and Enteritidis isolates were MDR which is consistent with previous findings [19, 36]. Importantly, none of the isolates from GEMS African sites were resistant to ciprofloxacin or ceftriaxone in contrast to isolates from Asia, suggesting a difference in utilization of these antibiotics. Five (of 8) Typhi from India and Pakistan were MDR but none were extensively drug resistant (XDR) as seen in the recent typhoid fever outbreak in Hyderabad, Pakistan [37].

Antibiotics are not recommended for treatment of NTS gastroenteritis in pediatric patients due to the predisposition for extended excretion of bacteria and relapse of infection [17, 38, 39]. However, our data suggest that children with NTS disease are being prescribed antibiotics (presumably not to treat the NTS since the treating clinician was unlikely to have known that the infection was due to NTS), which has selected for resistant bacteria. We observed high prescription rates for ciprofloxacin and other fluoroquinolones in Asia and not surprisingly, also high resistance of *Salmonella* to ciprofloxacin in Asia, but not Africa (where ciprofloxacin was rarely prescribed). In contrast, we recorded high antibiotic prescription rates of TMP/SMX in Africa which possibly led to the high resistance observed in Africa. TMP/SMX in combination with highly active antiretroviral therapy (HAART) has been used routinely as prophylaxis for opportunistic infections in patients with HIV in Africa [40].

The low frequency of *Salmonella* in MSD cases from Mali, The Gambia and Mozambique was somewhat unexpected given that these countries report high iNTS disease burdens [3, 30, 31]. However, the incidence of iNTS disease during GEMS (2007-2010) in these three countries decreased relative to earlier estimates, concomitant with a reduction in clinical malaria [3, 30, 31, 41]. Indeed, there is growing evidence to suggest that iNTS disease is correlated with clinical malaria and that efforts to control malaria have resulted in reduced iNTS disease incidence [10, 42]. A re-analysis of GEMS using quantitative molecular diagnostic methods showed higher attributable fractions for *Salmonella* in all age groups at all sites [43]. We speculate that the low frequency of *Salmonella* isolation in Mali, The Gambia and Mozambique reflects differences in conventional clinical microbiology techniques that were used at these sites.

Our findings have three main implications: 1) the prevalence data could be used to refine incidence estimates for individual *Salmonella* serovars; 2) we report for the first time the association of Typhimurium ST313 with acute diarrhea, thereby showing that these bacteria are not just associated with invasive disease; and 3) our data demonstrates wide-spread asymptomatic carriage of ST313, a key cause of iNTS infections. Because we have found that humans are carriers of MDR *Salmonella* strains that also cause iNTS [44], it is possible that these individuals serve as intermediaries in transmission and maintenance of these bacteria in the community.

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**Conflicts of Interest**

Drs Tennant and Levine are co-inventors (covered by multiple patents) of a trivalent *Salmonella* (Enteritidis/ Typhimurium/ Typhi Vi) conjugate vaccine and live attenuated NTS vaccines.

**Disclaimer**

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention.

**REFERENCES**

1. Jones TF, Ingram LA, Cieslak PR, et al. Salmonellosis outcomes differ substantially by serotype. J Infect Dis **2008**; 198(1): 109-14.

2. Cardemil CV, Sherchand JB, Shrestha L, et al. Pathogen-Specific Burden of Outpatient Diarrhea in Infants in Nepal: A Multisite Prospective Case-Control Study. J Pediatric Infect Dis Soc **2017**; 6(3): e75-e85.

3. Kwambana-Adams B, Darboe S, Nabwera H, et al. *Salmonella* Infections in The Gambia, 2005-2015. Clin Infect Dis **2015**; 61 Suppl 4: S354-62.

4. Leung DT, Das SK, Malek MA, et al. Non-typhoidal *Salmonella* gastroenteritis at a diarrheal hospital in Dhaka, Bangladesh, 1996-2011. Am J Trop Med Hyg **2013**; 88(4): 661-9.

5. Taniuchi M, Sobuz SU, Begum S, et al. Etiology of diarrhea in Bangladeshi infants in the first year of life analyzed using molecular methods. J Infect Dis **2013**; 208(11): 1794-802.

6. O'Reilly CE, Jaron P, Ochieng B, et al. Risk factors for death among children less than 5 years old hospitalized with diarrhea in rural western Kenya, 2005-2007: a cohort study. PLoS Med **2012**; 9(7): e1001256.

7. Estimates of global, regional, and national morbidity, mortality, and aetiologies of diarrhoeal diseases: a systematic analysis for the Global Burden of Disease Study 2015. Lancet Infect Dis **2017**; 17(9): 909-48.

8. Feasey NA, Dougan G, Kingsley RA, Heyderman RS, Gordon MA. Invasive non-typhoidal *Salmonella* disease: an emerging and neglected tropical disease in Africa. Lancet **2012**; 379(9835): 2489-99.

9. Marks F, von Kalckreuth V, Aaby P, et al. Incidence of invasive *Salmonella* disease in sub-Saharan Africa: a multicentre population-based surveillance study. Lancet Glob Health **2017**; 5(3): e310-e23.

10. Park SE, Pak GD, Aaby P, et al. The Relationship Between Invasive Nontyphoidal *Salmonella* Disease, Other Bacterial Bloodstream Infections, and Malaria in Sub-Saharan Africa. Clin Infect Dis **2016**; 62 Suppl 1: S23-31.

11. Feasey NA, Masesa C, Jassi C, et al. Three Epidemics of Invasive Multidrug-Resistant *Salmonella* Bloodstream Infection in Blantyre, Malawi, 1998-2014. Clin Infect Dis **2015**; 61 Suppl 4: S363-71.

12. Okoro CK, Barquist L, Connor TR, et al. Signatures of adaptation in human invasive *Salmonella* Typhimurium ST313 populations from sub-Saharan Africa. PLoS Negl Trop Dis **2015**; 9(3): e0003611.

13. Kingsley RA, Msefula CL, Thomson NR, et al. Epidemic multiple drug resistant *Salmonella* Typhimurium causing invasive disease in sub-Saharan Africa have a distinct genotype. Genome Res **2009**; 19(12): 2279-87.

14. Achtman M, Wain J, Weill FX, et al. Multilocus sequence typing as a replacement for serotyping in *Salmonella enterica*. PLoS Pathog **2012**; 8(6): e1002776.

15. Molly Freeman HK, Hannington Tasimwa , Susan Van Duyne , Hayat Caidi , Ana Lauer , Matthew Mikoleit. Characterization of *Salmonella* Isolates From Invasive Infections Collected During Acute Febrile Illness (AFI) Surveillance n Uganda From 2016-2018. SABIN Coaliation Against Typhoid Conference Booklet-Hanoi Vietnam, March **2019**: Abstract 37.

16. Akullian A, Montgomery JM, John-Stewart G, et al. Multi-drug resistant non-typhoidal *Salmonella* associated with invasive disease in western Kenya. PLoS Negl Trop Dis **2018**; 12(1): e0006156.

17. Rossier P, Urfer E, Burnens A, et al. Clinical features and analysis of the duration of colonisation during an outbreak of *Salmonella* Braenderup gastroenteritis. Schweiz Med Wochenschr **2000**; 130(34): 1185-91.

18. Onwuezobe IA, Oshun PO, Odigwe CC. Antimicrobials for treating symptomatic non-typhoidal *Salmonella* infection. Cochrane Database Syst Rev **2012**; 11: Cd001167.

19. Kariuki S, Revathi G, Kariuki N, et al. Invasive multidrug-resistant non-typhoidal *Salmonella* infections in Africa: zoonotic or anthroponotic transmission? J Med Microbiol **2006**; 55(Pt 5): 585-91.

20. Kariuki S, Gordon MA, Feasey N, Parry CM. Antimicrobial resistance and management of invasive *Salmonella* disease. Vaccine **2015**; 33 Suppl 3: C21-9.

21. Kotloff KL, Nataro JP, Blackwelder WC, et al. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. Lancet **2013**; 382(9888): 209-22.

22. Kotloff KL, Blackwelder WC, Nasrin D, et al. The Global Enteric Multicenter Study (GEMS) of diarrheal disease in infants and young children in developing countries: epidemiologic and clinical methods of the case/control study. Clin Infect Dis **2012**; 55 Suppl 4: S232-45.

23. Panchalingam S, Antonio M, Hossain A, et al. Diagnostic microbiologic methods in the GEMS-1 case/control study. Clin Infect Dis **2012**; 55 Suppl 4: S294-302.

24. Levy H, Diallo S, Tennant SM, et al. PCR method to identify *Salmonella* *enterica* serovars Typhi, Paratyphi A, and Paratyphi B among *Salmonella* Isolates from the blood of patients with clinical enteric fever. J Clin Microbiol **2008**; 46(5): 1861-6.

25. Tennant SM, Diallo S, Levy H, et al. Identification by PCR of non-typhoidal *Salmonella* *enterica* serovars associated with invasive infections among febrile patients in Mali. PLoS Negl Trop Dis **2010**; 4(3): e621.

26. Blackwelder WC, Biswas K, Wu Y, et al. Statistical methods in the Global Enteric Multicenter Study (GEMS). Clin Infect Dis **2012**; 55 Suppl 4: S246-53.

27. Okoro CK, Kingsley RA, Connor TR, et al. Intracontinental spread of human invasive *Salmonella* Typhimurium pathovariants in sub-Saharan Africa. Nat Genet **2012**; 44(11): 1215-21.

28. Levine MM, Robins-Browne RM. Factors that explain excretion of enteric pathogens by persons without diarrhea. Clin Infect Dis **2012**; 55 Suppl 4: S303-11.

29. Molbak K, Wested N, Hojlyng N, et al. The etiology of early childhood diarrhea: a community study from Guinea-Bissau. J Infect Dis **1994**; 169(3): 581-7.

30. Mandomando I, Bassat Q, Sigauque B, et al. Invasive *Salmonella* Infections Among Children From Rural Mozambique, 2001-2014. Clin Infect Dis **2015**; 61 Suppl 4: S339-45.

31. Tapia MD, Tennant SM, Bornstein K, et al. Invasive Nontyphoidal *Salmonella* Infections Among Children in Mali, 2002-2014: Microbiological and Epidemiologic Features Guide Vaccine Development. Clin Infect Dis **2015**; 61 Suppl 4: S332-8.

32. Breurec S, Reynaud Y, Frank T, et al. Serotype distribution and antimicrobial resistance of human *Salmonella enterica* in Bangui, Central African Republic, from 2004 to 2013. PLoS Negl Trop Dis **2019**; 13(12): e0007917.

33. Phoba MF, Barbé B, Ley B, et al. High genetic similarity between non-typhoidal *Salmonella* isolated from paired blood and stool samples of children in the Democratic Republic of the Congo. PLoS Negl Trop Dis **2020**; 14(7): e0008377.

34. Post AS, Diallo SN, Guiraud I, et al. Supporting evidence for a human reservoir of invasive non-Typhoidal *Salmonella* from household samples in Burkina Faso. PLoS Negl Trop Dis **2019**; 13(10): e0007782.

35. McInnes RS, McCallum GE, Lamberte LE, van Schaik W. Horizontal transfer of antibiotic resistance genes in the human gut microbiome. Curr Opin Microbiol **2020**; 53: 35-43.

36. Gordon MA, Graham SM, Walsh AL, et al. Epidemics of invasive *Salmonella enterica* serovar Enteritidis and *S*. *enterica* serovar Typhimurium infection associated with multidrug resistance among adults and children in Malawi. Clin Infect Dis **2008**; 46(7): 963-9.

37. Qamar FN, Yousafzai MT, Khalid M, et al. Outbreak investigation of ceftriaxone-resistant *Salmonella enteric*a serotype Typhi and its risk factors among the general population in Hyderabad, Pakistan: a matched case-control study. Lancet Infect Dis **2018**; 18(12): 1368-76.

38. Nelson JD, Kusmiesz H, Jackson LH, Woodman E. Treatment of Salmonella gastroenteritis with ampicillin, amoxicillin, or placebo. Pediatrics **1980**; 65(6): 1125-30.

39. Stapels DAC, Hill PWS, Westermann AJ, et al. Salmonella persisters undermine host immune defenses during antibiotic treatment. Science **2018**; 362(6419): 1156-60.

40. Crook AM, Turkova A, Musiime V, et al. Tuberculosis incidence is high in HIV-infected African children but is reduced by co-trimoxazole and time on antiretroviral therapy. BMC Med **2016**; 14: 50.

41. Institut National de la Statistique (INSTAT) CdPedSSS-DvSePdlFCS-D-PeI. Enquête Démographique et de Santé au Mali 2018. Bamako, Mali et Rockville, Maryland, USA

42. Tabu C, Breiman RF, Ochieng B, et al. Differing burden and epidemiology of non-Typhi *Salmonella* bacteremia in rural and urban Kenya, 2006-2009. PLoS One **2012**; 7(2): e31237.

43. Liu J, Platts-Mills JA, Juma J, et al. Use of quantitative molecular diagnostic methods to identify causes of diarrhoea in children: a reanalysis of the GEMS case-control study. Lancet **2016**; 388(10051): 1291-301.

44. Collaborators GBDN-TSID. The global burden of non-typhoidal *Salmonella* invasive disease: a systematic analysis for the Global Burden of Disease Study 2017. Lancet Infect Dis **2019**; 19(12): 1312-24.

**Table 1. Characteristics of children with moderate-to-severe diarrhea (MSD) and from whom *Salmonella* were isolated**

|  |  |  |  |
| --- | --- | --- | --- |
| **Clinical signs and symptoms**a | **0-11 m (n=86)** | **12-23 m (n=55)** | **24-59 m (n=49)** |
| **Stool consistency** |  |  |  |
| Mucus | 72.94% | 61.82% | 53.06% |
| Pus | 3.49% | 9.09% | 12.24% |
| Bloody | 24.42% | 12.73% | 0 |
| Watery | 75.58% | 87.27% | 100 |
| **Medical history** |  |  |  |
| Vomiting >3 times/day | 40.70% | 40.0% | 48.98% |
| Drank much less than usual | 19.77% | 21.82% | 14.29% |
| Very thirsty | 59.3% | 67.27% | 83.33% |
| Decreased activity or lethargy | 36.05% | 54.55% | 53.06% |
| Irritable or restless | 45.35% | 61.82% | 55.10% |
| Fever >38C or parent perception | 73.26% | 72.73% | 77.55% |
| **Physical examination** |  |  |  |
| Admitted to the hospital | 17.44% | 18.18% | 22.45% |
| Undernutrition | 9.30% | 16.36% | 12.24% |
| Loss of skin turgor | 26.74% | 25.45% | 36.73% |
| Dry mouth | 54.65% | 74.55% | 81.63% |
| Sunken eyes | 65.12% | 85.45% | 87.76% |
| Axillary temperature >38.3C | 18.60% | 21.82% | 26.53% |
| **Gender** |  |  |  |
| Female gender | 37.21% | 45.45% | 40.82% |

**Table 2. Prevalence of NTS at GEMS sites among children with moderate-to-severe-diarrhea (cases) and controls**

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | Basse, The Gambia | | Bamako, Mali | | Manhica, Mozambique | | Nyanza Province, Kenya | | Kolkata, India | | Mirzapur, Bangladesh | | Karachi (Bin Qasim Town), Pakistan | |
| Prevalence of NTS | Cases | Controls | Cases | Controls | Cases | Controls | Cases | Controls | Cases | Controls | Cases | Controls | Cases | Controls |
| **0-11 months** |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| No. of participants | 400 | 585 | 727 | 727 | 374 | 697 | 673 | 673 | 672 | 685 | 550 | 878 | 633 | 633 |
| **No. of NTS** | **5 (1.3%)** | **12 (2.1%)** | **0** | **0** | **4 (0.8%)** | **1 (0.1%)** | **34 (5.1%)** | **29 (4.3%)** | **1 (0.1%)** | **10 (1.5%)** | **27 (4.9%)** | **14 (1.6%)** | **15 (2.4%)** | **24 (3.8%)** |
| Typhimurium | 0 | 0 | 0 | 0 | 0 | 0 | 22 (3.3%) | 9 (1.3%) | 0 | 2 (0.6%) | 1 (0.2%) | 0 | 1 (0.2%) | 1 (0.2%) |
| Enteritidis | 0 | 0 | 0 | 0 | 1 (0.3%) | 0 | 2 (0.3%) | 2 (0.3%) | 0 | 0 | 0 | 0 | 0 | 0 |
| Paratyphi B Java | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 (0.5%) | 11 (2.0%) | 8 (0.9%) | 1 (0.2%) | 0 |
| Serogroup O:4 | 0 | 0 | 0 | 0 | 0 | 1 (0.1%) | 2 (0.3%) | 5 (0.7%) | 0 | 1 (0.5%) | 1 (0.2%) | 0 | 2 (0.3%) | 2 (0.3%) |
| Serogroup O:6,7 | 1 (0.3%) | 2 (0.3%) | 0 | 0 | 0 | 0 | 3 (0.4%) | 3 (0.4%) | 0 | 0 | 9 (1.6%) | 4 (0.5%) | 4 (0.6%) | 4 (0.6%) |
| Serogroup O:8 | 0 | 1 (0.2%) | 0 | 0 | 1 (0.3%) | 0 | 2 (0.3%) | 4 (0.6%) | 1 (0.1%) | 1 (0.5%) | 5 (0.9%) | 1 (0.1%) | 7 (1.1%) | 13 (2.1%) |
| Other serovars | 4 (1.0%) | 9 (1.5%) | 0 | 0 | 2 (0.5%) | 0 | 3 (0.4%) | 6 (0.9%) | 0 | 5 (0.7%) | 0 | 1 (0.1%) | 0 | 4 (0.6%) |
| **12-23 months** |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| No. of participants | 455 | 639 | 682 | 695 | 195 | 391 | 410 | 621 | 588 | 598 | 476 | 761 | 399 | 676 |
| **No. of NTS** | **6 (1.3%)** | **8 (1.3%)** | **1 (0.1%)** | **0** | **1 (0.5%)** | **0** | **24 (5.9%)** | **21 (3.4%)** | **1 (0.2%)** | **2 (0.3%)** | **7 (1.5%)** | **9 (1.2%)** | **14 (3.5%)** | **19 (2.8%)** |
| Typhimurium | 0 | 0 | 0 | 0 | 0 | 0 | 15 (3.7%) | 6 (1.0%) | 0 | 0 | 2 (0.4%) | 1 (0.1%) | 1 (0.3%) | 3 (0.4%) |
| Enteritidis | 0 | 0 | 0 | 0 | 0 | 0 | 4 (1.0%) | 2 (0.3%) | 0 | 0 | 2 (0.4%) | 0 | 1 (0.3%) | 1 (0.1%) |
| Paratyphi B Java | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 3 (0.6%) | 2 (0.3%) | 0 | 0 |
| Serogroup O:4 | 1 (0.2%) | 0 | 1 (0.1%) | 0 | 1 (0.5%) | 0 | 1 (0.2%) | 3 (0.5%) | 0 | 0 | 0 | 1 (0.1%) | 2 (0.5%) | 2 (0.3%) |
| Serogroup O:6,7 | 2 (0.4%) | 4 (0.6%) | 0 | 0 | 0 | 0 | 0 | 2 (0.3%) | 1 (0.2%) | 1 (0.2%) | 0 | 1 (0.1%) | 0 | 4 (0.6%) |
| Serogroup O:8 | 1 (0.2%) | 1 (0.2%) | 0 | 0 | 0 | 0 | 3 (0.7%) | 4 (0.6%) | 0 | 1 (0.2%) | 0 | 4 (0.5%) | 4 (1.0%) | 7 (1.0%) |
| Other serovars | 2 (0.4%) | 3 (0.5%) | 0 | 0 | 0 | 0 | 1 (0.2%) | 4 (0.6%) | 0 | 0 | 0 | 0 | 6 (1.5%) | 2 (0.3%) |
| **24-59 months** |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| No. of participants | 174 | 345 | 624 | 642 | 112 | 208 | 393 | 589 | 308 | 731 | 368 | 826 | 226 | 529 |
| **No. of NTS** | **4 (2.3%)** | **0** | **1 (0.2%)** | **0** | **0** | **0** | **20 (5.1%)** | **11 (1.9%)** | **1 (1.0%)** | **2 (0.3%)** | **7 (2.2%)** | **6 (0.7%)** | **10 (5.8%)** | **10 (2.3%)** |
| Typhimurium | 0 | 0 | 0 | 0 | 0 | 0 | 17 (4.3%) | 5 (0.8%) | 0 | 0 | 0 | 0 | 0 | 1 (0.2%) |
| Enteritidis | 0 | 0 | 0 | 0 | 0 | 0 | 1 (0.3%) | 1 (0.2%) | 0 | 0 | 1 (0.3%) | 0 | 0 | 0 |
| Paratyphi B Java | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 (0.5%) | 2 (0.2%) | 0 | 0 |
| Serogroup O:4 | 0 | 0 | 1 (0.2%) | 0 | 0 | 0 | 1 (0.3%) | 0 | 0 | 0 | 0 | 0 | 1 (0.4%) | 3 (0.6%) |
| Serogroup O:6,7 | 3 (1.7%) | 0 | 0 | 0 | 0 | 0 | 1 (0.3%) | 0 | 1 (0.3%) | 2 (0.3%) | 2 (0.5%) | 2 (0.2%) | 2 (0.9%) | 1 (0.2%) |
| Serogroup O:8 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 (0.3%) | 0 | 0 | 0 | 0 | 3 (1.3%) | 1 (0.2%) |
| Other serovars | 1 (0.6%) | 0 | 0 | 0 | 0 | 0 | 0 | 3 (0.5%) | 0 (0.6%) | 0 | 2 (0.8%) | 2 (0.2%) | 4 (3.1%) | 4 (1.1%) |
| **Total participants (all ages)** | **1029** | **1569** | **2033** | **2064** | **681** | **1296** | **1476** | **1883** | **1568** | **2014** | **1394** | **2465** | **1258** | **1838** |
| **Total no. of NTS (all ages)** | **15 (1.5%)** | **20 (1.3%)** | **2 (0.1%)** | **0** | **5 (0.9%)** | **1 (0.1%)** | **78 (5.3%)** | **61 (3.2%)** | **3 (0.2%)** | **14 (0.7%)** | **41 (2.9%)** | **29 (1.2%)** | **39 (3.1%)** | **53 (2.9%)** |

**Table 3. Proportion of children with MSD and positive for *Salmonella* prescribed any antimicrobial agents after seeking care at sentinel health facilities (SHF) that participated in GEMS at sites in Africa and Asia**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Antimicrobial agenta | Africa | The Gambia | Mali | Mozambique | Kenya | Asia | India | Bangladesh | Pakistan |
| No. of casesb | 100 | 15 | 2 | 5 | 78 | 90 | 5 | 42 | 43 |
| No antibiotics prescribed / given | 23 (23.0%) | 4 (26.7%) | 0 | 0 | 19 (24.4%) | 36 (40.0%) | 0 | 1 (2.4%) | 35 (81.4%) |
| Any antibiotics prescribed / given | 77 (77.0%) | 11 (73.3%) | 2 (100%) | 5 (100%) | 59 (75.6%) | 54 (60.0%) | 5 (100%) | 41 (97.6%) | 8 (18.6%) |
| **Ampicillin** | **1 (1.0%)** | **0** | **0** | **1 (20.0%)** | **0** | **0** | **0** | **0** | **0** |
| **Chloramphenicol** | **6 (6.0%)** | **3 (20.0%)** | **0** | **3 (60.0%)** | **0** | **0** | **0** | **0** | **0** |
| **Ciprofloxacin/ other fluoroquinolone** | **5 (5.0%)** | **1 (6.7%)** | **0** | **0** | **4 (5.1%)** | **43 (47.8%)** | **4 (80.0%)** | **31 (73.8%)** | **8 (18.6%)** |
| **Trimethoprim /Sulfamethoxazole** | **59 (59.0%)** | **8 (53.3%)** | **2 (100%)** | **1 (20.0%)** | **48 (61.5%)** | **6 (6.7%)** | **1 (20.0%)** | **5 (11.9%)** | **0** |
| **Gentamicin** | **12 (12.0%)** | **0** | **0** | **2 (40.0%)** | **10 (12.8%)** | **0** | **0** | **0** | **0** |
| Amoxicillin | 4 (4.0%) | 1 (6.7%) | 0 | 1 (20.0%) | 2 (2.6%) | 0 | 0 | 0 | 0 |
| Azithromycin | 0 | 0 | 0 | 0 | 0 | 4 (4.4%) | 0 | 4 (9.5%) | 0 |
| Erythromycin | 1 (1.0%) | 0 | 0 | 0 | 1 (1.3%) | 1 (1.1%) | 0 | 1 (2.4%) | 0 |
| Penicillin | 9 (9.0%) | 0 | 0 | 2 (40.0%) | 7 (9.0%) | 0 | 0 | 0 | 0 |
| Selexid / Pivmecillinam | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Other macrolides | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |

a Bold,antimicrobial agents that were tested for susceptibility

**b** Total number of Moderate-to-Severe Diarrhea (MSD) cases who were prescribed or given any antimicrobial agent at any participating SHF

|  |  |  |  |
| --- | --- | --- | --- |
| **Table 4. The sequence types (ST) of Typhimurium isolates identified in GEMS stools were determined using MLST PCR and/or whole genome sequencing** | | | |
| Site | Sequence type | | Total |
|  | ST36 | ST313 |  |
| **Africa** | **0** | **74** | **74** |
| Kenya | 0 | 74 | 74 |
| **Asia** | **9** | **4** | **13** |
| Bangladesh | 4 | 0 | 4 |
| Pakistan | 3 | 4 | 7 |
| India | 2 | 0 | 2 |
| **All sites** | **9** | **78** | **87** |

**Figure legends**

**Figure 1.** Distribution of *Salmonella* serogroups and serovars isolated from MSD cases and diarrhea-free asymptomatic controls. *Salmonella* spp. isolated from stools at A) all seven GEMS sites, B) Africa and C) Asia.

**Figure 2.** Percent of NTS non-susceptible to any of six commonly used antimicrobial agents. NTS isolated from A) Africa and B) Asia. NTS, non-typhoidal *Salmonell*a; CIP, ciprofloxacin; CRO, ceftriaxone; GEN, gentamicin; CHL, chloramphenicol; AMP, ampicillin; TMP/SMX, trimethoprim/sulfamethoxazole; MDR, multidrug resistant.

**Figure 3**. Percent of NTS that were non-susceptible to six antibiotics by serotype or serogroup. Only serotypes or serogroups that showed non-susceptibility to antibiotics are shown. NTS, non-typhoidal *Salmonell*a; CIP, ciprofloxacin; CRO, ceftriaxone; GEN, gentamicin; CHL, chloramphenicol; AMP, ampicillin; TMP/SMX, trimethoprim/sulfamethoxazole; MDR, multidrug resistant.

**Figure 4.** Genetic relationship between *Salmonella* Typhimurium ST313 isolated from MSD cases and diarrhoea-free controls. The core genome maximum likelihood tree is shown for *Salmonella* Typhimurium ST313 isolated from the stool of cases and controls of children aged under 5 years in Kenya and Pakistan (one isolate) which were collected as part of the GEMS study. Scale bars in Single Nucleotide Polymorphisms (SNPs) are shown beneath the phylogeny. Patient group, age range and antimicrobial resistance (AMR) data for the isolates are displayed using color strips created on ITOL and are labelled and coloured according to the inlaid key. Isolates which cluster with the lineage 1 or lineage 2 reference genomes are indicated. The tree is rooted using ST19.

**Supplementary files**

**Supplementary text.** Supplementary methods.

**Supplementary Table 1.** Meta-data of all *Salmonella* isolates from GEMS, including antibiotic susceptibility data and accession numbers of the 235 genomes that met quality control standards.

**Supplementary Table 2.** Odds ratios for various *Salmonella* serovars identified in stools.