SARS-CoV-2 neutralising antibodies in dogs and cats in the United Kingdom

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Abstract

Companion animals are susceptible to SARS-CoV-2 infection and sporadic cases of pet infections have occurred in the United Kingdom. Here we present the first large-scale serological survey of SARS-CoV-2 neutralising antibodies in dogs and cats in the UK. Results are reported for 688 sera (454 canine, 234 feline) collected by a large veterinary diagnostic laboratory for routine haematology during three time periods; pre-COVID-19 (January 2020), during the first wave of UK human infections (April-May 2020) and during the second wave of UK human infections (September 2020-February 2021). Both pre-COVID-19 sera and those from the first wave tested negative. However, in sera collected during the second wave, 1.4% (n=4) of dogs and 2.2% (n=2) of cats tested positive for neutralising antibodies. The low numbers of animals testing positive suggests pet animals are unlikely to be a major reservoir for human infection in the UK. However, continued surveillance of in-contact susceptible animals should be performed as part of ongoing population health surveillance initiatives.

Key words

SARS-CoV-2, serology, dogs, cats, animal disease surveillance

1. Introduction

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) emerged in Wuhan, China at the end of 2019 [1] and rapidly spread around the world. The main route of transmission remains human-to-human. However, there is evidence that the virus can infect animals [2] and it is important that we remain vigilant of such infections; particularly in companion animals with whom humans often have close contact.

Although initially there were only sporadic cases of infection in cats and dogs [3-5], there are now numerous reports of infection detected by RT-PCR or virus isolation [6-10], including in the UK [11]. Evidence of infection of cats and dogs has also been provided by the detection of anti-SARS-CoV-2 antibodies in several studies worldwide [12-17]. Experimental infections have shown that cats and, to a lesser extent, dogs are susceptible to SARS-CoV-2 and that cats can transmit the virus to other cats [18-20]. Infections in companion animals appear to have occurred as a result of human-to-animal transmission; however, the reported transmission of SARS-CoV-2 from farmed mink to in-contact humans, cats and dogs [21, 22] and the detection of the virus in stray dogs and cats [23, 24], suggest it is important to continue surveillance in companion animals. Here we conducted a survey of SARS-CoV-2 neutralising antibodies in cats and dogs attending UK veterinary practices.

1. Methods
	1. Samples

Canine and feline sera used in this study were obtained from the UK Virtual Biobank, which uses health data from commercial diagnostic laboratories participating in the Small Animal Veterinary Surveillance Network (SAVSNET) to target left over diagnostic samples in the same laboratories for enhanced phenotypic and genomic analyses [25]. All samples were residual sera remaining after routine diagnostic testing and were sent by one contributing laboratory based on convenience within the following parameters: samples were requested from UK cats and dogs collected over two time periods; March and April 2020 (during the first wave in humans) for both cats and dogs, then September 2020 to February 2021 for dogs, and January 2021 for cats (during the second wave in humans). Serum samples collected from the same laboratory in early January 2020 were also tested as pre-COVID-19 controls. All samples were linked to electronic health data for that sample (species, breed, sex, postcode of the submitting veterinary practice, date received by the diagnostic laboratory) held in the SAVSNET database, using a unique anonymised identifier. Data on SARS-CoV-2 exposure or symptoms was not available. Ethical approval to collect electronic health data (SAVSNET) and physical samples from participating laboratories (National Virtual Biobank) was granted by the Research Ethics Committee at the University of Liverpool (RETH000964).

2.2. Neutralising antibody detection in serum samples

Serum samples were screened for SARS-CoV-2 neutralising antibodies using the plaque reduction neutralisation test (PRNT) as previously described [15], with the SARS-CoV-2/human/Liverpool/REMRQ0001/2020 isolate cultured in Vero E6 cells [26]. Briefly, sera were heat inactivated at 56°C for 30 mins and stored at -20°C until use. DMEM containing 2% FBS was used to dilute sera ten-fold followed by serial two-fold dilution. SARS-CoV-2 at 800 plaque forming units (PFU)/ml was added to diluted sera and incubated at 37°C for 1 h. The virus/serum mixture was then inoculated onto Vero E6 cells, incubated at 37°C for 1 h, and overlaid as in standard plaque assays [27]. Cells were incubated for 48 h at 37°C and 5% CO2, ﬁxed with 10% formalin and stained with 0.05% crystal violet solution. PRNT80 was determined by the highest dilution with 80% reduction in plaques compared to the control. Samples with detectable neutralising antibody titre were repeated as technical replicates for conﬁrmation. Where titres differed between technical replicates, the lowest dilution was reported.

1. Results

A total of 732 samples were received from the diagnostic laboratory and tested for SARS-CoV-2 neutralising antibodies. Linking of data to the samples found that 22 samples were duplicates (duplicate samples gave the same result in each replicate and are therefore reported as one sample). Seven samples were from animals with non-UK postcodes, two samples did not have species data, two samples were received as dogs but were actually from cats and were collected outside the two time periods of cat sample collection and eleven samples were missing postcodes; these samples were excluded. Results are therefore reported for 688 sera (454 canine, 234 feline) of which 558 (372 dogs, 186 cats) were collected during the SARS-CoV-2 pandemic and 130 (82 dogs, 48 cats) were collected from animals before the first confirmed human case in the UK (21st January 2020 [28]) - pre-COVID-19 samples; these samples were distributed across the UK (Figure 1). Of the dog sera collected during the pandemic, 0/85 (0%) collected in March/April 2020 and 4/287 (1.4%) collected September 2020-February 2021 tested positive for neutralising antibodies with titres of 1:20 (n=2) and 1:80 (n=2). In cats, 0/96 (0%) sera collected in March/April 2020 tested positive for neutralising antibodies and 2/90 (2.2%) collected in January 2021 tested positive with titres of 1:40 and 1:80. Pre-COVID-19 sera from both dogs (n=82) and cats (n=48) tested negative for neutralising antibodies. Positive samples in dogs were collected in November 2020 (n=1), January 2021 (n=2) and February 2021 (n=1) and were collected in Kent, Buckinghamshire, Worcestershire and Yorkshire, respectively (Figure 1). The two positive cats were collected in January 2021; one in Birmingham and the other in London (Figure 1).

1. Discussion

Studies of companion animals have shown that they can be infected with SARS-CoV-2 [2]. In the UK, there have been sporadic reports of infection in cats and dogs, including one that tested 387 cats showing respiratory signs using RT-PCR and found one cat to be positive for the virus [11, 29]. However, there has been no large scale sero-survey of infection. Here we show that a small proportion of UK dogs and cats, sampled at a time of active human transmission, tested positive for SARS-CoV-2 neutralising antibodies.

Sera from two time points during the pandemic were analysed. Sera collected during March and April 2020 (first wave) from both cats and dogs were negative for neutralising antibodies. Previous studies using European samples have shown a low level of infection, highest in Italy, where 3.3% (15/451) of dog sera and 5.8% (11/191) cat sera collected between March and May 2020 had measurable neutralising antibody titres [15]. These samples were purposefully collected from regions of Italy with a high prevalence of infection in humans, in some cases from households known to contain recently diagnosed human cases. Our results in contrast, are more consistent with a survey from a similar population of cats in Germany, that found 0/221 samples collected in April and May of 2020 to be positive for anti-SARS-CoV-2 antibodies using ELISA [13], and with a survey in the Netherlands in April-May 2020, that found 0.4% of cats and 0.2% of dogs to be seropositive [30]. The lack of positive samples from March-April 2020 in the UK, that we report here, likely reflects the selection criteria of the animals assayed (undergoing routine haematological testing and not selected based on location), and the relatively low rate of human disease at the time compared to Italy.

In sera collected during the second wave of the pandemic, 4/287 (1.4%) dogs and 2/90 (2.2%) cats tested positive for SARS-CoV-2 neutralising antibodies. Positive dog samples were collected in November 2020 and January and February of 2021. Positive cat samples were collected in January 2021. This is again broadly in line with a recent German survey conducted from September 2020 to February 2021, showing a seroprevalence of 1.36% (increased from 0.69% earlier in the pandemic), that the authors concluded corresponded with the rise of reported cases in the human population, and was suggestive of ongoing transmission from owners to their cats [14]. The detection of neutralising antibodies in dogs and cats during the second wave in this study also likely reflects the increased numbers of humans that had been infected. However, it is unknown whether the main variant circulating in the UK human population during the second wave (B.1.1.7) has altered transmissibility to cats and dogs as it does in humans [31]. Whilst there is some work in model species testing new variants in e.g. hamsters [32, 33], to our knowledge none of the early experimental infections performed in cats and dogs using the original SARS-CoV-2 have been repeated with newer variants.

Cats and dogs can be infected with other coronaviruses, leading to the possibility that SARS-CoV-2 neutralising antibodies detected in this study may result from previous infection with a different virus. We and others have previously demonstrated a lack of cross-reactivity between SARS-CoV-2 and samples containing antibodies to feline coronavirus (FCoV), canine enteric coronavirus (CeCoV) and canine respiratory coronavirus (CRCoV) [13, 15-17]. The reverse has also been demonstrated whereby cat or dog samples containing antibodies to SARS-CoV-2 did not react with known feline or canine coronavirus antigens [13, 34]. Here we also tested samples from UK cats and dogs collected before the human index case in the UK (21st January 2020 [28]). All pre-COVID-19 samples were negative for SARS-CoV-2 neutralising antibodies. Although there are no recent sero-surveys of cat and dog coronaviruses in the UK [35-37], FCoV, CeCoV and CRCoV are considered to be endemic. Furthermore, a recent serological survey in the Netherlands testing for anti-SARS-CoV-2 antibodies also found 71% of cat and 40% of dog pandemic sera to have antibodies to other known cat or dog coronaviruses [30]. If cross-reactivity of SARS-CoV-2 with antibodies to cat and dog coronaviruses was occurring in the study reported here, this would be apparent in the pre-COVID-19 samples. We would also expect to see higher numbers of animals with SARS-CoV-2 neutralising antibodies in the pandemic samples, again suggesting that antibodies produced following infection by endemic cat and dog coronaviruses do not cross-react with SARS-CoV-2.

Here we made use of samples collected from a commercial diagnostic laboratory contributing data to a voluntary national surveillance scheme (SAVSNET) to efficiently test for evidence of prior SARS-CoV-2 infection in UK cats and dogs. The major limitations of such a system are the relatively sparse data available for each sample such that individual animals, that are not identifiable, may have been sampled twice or have come from the same household. In addition, such samples lack detailed information on the health of the animals and whether they were from a COVID-19-positive household. However, acquiring such samples from the UK Virtual Biobank, offers a responsive resource for studying national patterns of disease in UK pets [25].

We report here the detection of a low level of SARS-CoV-2 neutralising antibodies in dogs and cats during the second wave of human infections in the UK. The use of a neutralisation assay will not detect animals that have sero-converted but not produced neutralising antibodies (such that might test positive by ELISA but not by PRNT). Indeed, other studies have shown that sera positive by ELISA for anti-SARS-CoV-2 antibodies may not have detectable neutralising antibodies [13, 34]. As such, our results may underestimate the number of animals exposed to SARS-CoV-2.

We and others report that cats and dogs can become infected with SARS-CoV-2, likely through their interactions with humans. Although animal-to-animal transmission has been reported, for example on mink farms and in experimental infections [18-20, 22, 38], the small numbers of companion animals testing positive in the field in the UK and elsewhere suggest that pets are not currently acting as a significant reservoir for infection, and that the pandemic will be controlled by measures largely focussed on minimising human-to-human transmission. Current UK guidance is that COVID-19 positive humans should limit contact with their pets. Studies like that presented here strongly argue for continued surveillance of in-contact, susceptible animal species, particularly with the emergence of new variants, which will help determine whether in the future, more targeted control measures are needed for pet animals, particularly in regions that are gaining control of infection in their human populations.

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Conflict of interest

NM and LR are employed by IDEXX Laboratories. All other authors declare no competing interests.

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Figure 1: Schematic map showing the location of samples for which testing of SARS-CoV-2 neutralising antibodies is reported. Red dots indicate samples that were positive for SARS-CoV-2 neutralising antibodies using PRNT80. Blue dots indicate samples that were negative.