**Advances in the Pharmaceutical Treatment Options for Canine Osteoarthritis**

**Abstract**

Canine osteoarthritis is a significant cause of pain in many dogs and can therefore compromise animal welfare. As the understanding of the biology and pain mechanisms underpinning osteoarthritis grows, so do the number of treatments available to manage it. Over the last decade, there have been a number of advances in the pharmaceutical treatment options available for dogs with osteoarthritis, as well as an increasing number of clinical trials investigating the efficacy of pre-existing treatments. This review aims to examine the current evidence behind pharmaceutical treatment options for canine osteoarthritis, including non-steroidal anti-inflammatory drugs, piprants, monoclonal antibodies, adjunctive analgesics, structure modifying osteoarthritis drugs and regenerative therapies.

**Introduction**

Osteoarthritis (OA) is a significant cause of pain, lameness and morbidity in dogs and many other species, including humans, across the world (Brown et al., 2013b, Knazovicky et al., 2016, Anderson et al., 2018, Cui et al., 2020). It is a multifactorial, progressive, degenerative disease of synovial joints, affecting not only the articular cartilage, but other structures within the specific synovial joint (Loeser *et al*., 2012). Degradation of articular cartilage, subchondral bone sclerosis, osteophytosis, varying degrees of synovitis, meniscal and ligament degeneration are all characteristics of the disease process. However, there is still much to be understood regarding the underlying pathogenesis of OA (Glyn-Jones *et al*., 2015).

Canine OA most commonly arises as a result of inciting factors, such as coxofemoral (hip) joint dysplasia, elbow dysplasia, cranial cruciate ligament (CCL) disease, patella luxation, limb malformations and articular fractures (Johnston, 1997). A number of risk factors for canine OA have been identified, including genetic predispositions, diet and obesity, all of which can play a role in disease progression (Anderson *et al*., 2018).

Estimates of the prevalence of dogs presenting with OA to primary care veterinary practices in the UK vary depending on the study, but have recently been reported to be between 2.5% (Anderson *et al*., 2018) and 6.6% (O'Neill *et al*., 2014b). A previous study of dogs attending referral hospitals in the USA reported an estimated prevalence of OA of up to 20% in dogs over one year old (Johnston, 1997). The true prevalence of the disease, however, is likely to be higher when unreported cases and the discrepancies in recording systems are taken into consideration (O'Neill *et al*., 2014a). With an estimated UK canine population of around 12.5 million (Pet Food Manufacturers Association, 2021) and 77 million in the USA (American Veterinary Medical Association, 2017), this represents a significant number of affected dogs, as well as a substantial number of dog owners and caregivers charged with the responsibility (and economic cost) of managing their treatment (Belshaw *et al*., 2020).

As OA therefore presents a welfare problem for many dogs, it is important that information regarding treatment options for these animals is both up to date and evidence based, aiding veterinary practitioners and dog owners in effectively managing these cases. With a plethora of treatment options available, and novel drugs having gained market authorisation for the treatment of canine OA in the last decade, the aim of this review is to examine the recent evidence in the literature underpinning the pharmaceutical treatment options for the management of canine OA. In this review, pharmaceutical treatments of canine OA will be discussed, focussing on novel therapies, and updated evidence for existing treatments.

Some of the studies referenced in this review compare pharmaceutical treatments of OA to a placebo. It should be noted that using a placebo as a control in a disease that is known to be painful and for which there are licensed treatments available has ethical implications. In the UK, these would, as a minimum, require an Animal Test Certificate from the Veterinary Medicines Directorate or may require authorisation under the Animal (Scientific Procedures) Act 1986 depending on the nature of the study (Veterinary Medicine Directorate, 2018). As an alternative to placebo controlled trials, some studies use a medication with known efficacy as a positive control to compare to the treatment group (Reymond *et al*., 2012). In these cases the term “non-inferior” is used if the treatment group is not worse than the positive control (Freise *et al*., 2013).

Licensing guidelines for the UK market will be used as standard unless stated otherwise. Veterinary practitioners should also be aware of non-pharmaceutical therapies in the non-surgical management of canine OA, such as weight management and physiotherapy, enabling a multi-modal treatment approach to this disease.

**Pain Mechanisms in OA**

OA is a painful chronic disease. The pain mechanisms involved are complex (Fu *et al*., 2018). Both peripheral components of pain and central processes are involved, with nociceptive, inflammatory and neuropathic types of pain occurring to varying degrees (White and Hunt, 2019). There are several classes of analgesic medications available, with different mechanisms of action, targeting nociception at different steps along the pain pathway (Figure 1). With this in mind, it is important to consider multi-modal analgesia and management when treating canine OA patients if an insufficient response to one type of medication is shown (Lascelles *et al*., 2008).

**Pharmaceutical Treatments of Canine OA**

Non-Steroidal Anti-Inflammatory Drugs (NSAIDs)

Non-steroidal anti-inflammatory drugs (NSAIDs) have been first-line analgesics for the management of canine OA pain for many years. Conventional NSAIDs exert their analgesic effects by inhibiting the enzyme cyclooxygenase (COX), which is responsible for the production of prostaglandins from arachidonic acid. There are two peripheral isoforms of COX: COX-1 and COX-2. Unwanted side effects caused by the inhibition of COX-1, such as gastrointestinal and renal effects, has led to the development of preferential and selective COX-2 inhibitors (KuKanich *et al*., 2012). The mechanism of action of NSAIDs is outlined in Figure 2.

The efficacy of NSAIDs in reducing pain related to OA has been well documented. Of the NSAIDs licensed for veterinary use, previous systematic reviews of treatments for canine OA have shown a high level of evidence in the literature for the efficacy of meloxicam, carprofen and firocoxib (Sandersoln *et al*., 2009, Aragon *et al*., 2007). Since the publication of these systematic reviews, four other coxibs have been licensed in the UK for the treatment of OA in dogs, namely robenacoxib (Onsior®; Elanco), mavacoxib (Trocoxil®; Zoetis), cimicoxib (Cimalgex®; Vetoquinol) and enflicoxib (Daxocox®; Animalcare Limited). Similar to firocoxib, these NSAIDs are highly selective inhibitors of COX-2, therefore theoretically reducing the risk of adverse side effects caused by COX-1 inhibition (Toutain *et al*., 2018). In humans, coxibs have been shown to increase the risk of myocardial infarction, leading to some products losing their market authorisation (Arora *et al*., 2020). However, this has not been to date demonstrated in dogs. Due to a lack of sufficiently powered, randomised, blinded, placebo controlled studies when comparing the incidence of adverse effects and analgesic efficacy of different NSAIDs, it is not known whether selective COX-2 inhibitors are superior to preferential COX-2 inhibitors (Monteiro-Steagall *et al*., 2013).

A summary of recent clinical trials involving NSAIDs licensed for use in canine OA is shown in Table 1. One study demonstrated a similar efficacy in reducing pain related to OA with robenacoxib compared to carprofen over a 12 week treatment period in 188 dogs (Reymond *et al*., 2012). Clinical trials involving cimicoxib for the treatment of OA in dogs are lacking. However, one study showed cimicoxib to provide analgesia superior to tramadol post-operatively after tibial plateau levelling osteotomy (TPLO) surgery (Piras *et al*., 2021). Enflicoxib, the most recently licensed of the coxibs, is dosed at weekly intervals, with an initial loading dose of 8mg/kg, followed by once weekly doses of 4mg/kg (National Office of Animal Health, 2021a). A recent study involving 242 dogs randomised to receive either enflicoxib, mavacoxib or placebo, found an improvement in veterinary and owner assessment of clinical signs related to OA with both NSAIDs compared to placebo over the six-week trial period (Salichs *et al*., 2021). Mavacoxib differs from the other licensed NSAIDs, as it has a much longer half-life, and the dose regime involves repeating the initial oral dose 14 days after the first treatment, then at monthly intervals (Lees *et al*., 2015). Two clinical trials examined the clinical efficacy of mavacoxib treatment in 111 and 124 dogs with naturally occurring OA compared to either meloxicam (Walton *et al*., 2014) or carprofen (Payne-Johnson *et al*., 2015) respectively. Both studies found mavacoxib to be non-inferior to the control NSAID over the study periods of 12 weeks and 134 days with a similar rate of adverse events. Due to its prolonged half-life however, there is a potential for increased risk of inadvertent overdose by owners, and an inability to cease daily dosing if unwanted side effects do occur (European Medicines Agency, 2008).

All NSAIDs have potential side effects, the most common of which include gastrointestinal signs such as vomiting, diarrhoea and inappetence, with severe side effects, including gastrointestinal ulceration and renal toxicity, being noted on datasheets of medications to happen rarely to very rarely. The true overall incidence of adverse effects, and whether there is a significant difference between licensed NSAIDs in terms of safety, is unknown (Monteiro-Steagall *et al*., 2013, Hunt *et al*., 2015).

**In summary, NSAIDs provide analgesia for dogs with OA pain, and although there are many licensed for use in dogs with OA, no one licensed NSAID has been shown to be consistently superior to another in terms of efficacy or safety.**

Piprants

Grapiprant

Recently, a novel class of non-steroidal, non-COX inhibiting drugs, the piprants, have been pursued as treatment options for the management of canine OA. This has led to the development of a prostaglandin E₂ (PGE₂) EP4 receptor antagonist (PRA), grapiprant, as a licensed treatment for dogs with mild to moderate OA pain (Galliprant®; Elanco).

EP4 is a receptor through which PGE₂, a key mediator of inflammation and pain, exerts its effects (Nakao *et al*., 2007). By antagonising EP4, grapiprant blocks PGE2 mediated sensitisation of sensory neurons and PGE2 mediated inflammation, thus producing anti-inflammatory and analgesic effects without inhibiting the production of prostaglandins as a whole (Lin *et al*., 2006) (Figure 2). Therefore, grapiprant could *theoretically* reduce the risk of side effects caused by the inhibition of COX enzymes (Kirkby Shaw *et al*., 2016).

Only one peer reviewed clinical trial involving grapiprant use in dogs for the treatment of naturally occurring OA pain has been published to date. This prospective, randomised, blinded, placebo-controlled study involving 265 client owned dogs with OA confirmed by radiography in at least one appendicular joint examined the clinical effects of grapiprant (2mg/kg once a day *per os*) compared to a placebo over the course of 28 days treatment (Rausch-Derra *et al*., 2016). Outcomes were measured using a clinical metrology instrument, the Canine Brief Pain Inventory (CBPI) (Brown *et al*., 2008) and veterinary assessments, with safety measured by physical examination, clinical pathology results and owner observations. Grapiprant was generally well tolerated, and the authors found grapiprant to have improved pain scores compared to placebo, with a treatment success rate of 48.1% for grapiprant treated dogs compared to 31.3% for dogs receiving a placebo (*p*=0.0315). These measurements of treatment success were based on a previous definition by Brown et al. (2013a), using the CBPI and an overall impression score of the same or better on day 28 compared to day 0 . Studies comparing the analgesic efficacy of grapiprant to NSAIDs have so far only examined acute pain control for the 24 to 48-hour period following experimental induction of arthritis or synovitis. Both studies found the NSAID (firocoxib or carprofen) to have superior analgesic properties in that timeframe (de Salazar Alcalá *et al*., 2019, Budsberg *et al*., 2019) (Table 1). However, it should be noted that this is a very short timeframe for treatment and is in an acute pain experimental model of induced OA rather that a clinical setting. Further studies comparing the efficacy of grapiprant to that of NSAIDs would be of use in improving the evidence behind its use and as a first line treatment for OA pain in dogs.

Potential side effects of grapiprant include vomiting (very common [more than one in ten animals treated]), diarrhoea and inappetence (common [between one and ten animals in 100 animals treated]), with these side effects being generally mild and transient. In very rare cases (less than one in 10,000 animals treated), haemorrhagic diarrhoea or haematemesis has been reported (National Office of Animal Health, 2021b). The recommended daily dose of grapiprant is 2mg/kg once a day. In a study examining the safety of daily grapiprant administration at doses up to 50mg/kg *per os* once a day for 9 months in healthy Beagles in an experimental setting, it was well tolerated with no renal or hepatic toxicity noted suggesting the safety of long-term oral administration of grapiprant to dogs (Rausch-Derra *et al*., 2015).

**Piprants are a new class of drugs that offer a more targeted mechanism of action than COX-inhibiting NSAIDs (Kirkby Shaw *et al*., 2016). However, their clinical efficacy in the long-term treatment of canine OA compared to NSAIDs is, as yet, unknown, and larger clinical trials comparing these drugs would improve the evidence.**

Paracetamol and Paracetamol/Codeine

Paracetamol (acetaminophen) is an analgesic and anti-pyretic, and is a common first line treatment for OA pain in humans (Onakpoya, 2020). It has a complex mechanism of action which is still not completely understood, functioning as both an inhibitor of COX peripherally and centrally, as well as acting on other central anti-nociception pathways such as serotonergic pathways, the endocannabinoid system and the L-arginine/NO pathway (Przybyła *et al*., 2021). Paracetamol on its own is not licensed for use in dogs in the UK, but a preparation of paracetamol and codeine phosphate (400mg Paracetamol/9mg Codeine Phosphate) is licensed in the UK for the treatment of acute pain of traumatic origin, as a complementary treatment in pain associated with other conditions, and for post-operative analgesia (Pardale-V®, Dechra Veterinary Products). This is licensed for a treatment duration of up to five days. Codeine, an opioid, has a low oral bioavailability in dogs and it is unknown whether it effectively contributes to the analgesic effects of this product (KuKanich, 2010).

In humans, recent Osteoarthritis Research Society International (OARSI) treatment guidelines no longer recommend the use of paracetamol as a single-agent in the treatment of knee, hip and polyarticular OA (Bannuru *et al*., 2019). Previously, the use of paracetamol had been advised as a first line treatment, but evidence in recent meta-analyses of human trials show it to have little to no efficacy in the treatment of OA in humans (Machado *et al*., 2015, da Costa *et al*., 2017, Bannuru *et al*., 2019). In canines, there are no published studies examining the analgesic efficacy of paracetamol alone, in combination with codeine, or as an adjunctive analgesic with other medications such as NSAIDs in dogs with chronic OA pain. This is a gap in the current evidence base. In practice, paracetamol can be used (off license) concurrently with an NSAID as an adjunctive analgesia, or as an alternative to an NSAID in those dogs where NSAID use is not tolerated (Pettitt and German, 2015).

**There is no published data on the analgesic efficacy of paracetamol in canine OA specifically, and use of Pardale-V® (Dechra) of more than five days duration, or concurrently with an NSAID, is off licence. This highlights a gap in the current evidence base of pharmaceutical treatments for canine OA.**

Anti-Nerve Growth Factor Monoclonal Antibodies

Nerve growth factor (NGF) is a soluble signalling protein, released from peripheral tissues in response to noxious stimuli. It has an important role in nociceptor sensitisation in both acute and chronic pain states, including OA, by increasing peripheral sensitisation through phenotypic alterations, increasing the expression of pro-nociceptive neurotransmitters and inducing inflammatory mediator release in the periphery (Enomoto *et al*., 2019).

Bedinvetmab

In 2020, a canine-specific anti-NGF mAb product, bedinvetmab (Librela®; Zoetis), received approval from the European Medicines Agency. This product is licensed in the UK as a once monthly subcutaneous injection for dogs over 12 months of age, at a dose of 0.5 to 1.0mg/kg.

In a double blinded, placebo controlled, multicentre, randomised controlled trial, Corral *et al*. (2021) investigated the efficacy of monthly bedinvetmab injections in 287 dogs with OA. The study had a three-month comparative phase, where CBPI scores were used as primary outcomes in one group treated with bedinvetmab, compared to another group given a placebo. Improved CBPI scores where observed in all assessed time points in this trial in the bedinvetmab treated groups compared to placebo. The study then had a continuation phase, involving 89 dogs that showed clinical improvement with bedinvetmab treatment continuing with monthly injections of bedinvetmab for a further six months. Sustained efficacy was shown over this timeframe in terms of CBPI scores, although the study lacked objective outcome measurements such as force plate analysis.

Concurrent use of anti-NGF mAbs with NSAIDs in humans in clinical trials have shown a rapid progression of OA (Hefti, 2020). The concurrent use of bedinvetmab with an NSAID (carprofen) has only been investigated in young, healthy laboratory dogs without OA for a period of two weeks (Krautmann *et al*., 2021), and the long-term concurrent use of these medications in dogs with OA has not been investigated. Therefore, it cannot be currently recommended to administer bedinvetmab and an NSAID concurrently. The only noted adverse reaction on the datasheet for Librela®   
(Zoetis) is an uncommon mild reaction at the injection site (National Office of Animal Health, 2021c).

**Initial evidence for the effectiveness of anti-NGF mAbs shows promise in providing an alternative treatment option for canine OA (Webster *et al*., 2014, Lascelles *et al*., 2015, Corral *et al*., 2021), with bedinvetmab licensed for the alleviation of pain associated with OA in dogs.**

Opioids

Tramadol

Tramadol has been used as an analgesic in humans since the 1970s (Schenck and Arend, 1978) and exerts its analgesic effects via several mechanisms. It is a mu-opioid agonist and a serotonin and noradrenaline reuptake inhibitor (Grond and Sablotzki, 2004). The effects on the mu-opioid receptor are predominantly due to the tramadol metabolites, especially the O-desmethyltramadol (M1) metabolite. Some dogs (and it is unknown what proportion) are unable to produce this metabolite, hence reducing the mu opioid analgesic effects in some canines (Perez Jimenez *et al*., 2016, KuKanich and Papich, 2004).

Since 2018, there are now two licensed tramadol hydrochloride products available for use in the reduction of acute and chronic mild soft tissue and musculoskeletal pain in dogs in the UK (Tralieve®; Dechra and Tramvetol®; Virbac).

The evidence in the literature supporting the use of tramadol as an analgesic in the treatment of canine OA describes mixed results. Budsberg *et al*. (2018) compared subjective and objective outcome measurements in 40 dogs with OA when treated with either carprofen, tramadol or a placebo for 10 days. A significant improvement in all outcome measures were found with carprofen treatment, but not with placebo or tramadol. The authors concluded that tramadol provided no clinical benefit in the treatment of canine OA, however the time period of the study was short, and the number of dogs in each treatment group was small. Malek *et al*. (2012) however, found an improvement in owner assessed mobility scores in dogs treated with either tramadol or carprofen compared to ABT-116 (a transient receptor potential vanilloid 1 [TRPV1] antagonist) or placebo. However, no difference were found between groups on objective kinetic gait analysis. This study involved 49 dogs with OA treated for two weeks.

Two studies have examined concurrent use of tramadol with an NSAID compared to treatment with an NSAID alone (Monteiro *et al*., 2019, Miles *et al*., 2020). Monteiro *et al*. (2019) found an increased analgesic efficacy in 20 dogs when tramadol was used in conjunction with reduced dose ketoprofen over 28 days. Miles *et al*. (2020) also found an improvement in objective measurements of lameness such as force plate gait analysis in a group of 18 dogs receiving both tramadol and an NSAID for 28 days compared with NSAID as a sole agent.

**With a limited number of clinical studies investigating the efficacy of tramadol for the treatment of canine OA pain, and the known individual variability of the production of the M1 metabolite in dogs (KuKanich and Papich, 2004), it is difficult to draw firm conclusions over its clinical effectiveness. Budsberg *et al*. (2018) suggested that when tramadol was used as a sole analgesic for periods of less than two weeks it provided insufficient analgesia for OA pain. However, there may be a beneficial role when used in conjunction with other analgesics as part of a multi-modal approach to pain management in some dogs (Monteiro *et al*., 2019).**

Gabapentinoids

Gabapentin and Pregabalin

Gabapentin is a synthetic analogue of γ-aminobutyric acid (GABA), and is used in human medicine as an anti-epileptic drug, and in the treatment of chronic neuropathic pain and fibromyalgia (Calandre *et al*., 2016). Its mechanism of action is not fully understood, but it is believed to exert its main effects by selectively inhibiting voltage-gated calcium channels containing the alpha2delta-1 subunit, leading to reduced neurotransmitter release and lessening of post-synaptic excitability (Sills, 2006). Pregabalin has a similar mechanism of action to gabapentin, but it has a longer half-life and higher oral bioavailability (Salazar *et al*., 2009).

Previous pharmacokinetic studies have suggested a dosage of gabapentin in dogs of 10-20mg/kg *per os* every eight hours (Kukanich and Cohen, 2011) compared to a dosage of pregabalin of 4mg/kg every 12 hours to achieve the therapeutic plasma concentration seen in humans (Salazar *et al*., 2009). The therapeutic plasma concentrations in dogs are unknown and there are currently no licensed products of either gabapentin or pregabalin available for veterinary use in the UK.

There are a limited number of published studies involving the use of gabapentin and pregabalin in dogs, and only one published abstract describing a study examining the use of gabapentin as an adjunctive treatment with NSAIDs in canine OA (Miles *et al*., 2020). This study involved objective measurements of gait analysis of a small cohort of 24 dogs with OA, receiving either tramadol or gabapentin for four weeks in addition to an NSAID. Both tramadol and gabapentin led to an improvement in weight bearing (Miles *et al*., 2020).

**The evidence for the use of gabapentinoids in canine OA is currently lacking. More published high-quality clinical trials are needed to examine the efficacy of gabapentin and pregabalin in the dog, in order to give a greater evidence-base behind their usage.**

*N*‐Methyl d‐Aspartate (NMDA) Receptor Antagonists

Amantadine and Memantine

Amantadine, first developed as an anti-viral medication and also used to treat Parkinson’s disease in humans, exerts its analgesic effects by antagonising *N*‐methyl d‐aspartate (NMDA) receptors (Fisher *et al*., 2000). Memantine is also an NMDA receptor antagonist, and is a more potent congener of amantadine (Johnson and Kotermanski, 2006). Although neither drug is licensed in dogs, suggested doses of amantadine are 3.0 to 5.0 mg/kg *per os* once a day. A study examining the use of memantine in the treatment of compulsive disorders in dogs suggested doses of memantine of 0.3 to 0.5mg/kg *per os* twice a day initially, increasing to 1.0mg/kg twice a day if necessary (Schneider *et al*., 2009).

There is only one published study examining the use of amantadine in dogs with OA (Lascelles *et al*., 2008). This study showed an improvement in subjective veterinary assessment of OA pain, and in the Client Specific Outcome Measure (CSOM) clinical metrology instrument (CMI) in 31 dogs when amantadine was given in conjunction with meloxicam, compared to meloxicam with placebo.

**NMDA antagonists could be beneficial as adjunctive analgesics in managing chronic OA pain (Lascelles and other 2008), however this was found in one small study, and larger clinical trials are required to improve the evidence behind their use. There are no published clinical trials involving the use of memantine in dogs with OA and both the use of amantadine and memantine are off licence in the UK.**

Cannabinoids

Cannabidiol (CBD oil)

Cannabinoids have gained attention in recent years for their potential efficacy as analgesics in patients with chronic pain. They have a complex mechanism of action, acting on peripheral, spinal and supra-spinal sites to exert anti-nociceptive and anti-hyperalgesic effects (Richardson, 2000, Landa *et al*., 2016).

Clinical trials investigating the use of CBD oil in dogs with OA have had mixed results, and are all of low sample size, involving up to a total of 32 dogs (Table 1). Two randomised crossover studies comparing CBD oil treatment to placebo found a beneficial effect in reduction of clinical signs by subjective measurements and validated CMIs over study periods of four weeks in 20 and 22 dogs respectively (Verrico *et al*., 2020, Gamble *et al*., 2018). However in another randomised, double-blinded, crossover study involving 23 dogs, Mejia *et al*. (2021) found no significant difference between CBD oil treatment and placebo over the course of six weeks, based on CMI outcomes and objective pressure gait analysis. Two other published studies have suggested an improvement in clinical signs of OA in dogs treated with CBD oil as part of a multimodal analgesic plan (Brioschi *et al*., 2020), and with gabapentin (Kogan *et al*., 2020). However, both studies have limitations. The latter study had no control group, and did not use validated pain scoring systems to subjectively measure outcomes (Kogan *et al*., 2020), and the former involved dogs already receiving an NSAID, gabapentin and amitriptyline (Brioschi *et al*., 2020). Both also involved small numbers of dogs, with sample sizes of 32 and 23 respectively.

**Currently, there is no licence for the use of CBD oil in dogs in the UK, and veterinary surgeons should be aware of the legal position regarding prescribing these medications. A recently published information sheet from the British Small Animal Veterinary Association (BSAVA) outlines the current position in the UK (Wessmann *et al*., 2016). There is currently very limited available evidence for the efficacy of CBD oil for the treatment of canine OA pain.**

Tricyclic Antidepressants

Amitriptyline

Amitriptyline is a tricyclic antidepressant (TCA), and commonly used as a treatment for neuropathic pain in humans. TCAs exert their analgesic effects by several mechanisms of action, including NMDA and adrenergic receptor antagonism, serotonin and noradrenaline reuptake inhibition, voltage-gated sodium channel blockade, and enhancement of adenosine and GABA B receptor activity (Dharmshaktu *et al*., 2012).

**There are currently no published clinical trials investigating the analgesic efficacy of TCAs in dogs with OA pain, and there is no licensed preparation of amitriptyline for dogs.**

Corticosteroids

Corticosteroids are thought to have an analgesic effect in OA due to their anti-inflammatory action (Johnston and Budsberg, 1997). Their use in OA management is controversial (Behrens and others 1975, Murphy and others 2000), and due to the well documented risks of long-term corticosteroid use they should be used with caution if given long term. Prednoleucotropin, an oral prednisolone and cinchophen combination product has been used for the treatment of OA in dogs (McKellar *et al*., 1991) but is no longer licensed in the UK.

Intra-articular (IA) injections of long-acting preparations of corticosteroids (methylprednisolone acetate, triamcinolone acetonide, and triamcinolone hexacetonide) in humans with OA, have been shown to have a short-term benefit in alleviation of pain compared to placebo (Najm *et al*., 2021). Methylprednisolone acetate is licensed for IA use in the dog for inflammatory conditions (Depo-medrone V; Zoetis), therefore its use in osteoarthritis (typically considered a non-inflammatory condition, although now understood to have an inflammatory component) under the licence is debatable (Sokolove and Lepus, 2013). Clinical trials into its effectiveness and safety in naturally occurring OA are lacking. A recent clinical trial found improvements in weight bearing for up to 90 days in a group of 20 dogs following a single IA injection of an unlicensed preparation of the corticosteroid triamcinolone hexacetonide in OA hip joints compared to a control group receiving a placebo (Alves *et al*., 2021b).

**Veterinary practitioners should consider the contraindications for corticosteroids, such a septic arthritis, and safety concerns over potential cartilage damage with long term IA corticosteroids before use (Chunekamrai *et al*., 1989, Farquhar *et al*., 1996, Murphy *et al*., 2000). Long term efficacy and safety studies of IA corticosteroid use in clinical trials in dogs are lacking.**

Candidate Structure Modifying OA Drugs

Several drugs have been investigated as potential structure modifying OA drugs (SMOADs). SMOADs are defined as drugs that can delay, stabilise or repair OA lesions in affected joints rather than just alleviating the symptoms of OA (Sevalla *et al*., 2000, Sunaga *et al*., 2012). However, no evidence currently exists that these drugs are able to stabilise or repair OA lesions *in vivo*. Examples of these medications include pentosan polysulphate (PPS), polysulphated glycosaminoglycans (PSGAGs), hyaluronic acid (HA), and doxycycline. Little new evidence has emerged regarding the use of these treatments in canine OA over the past decade.

Sodium PPS is licensed for treatment for OA in dogs in the UK (Osteopen; Chanelle Pharma, Cartrophen Vet; Arthropharm (Europe) Ltd). It has a wide range of pharmacological activities, including anti-catabolic activities in articular cartilage, anti-inflammatory actions, increasing hyaluronan production from synoviocytes and thrombolytic activity that could enhance blood supply to affected joints (Ghosh, 1999). Studies investigating the efficacy of PPS as a treatment for OA in dogs have shown improvements in outcomes compared to placebo, however they are limited and not all based upon objective outcomes (Bouck et al., 1995, Read et al., 1996, Innes et al., 2000, Smith et al., 2001, Budsberg et al., 2007).

The mechanisms by which polysulphated glycosaminoglycans (PSGAGs) is believed to be through the inhibition of matrix metalloprotease (MMP) enzymes, therefore having a preventative effect on matrix molecule degradation in articular cartilage (Sevalla et al., 2000). PSGAGs were found to have a moderate level of evidence in a previous systematic review based on two included randomised controlled trials, although the authors concluded that further studies were needed (Sandersoln et al., 2009).

Doxycycline has been investigated as a SMOAD due to *in vitro* studies suggesting a potential to slow cartilage degeneration (Shlopov et al., 1999). However, evidence behind its use as a treatment for canine OA in a clinical setting is limited, and previous systematic reviews have found no evidence for its use in the treatment of canine OA (Aragon et al., 2007, Sandersoln et al., 2009)**.**

A recent small cohort clinical trial in dogs with naturally occuring OA secondary to hip dysplasia, compared eight dogs treated with IA injections of a low molecular weight HA with a control group receiving saline IA injections plus oral carprofen and a nutraceutical containing glucosamine, chondroitin sulphate and collagen (Carapeba et al., 2016) (Table 1). Dogs in both groups showed an improvement from baseline scores when assessed by subjective measurements (CPBI, Helsinki Chronic Pain Scale [HCPS] and veterinary assessments) up to 90 days post treatment. However, a greater improvement was shown in the dogs treated with IA HA. There is currently no licensed preparation of IA HA for use in dogs in the UK, and further evidence behind its use in canine OA is required before conclusions can be drawn as to its effectiveness in clinical cases.

**As this review is an update focusing primarily on recent treatments, or updated clinical evidence behind existing treatments, we refer the reader to previous systematic reviews outlining the evidence behind candidate SMOADs (Aragon *et al*., 2007, Sandersoln *et al*., 2009).**

Mesenchymal Stem Cells (MSCs)

Mesenchymal stem cells (MSCs) are progenitor cells with the ability to differentiate into numerous cell types, such as cells of connective tissue, bone and cartilage (Caplan, 1991). They can be derived from a variety of tissue including adipose tissue and bone marrow, which have both been shown to be a source of MSCs in dogs (Screven *et al*., 2014).

Although MSC treatment is often termed a regenerative therapy, the exact mechanisms by which MSCs exert their effect is still under investigation. It was previously thought that the main mechanism of action of the stem cells was that, once injected into the site of an OA lesion, they undergo differentiation into chondrocytes, therefore repairing the OA lesion (Scharstuhl *et al*., 2007). However, it is now thought that the effects of MSCs are exerted primarily through their secreted factors, including extracellular vesicles (EVs) and bioactive molecules such as chemokines, cytokines and growth factors, termed the secretome (Tofiño-Vian *et al*., 2018, Villatoro *et al*., 2019). These paracrine factors have a range of immunomodulatory, anti-inflammatory, angiogenic and anti-apoptotic properties (Phinney and Pittenger, 2017, Mocchi *et al*., 2020). Because of this growing knowledge of the importance of these secreted factors in the mechanism of action of MSCs, it has recently been proposed that they should now be known as “Medicinal Signalling Cells” rather than mesenchymal stem cells (Caplan, 2017).

Adipose tissue derived MSCs (ADSCs) are the favoured source of MSCs for clinical use in the dog due to the relative ease of accessibility and rapid rate of proliferation in culture (Zhu *et al*., 2008). MSCs can be autologous, allogenic or xenogenic depending on whether the cells are derived from the same dog that will be the recipient of the cells, from a different donor, or from a different species respectively (Cuervo *et al*., 2014, Cabon *et al*., 2019, Daems *et al*., 2019). In autologous canine ADSC treatment, subcutaneous adipose tissue is collected from the dog that will be the recipient of the ADSCs, and the cells are cultured and expanded *in vitro*, followed by IA injection into the affected joint (Cuervo *et al*., 2014)*.* The process of culturing the cells *in vitro* ensures a consistent and enriched MSC population, but can take seven to ten days, therefore increasing the cost of treatment (Voga *et al*., 2020). Alternatively, autologous MSC preparations can be performed by minimal manipulation, allowing for a faster preparation of an MSC product patient-side. These products, such as stromal vascular fraction (SVF) contain less MSCs per millilitre, and also contain other cell types (Franklin *et al*., 2018).

Clinical trials involving the use of MSCs in clinical cases of dogs with OA are generally of low sample size and involve different methodologies, but have shown some beneficial outcomes. A systematic review of the treatment of naturally occurring hip OA in dogs with ADSCs has been recently published (Olsson *et al*., 2021). This systematic review included six clinical trials, and concluded that there was evidence that the use of ADSCs by IA injection led to an improvement in clinical signs associated with OA in dogs and that both autologous and allogenic ADSCs were well tolerated with no adverse events noted. One of these studies involved comparing 38 dogs with OA treated with IA ADSCs compared to 36 given a saline placebo, and found significant improvement in subjective CSOM in the treated group (Harman and others 2016). Other studies included smaller numbers of dogs with 18 dogs or less. Only two studies utilised objective outcome measurements of lameness with force plate gait analysis (Vilar *et al*., 2014, Vilar *et al*., 2016), with the other four studies using subjective outcome measurements, including owner visual analogue scales (VAS) (Cuervo *et al*., 2014), CSOM (Srzentić Dražilov *et al*., 2018, Harman *et al*., 2016) and veterinary assessments (Cuervo *et al*., 2014, Marx *et al*., 2014, Harman *et al*., 2016, Vilar *et al*., 2016). Clinical trials have also examined the effect of MSC treatment on OA in the stifle joint and elbow joint in the dog (Kriston-Pál *et al*., 2017, Mohoric *et al*., 2016). A larger study involving cases of OA in various joints in 203 dogs found significant improvements on subjective veterinary assessments of clinical signs of OA ten weeks after either IA (n=128) or IV (n=65) treatment with an allogenic MSC preparation, although these improvements were more significant after IA than IV treatment (Shah *et al*., 2018). However, there was no control group in this study, and the outcomes were not assessed by a validated CMI.

The length of treatment efficacy after a single IA injection of MSCs differed between studies, and is limited by the length of some studies but has been reported to be from one month (Vilar *et al*., 2014) to up to six months following a single IA injection (Cabon *et al*., 2019, Cuervo *et al*., 2014).

The collection of adipose tissue or bone marrow for autologous MSC preparation involves some risk, with the potential for donor site morbidity to occur, and the need for a general anaesthetic (Redondo *et al*., 2007, Espinel-Rupérez *et al*., 2019). Allogenic MSC preparations can reduce this risk, but potentially hold a greater risk of an adverse immune response to treatment (Cabon *et al*., 2019, Oliveira *et al*., 2017). The administration of the product, like other IA products, also requires sedation or general anaesthesia (Redondo *et al*., 2007). However, minimal adverse effects to MSC treatment in dogs with OA have been reported in studies to date. These include local injection site reactions (Cabon *et al*., 2019), and a mild skin allergy in one dog (Shah *et al*., 2018).

A future area for development in MSC treatment research lies in examining extracellular vesicles (EVs) released from MSCs, as a potential cell-free version of treatment. EVs contain a number of biologically active signalling molecules that lead to the beneficial effects of MSCs in OA treatment , and by isolating EVs for use in therapies towards OA there may be less of an immune response towards treatment (Bari *et al*., 2019, Li *et al*., 2019). This area of research, however, is still in its infancy.

In veterinary practice, there are currently no defined guidelines for the dose or frequency of MSC treatment in clinical scenarios. Several companies offer assistance in the culture and preparation of MSCs, but there is currently a lack of standardisation of protocols between companies in terms of methods and clinical approach.

**Overall, MSCs show potential as an alternative, or adjunctive, treatment option in cases where conventional treatment is not providing adequate outcomes. Commercial systems are available, but there is a need for greater regulation and standardisation of methods. There is also a need for larger, multi-centre, randomised controlled clinical trials using a standardised clinical approach and method to better evaluate outcomes in clinical cases of canine OA (Sasaki *et al*., 2019, Olsson *et al*., 2021).**

Platelet Rich Plasma (PRP)

PRP is a blood derived product, and consists of plasma with a higher concentration of platelets than is present in peripheral blood (Rossi *et al*., 2019). It can also contain varying amounts of leukocytes (Carr *et al*., 2015, Alves *et al*., 2021a).Platelets are integral to blood clotting, and release growth factors such as platelet-derived growth factor, transforming growth factor beta, epidermal growth factor, and vascular endothelial growth factors (Verheul *et al*., 1997, Coppinger *et al*., 2004) . These growth factors stimulate processes such as angiogenesis, and chondrocyte proliferation, and reduce processes such as chondrocyte apoptosis (Collins *et al*., 2021).

The clinical use of PRP therapy for canine OA involves drawing a blood sample from the affected dog, which is then processed to produce plasma highly concentrated in platelets (Perego *et al*., 2020). This is then injected into the intra-articular space of the OA affected joint. Different commercial systems, both external and in-house, for the processing of canine PRP exists, and, like MSCs, there is currently no standardised method of preparation (Carr *et al*., 2015, Franklin *et al*., 2015).

The efficacy of PRP in OA treatment in humans is still under debate, with some systematic reviews on their use in human medicine showing an overall low level of evidence (Gato-Calvo *et al*., 2019), and others concluding that PRP was more efficacious than other conservative methods of OA treatment (Hong *et al*., 2021). In dogs, there are a limited number of clinical trials published, all involving small numbers of dogs (up to 24 dogs), and different methodologies (Cuervo *et al*., 2020).

Venator *et al*. (2020) investigated objective force plate analysis in dogs with non-stabilised CCL rupture treated with a single IA PRP injection. The authors found an improvement in kinetics for a minimum of four weeks post treatment, but there was no control group to act as a comparison. Upchurch *et al*. (2016) also found improvements in objective and subjective outcome measures in dogs with hip OA after IA and intravenous (IV) PRP and MSC treatment compared to placebo in 12 dogs. Another study examined objective lameness outcomes in dogs receiving IA PRP compared to another group receiving IA PRP as well as physiotherapy (Cuervo *et al*., 2020). Both groups improved compared to baseline levels. Five case reports of PRP treatment for refractory OA describe beneficial outcomes, but it is difficult to extrapolate these outcomes to the wider dog population due to the low number of case studies and lack of a control group (Catarino *et al*., 2020). A recent study by Alves *et al*. (2021a) compared outcomes of groups of 20 dogs with naturally occurring hip OA receiving either two injections of PRP given 14 days apart, or two IA saline injections as a negative control. Outcomes were measured by four different validated CMIs (LOAD, CBPI, Canine Orthopaedic Index [COI] and Hudson Visual Analogue Scale [HVAS]), and dogs were assessed at intervals up to 180 days post-treatment. A significant improvement was found in the PRP group compared to control, lasting for approximately 130 days.

No severe adverse reactions have been reported in the literature to date, although studies have described adverse reactions related to transient pain at the injection site which usually improves within 48 to 72 hours (Alves *et al*., 2021a). The need for sedation or general anaesthetic to administer the IA treatment also holds some level of risk (Redondo *et al*., 2007).

**Overall, PRP treatment requires further standardisation and regulation of different methods (Carr *et al*., 2015). Larger scale, multi-centre, randomised controlled clinical trials are required before firm conclusions over its effectiveness as a treatment for canine OA in practice can be drawn.**

Gene Therapy

Anti-inflammatory Cytokine Plasmid DNA Therapy

Anti-inflammatory cytokines, such as interleukin (IL)-10 can inhibit the production of pro-inflammatory cytokines, such as IL-6, IL-1β and tumour necrosis factor (TNF) α, as well as downregulating MMP production and preventing chondrocyte apoptosis (John *et al*., 2007, Kapoor *et al*., 2011). Previous studies investigating the use of IL-10 as an OA therapy have found limited benefits due to the short half-life *in vivo* (Chernoff *et al*., 1995). Recent investigations into novel therapies for OA have included research into the use of gene therapies that encode for anti-inflammatory cytokines, such as IL-10. In a recent randomised double-blinded placebo controlled clinical trial in 14 dogs with naturally occurring OA, IA targeted IL-10 plasmid DNA therapy demonstrated some beneficial effects on reducing clinical signs of OA based on owner and veterinary subjective VAS. The same paper described safety studies, and concluded that the treatment was well tolerated (Watkins *et al*., 2020).

**More work is required before IL-10 plasmid DNA therapies are available commercially, but this is an area for future development in OA treatment (Schulze-Tanzil, 2021).**

Conclusion

A greater understanding of the biology and pain mechanisms of OA has led to a growing number of pharmaceutical treatment options for canine OA in the past decade. With the advent of novel medications such as anti-NGF mAbs and piprants, as well as a growing number of adjunctive analgesics, and greater availability of regenerative therapies in veterinary medicine, veterinary practitioners now have more therapeutics to offer, hopefully improving the welfare of dogs with this condition. Although a search for a curative treatment of OA is ongoing and, as yet, elusive, there are still exciting areas for future development, such as gene and mRNA therapy (Schulze-Tanzil 2021).

This review highlights that there is a need for larger scale, randomised controlled clinical trials to improve the evidence underpinning treatments to ensure that both veterinary professionals and animal caregivers can have more confidence in the effectiveness of treatments in clinical cases. It should also be noted that a multi-modal approach to treatment, incorporating not only different types of pharmaceuticals, but also weight management, nutraceuticals, physiotherapy and other complementary therapies is important in the non-surgical treatment of this whole joint disease (Mlacnik *et al*., 2006).

References

ALVES, J. C., SANTOS, A. & JORGE, P. 2021a. Platelet-rich plasma therapy in dogs with bilateral hip osteoarthritis. *BMC Veterinary Research,* 17**,** 207.

ALVES, J. C., SANTOS, A., JORGE, P., LAVRADOR, C. & CARREIRA, L. M. 2021b. The intra-articular administration of triamcinolone hexacetonide in the treatment of osteoarthritis. Its effects in a naturally occurring canine osteoarthritis model. *PLOS ONE,* 16**,** e0245553.

AMERICAN VETERINARY MEDICAL ASSOCIATION. 2017. *AVMA pet ownership and demographics sourcebook.* Colorado State University. Libraries.

ANDERSON, K. L., O’NEILL, D. G., BRODBELT, D. C., CHURCH, D. B., MEESON, R. L., SARGAN, D., SUMMERS, J. F., ZULCH, H. & COLLINS, L. M. 2018. Prevalence, duration and risk factors for appendicular osteoarthritis in a UK dog population under primary veterinary care. *Scientific Reports,* 8**,** 5641.

ARAGON, C. L. & BUDSBERG, S. C. 2005. Applications of evidence-based medicine: cranial cruciate ligament injury repair in the dog. *Vet Surg,* 34**,** 93-8.

ARAGON, C. L., HOFMEISTER, E. H. & BUDSBERG, S. C. 2007. Systematic review of clinical trials of treatments for osteoarthritis in dogs. *Journal of the American Veterinary Medical Association,* 230**,** 514-521.

ARORA, M., CHOUDHARY, S., SINGH, P. K., SAPRA, B. & SILAKARI, O. 2020. Structural investigation on the selective COX-2 inhibitors mediated cardiotoxicity: A review. *Life Sciences,* 251**,** 117631.

BANNURU, R. R., OSANI, M. C., VAYSBROT, E. E., ARDEN, N. K., BENNELL, K., BIERMA-ZEINSTRA, S. M. A., KRAUS, V. B., LOHMANDER, L. S., ABBOTT, J. H., BHANDARI, M., BLANCO, F. J., ESPINOSA, R., HAUGEN, I. K., LIN, J., MANDL, L. A., MOILANEN, E., NAKAMURA, N., SNYDER-MACKLER, L., TROJIAN, T., UNDERWOOD, M. & MCALINDON, T. E. 2019. OARSI guidelines for the non-surgical management of knee, hip, and polyarticular osteoarthritis. *Osteoarthritis Cartilage,* 27**,** 1578-1589.

BARI, E., PERTEGHELLA, S., CATENACCI, L., SORLINI, M., CROCE, S., MANTELLI, M., AVANZINI, M. A., SORRENTI, M. & TORRE, M. L. 2019. Freeze-dried and GMP-compliant pharmaceuticals containing exosomes for acellular mesenchymal stromal cell immunomodulant therapy. *Nanomedicine (Lond),* 14**,** 753-765.

BELSHAW, Z., DEAN, R. & ASHER, L. 2020. “You can be blind because of loving them so much”: the impact on owners in the United Kingdom of living with a dog with osteoarthritis. *BMC veterinary research,* 16**,** 1-10.

BOUCK, G. R., MILLER, C. W. & TAVES, C. L. 1995. A Comparison of Surgical and Medical Treatment of Fragmented Coronoid Process and Osteochondritis Dissecans of the Canine Elbow. *Vet Comp Orthop Traumatol,* 08**,** 177-183.

BRIOSCHI, F. A., DI CESARE, F., GIOENI, D., RABBOGLIATTI, V., FERRARI, F., D’URSO, E. S., AMARI, M. & RAVASIO, G. 2020. Oral Transmucosal Cannabidiol Oil Formulation as Part of a Multimodal Analgesic Regimen: Effects on Pain Relief and Quality of Life Improvement in Dogs Affected by Spontaneous Osteoarthritis. *Animals,* 10**,** 1505.

BROWN, D. C., BELL, M. & RHODES, L. 2013a. Power of treatment success definitions when the Canine Brief Pain Inventory is used to evaluate carprofen treatment for the control of pain and inflammation in dogs with osteoarthritis. *Am J Vet Res,* 74**,** 1467-73.

BROWN, D. C., BOSTON, R. C., COYNE, J. C. & FARRAR, J. T. 2008. Ability of the canine brief pain inventory to detect response to treatment in dogs with osteoarthritis. *Journal of the American Veterinary Medical Association,* 233**,** 1278-1283.

BROWN, D. C., BOSTON, R. C. & FARRAR, J. T. 2013b. Comparison of Force Plate Gait Analysis and Owner Assessment of Pain Using the Canine Brief Pain Inventory in Dogs with Osteoarthritis. *Journal of Veterinary Internal Medicine,* 27**,** 22-30.

BUDSBERG, S. C., BERGH, M. S., REYNOLDS, L. R. & STREPPA, H. K. 2007. Evaluation of Pentosan Polysulfate Sodium in the Postoperative Recovery from Cranial Cruciate Injury in Dogs: A Randomized, Placebo-Controlled Clinical Trial. *Veterinary Surgery,* 36**,** 234-244.

BUDSBERG, S. C., KLEINE, S. A., NORTON, M. M. & SANDBERG, G. S. 2019. Comparison of two inhibitors of E-type prostanoid receptor four and carprofen in dogs with experimentally induced acute synovitis. *American Journal of Veterinary Research,* 80**,** 1001-1006.

BUDSBERG, S. C., TORRES, B. T., KLEINE, S. A., SANDBERG, G. S. & BERJESKI, A. K. 2018. Lack of effectiveness of tramadol hydrochloride for the treatment of pain and joint dysfunction in dogs with chronic osteoarthritis. *J Am Vet Med Assoc,* 252**,** 427-432.

CABON, Q., FEBRE, M., GOMEZ, N., CACHON, T., PILLARD, P., CAROZZO, C., SAULNIER, N., ROBERT, C., LIVET, V., RAKIC, R., PLANTIER, N., SAAS, P., MADDENS, S. & VIGUIER, E. 2019. Long-Term Safety and Efficacy of Single or Repeated Intra-Articular Injection of Allogeneic Neonatal Mesenchymal Stromal Cells for Managing Pain and Lameness in Moderate to Severe Canine Osteoarthritis Without Anti-inflammatory Pharmacological Support: Pilot Clinical Study. *Frontiers in Veterinary Science,* 6.

CALANDRE, E. P., RICO-VILLADEMOROS, F. & SLIM, M. 2016. Alpha(2)delta ligands, gabapentin, pregabalin and mirogabalin: a review of their clinical pharmacology and therapeutic use. *Expert Rev Neurother,* 16**,** 1263-1277.

CAPLAN, A. I. 1991. Mesenchymal stem cells. *J Orthop Res,* 9**,** 641-50.

CAPLAN, A. I. 2017. Mesenchymal Stem Cells: Time to Change the Name! *Stem Cells Transl Med,* 6**,** 1445-1451.

CARAPEBA, G. O., CAVALETI, P., NICÁCIO, G. M., BRINHOLI, R. B., GIUFFRIDA, R. & CASSU, R. N. 2016. Intra-Articular Hyaluronic Acid Compared to Traditional Conservative Treatment in Dogs with Osteoarthritis Associated with Hip Dysplasia. *Evid Based Complement Alternat Med,* 2016**,** 2076921.

CARR, B. J., CANAPP, S. O., JR., MASON, D. R., COX, C. & HESS, T. 2015. Canine Platelet-Rich Plasma Systems: A Prospective Analysis. *Front Vet Sci,* 2**,** 73.

CATARINO, J., CARVALHO, P., SANTOS, S., MARTINS, Â. & REQUICHA, J. 2020. Treatment of canine osteoarthritis with allogeneic platelet-rich plasma: review of five cases. *Open veterinary journal,* 10**,** 226-231.

CHERNOFF, A. E., GRANOWITZ, E. V., SHAPIRO, L., VANNIER, E., LONNEMANN, G., ANGEL, J. B., KENNEDY, J. S., RABSON, A. R., WOLFF, S. M. & DINARELLO, C. A. 1995. A randomized, controlled trial of IL-10 in humans. Inhibition of inflammatory cytokine production and immune responses. *The Journal of Immunology,* 154**,** 5492-5499.

CHUNEKAMRAI, S., KROOK, L. P., LUST, G. & MAYLIN, G. A. 1989. Changes in articular cartilage after intra-articular injections of methylprednisolone acetate in horses. *American journal of veterinary research,* 50**,** 1733-1741.

COLLINS, T., ALEXANDER, D. & BARKATALI, B. 2021. Platelet-rich plasma: a narrative review. *EFORT Open Reviews,* 6**,** 225-235.

COPPINGER, J. A., CAGNEY, G., TOOMEY, S., KISLINGER, T., BELTON, O., MCREDMOND, J. P., CAHILL, D. J., EMILI, A., FITZGERALD, D. J. & MAGUIRE, P. B. 2004. Characterization of the proteins released from activated platelets leads to localization of novel platelet proteins in human atherosclerotic lesions. *Blood,* 103**,** 2096-2104.

CORRAL, M. J., MOYAERT, H., FERNANDES, T., ESCALADA, M., KIRA S TENA, J., WALTERS, R. R. & STEGEMANN, M. R. 2021. A prospective, randomized, blinded, placebo-controlled multisite clinical study of bedinvetmab, a canine monoclonal antibody targeting nerve growth factor, in dogs with osteoarthritis. *Veterinary Anaesthesia and Analgesia*.

CUERVO, B., RUBIO, M., CHICHARRO, D., DAMIÁ, E., SANTANA, A., CARRILLO, J. M., ROMERO, A. D., VILAR, J. M., CERÓN, J. J. & SOPENA, J. J. 2020. Objective Comparison between Platelet Rich Plasma Alone and in Combination with Physical Therapy in Dogs with Osteoarthritis Caused by Hip Dysplasia. *Animals (Basel),* 10.

CUERVO, B., RUBIO, M., SOPENA, J., DOMINGUEZ, J. M., VILAR, J., MORALES, M., CUGAT, R. & CARRILLO, J. M. 2014. Hip Osteoarthritis in Dogs: A Randomized Study Using Mesenchymal Stem Cells from Adipose Tissue and Plasma Rich in Growth Factors. *International Journal of Molecular Sciences,* 15**,** 13437-13460.

CUI, A., LI, H., WANG, D., ZHONG, J., CHEN, Y. & LU, H. 2020. Global, regional prevalence, incidence and risk factors of knee osteoarthritis in population-based studies. *EClinicalMedicine,* 29.

DA COSTA, B. R., REICHENBACH, S., KELLER, N., NARTEY, L., WANDEL, S., JÜNI, P. & TRELLE, S. 2017. Effectiveness of non-steroidal anti-inflammatory drugs for the treatment of pain in knee and hip osteoarthritis: a network meta-analysis. *Lancet,* 390**,** e21-e33.

DAEMS, R., VAN HECKE, L., SCHWARZKOPF, I., DEPUYDT, E., BROECKX, S. Y., DAVID, M., BEERTS, C., VANDEKERCKHOVE, P. & SPAAS, J. H. 2019. A Feasibility Study on the Use of Equine Chondrogenic Induced Mesenchymal Stem Cells as a Treatment for Natural Occurring Osteoarthritis in Dogs. *Stem Cells International,* 2019**,** 4587594.

DE SALAZAR ALCALÁ, A. G., GIODA, L., DEHMAN, A. & BEUGNET, F. 2019. Assessment of the efficacy of firocoxib (Previcox®) and grapiprant (Galliprant®) in an induced model of acute arthritis in dogs. *BMC Vet Res,* 15**,** 309.

DHARMSHAKTU, P., TAYAL, V. & KALRA, B. S. 2012. Efficacy of antidepressants as analgesics: a review. *J Clin Pharmacol,* 52**,** 6-17.

ENOMOTO, M., MANTYH, P. W., MURRELL, J., INNES, J. F. & LASCELLES, B. D. X. 2019. Anti-nerve growth factor monoclonal antibodies for the control of pain in dogs and cats. *The Veterinary record,* 184**,** 23-23.

ESPINEL-RUPÉREZ, J., MARTÍN-RÍOS, M. D., SALAZAR, V., BAQUERO-ARTIGAO, M. R. & ORTIZ-DÍEZ, G. 2019. Incidence of surgical site infection in dogs undergoing soft tissue surgery: risk factors and economic impact. *Veterinary record open,* 6**,** e000233-e000233.

EUROPEAN MEDICINES AGENCY. 2008. European Medicines Agency 2008. Trocoxil: EPAR- Scientific Discussion Available on <https://www.ema.europa.eu/en/documents/scientific-discussion/trocoxil-epar-scientific-discussion_en.pdf> [Accessed 11/10/21].

FARQUHAR, T., TODHUNTER, R. J., FUBINI, S. L., BURTON-WURSTER, N. & LUST, G. 1996. Effect of methylprednisolone and mechanical loading on canine articular cartilage in explant culture. *Osteoarthritis and Cartilage,* 4**,** 55-62.

FISHER, K., CODERRE, T. J. & HAGEN, N. A. 2000. Targeting the N-methyl-D-aspartate receptor for chronic pain management. Preclinical animal studies, recent clinical experience and future research directions. *J Pain Symptom Manage,* 20**,** 358-73.

FRANKLIN, S. P., GARNER, B. C. & COOK, J. L. 2015. Characteristics of canine platelet-rich plasma prepared with five commercially available systems. *American Journal of Veterinary Research,* 76**,** 822-827.

FRANKLIN, S. P., STOKER, A. M., BOZYNSKI, C. C., KUROKI, K., CLARKE, K. M., JOHNSON, J. K. & COOK, J. L. 2018. Comparison of Platelet-Rich Plasma, Stromal Vascular Fraction (SVF), or SVF with an Injectable PLGA Nanofiber Scaffold for the Treatment of Osteochondral Injury in Dogs. *J Knee Surg,* 31**,** 686-697.

FREISE, K. J., LIN, T. L., FAN, T. M., RECTA, V. & CLARK, T. P. 2013. Evidence-based medicine: the design and interpretation of noninferiority clinical trials in veterinary medicine. *J Vet Intern Med,* 27**,** 1305-17.

FU, K., ROBBINS, S. R. & MCDOUGALL, J. J. 2018. Osteoarthritis: the genesis of pain. *Rheumatology,* 57**,** iv43-iv50.

GAMBLE, L. J., BOESCH, J. M., FRYE, C. W., SCHWARK, W. S., MANN, S., WOLFE, L., BROWN, H., BERTHELSEN, E. S. & WAKSHLAG, J. J. 2018. Pharmacokinetics, Safety, and Clinical Efficacy of Cannabidiol Treatment in Osteoarthritic Dogs. *Front Vet Sci,* 5**,** 165.

GATO-CALVO, L., MAGALHAES, J., RUIZ-ROMERO, C., BLANCO, F. J. & BURGUERA, E. F. 2019. Platelet-rich plasma in osteoarthritis treatment: review of current evidence. *Therapeutic advances in chronic disease,* 10**,** 2040622319825567-2040622319825567.

GHOSH, P. 1999. The pathobiology of osteoarthritis and the rationale for the use of pentosan polysulfate for its treatment. *Semin Arthritis Rheum,* 28**,** 211-67.

GLYN-JONES, S., PALMER, A., AGRICOLA, R., PRICE, A., VINCENT, T., WEINANS, H. & CARR, A. 2015. Osteoarthritis. *The Lancet,* 386**,** 376-387.

GROND, S. & SABLOTZKI, A. 2004. Clinical pharmacology of tramadol. *Clin Pharmacokinet,* 43**,** 879-923.

HARMAN, R., CARLSON, K., GAYNOR, J., GUSTAFSON, S., DHUPA, S., CLEMENT, K., HOELZLER, M., MCCARTHY, T., SCHWARTZ, P. & ADAMS, C. 2016. A Prospective, Randomized, Masked, and Placebo-Controlled Efficacy Study of Intraarticular Allogeneic Adipose Stem Cells for the Treatment of Osteoarthritis in Dogs. *Frontiers in Veterinary Science,* 3.

HEFTI, F. 2020. Pharmacology of nerve growth factor and discovery of tanezumab, an anti-nerve growth factor antibody and pain therapeutic. *Pharmacol Res,* 154**,** 104240.

HONG, M., CHENG, C., SUN, X., YAN, Y., ZHANG, Q., WANG, W. & GUO, W. 2021. Efficacy and Safety of Intra-Articular Platelet-Rich Plasma in Osteoarthritis Knee: A Systematic Review and Meta-Analysis. *Biomed Res Int,* 2021**,** 2191926.

HUNT, J. R., DEAN, R. S., DAVIS, G. N. & MURRELL, J. C. 2015. An analysis of the relative frequencies of reported adverse events associated with NSAID administration in dogs and cats in the United Kingdom. *Vet J,* 206**,** 183-90.

INNES, J. F., BARR, A. R. & SHARIF, M. 2000. Efficacy of oral calcium pentosan polysulphate for the treatment of osteoarthritis of the canine stifle joint secondary to cranial cruciate ligament deficiency. *Vet Rec,* 146**,** 433-7.

JOHN, T., MÜLLER, R., OBERHOLZER, A., ZREIQAT, H., KOHL, B., ERTEL, W., HOSTMANN, A., TSCHOEKE, S. & SCHULZE-TANZIL, G. 2007. Interleukin-10 modulates pro-apoptotic effects of TNF-α in human articular chondrocytes in vitro. *Cytokine,* 40**,** 226-234.

JOHNSON, J. W. & KOTERMANSKI, S. E. 2006. Mechanism of action of memantine. *Current Opinion in Pharmacology,* 6**,** 61-67.

JOHNSTON, S. A. 1997. Osteoarthritis. Joint anatomy, physiology, and pathobiology. *Vet Clin North Am Small Anim Pract,* 27**,** 699-723.

JOHNSTON, S. A. & BUDSBERG, S. C. 1997. Nonsteroidal Anti-Inflammatory Drugs and Corticosteroids for The Management of Canine Osteoarthritis. *Veterinary Clinics of North America: Small Animal Practice,* 27**,** 841-862.

KAPOOR, M., MARTEL-PELLETIER, J., LAJEUNESSE, D., PELLETIER, J.-P. & FAHMI, H. 2011. Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. *Nature Reviews Rheumatology,* 7**,** 33.

KIRKBY SHAW, K., RAUSCH‐DERRA, L. C. & RHODES, L. 2016. Grapiprant: an EP 4 prostaglandin receptor antagonist and novel therapy for pain and inflammation. *Veterinary medicine and science,* 2**,** 3-9.

KNAZOVICKY, D., HELGESON, E. S., CASE, B., GRUEN, M. E., MAIXNER, W. & LASCELLES, B. D. X. 2016. Widespread somatosensory sensitivity in naturally occurring canine model of osteoarthritis. *Pain,* 157**,** 1325-1332.

KOGAN, L., HELLYER, P. & DOWNING, R. 2020. The use of Cannabidiol-rich hemp oil extract to treat canine osteoarthritis-related pain: a pilot study. *AHVMA J,* 58**,** 1-10.

KRAUTMANN, M., WALTERS, R., COLE, P., TENA, J., BERGERON, L. M., MESSAMORE, J., MWANGI, D., RAI, S., DOMINOWSKI, P., SAAD, K., ZHU, Y., GUILLOT, M. & CHOUINARD, L. 2021. Laboratory safety evaluation of bedinvetmab, a canine anti-nerve growth factor monoclonal antibody, in dogs. *The Veterinary Journal,* 276**,** 105733.

KRISTON-PÁL, É., CZIBULA, Á., GYURIS, Z., BALKA, G., SEREGI, A., SÜKÖSD, F., SÜTH, M., KISS-TÓTH, E., HARACSKA, L., UHER, F. & MONOSTORI, É. 2017. Characterization and therapeutic application of canine adipose mesenchymal stem cells to treat elbow osteoarthritis. *Canadian journal of veterinary research = Revue canadienne de recherche veterinaire,* 81**,** 73-78.

KUKANICH, B. 2010. Pharmacokinetics of acetaminophen, codeine, and the codeine metabolites morphine and codeine-6-glucuronide in healthy Greyhound dogs. *J Vet Pharmacol Ther,* 33**,** 15-21.

KUKANICH, B., BIDGOOD, T. & KNESL, O. 2012. Clinical pharmacology of nonsteroidal anti-inflammatory drugs in dogs. *Veterinary Anaesthesia and Analgesia,* 39**,** 69-90.

KUKANICH, B. & COHEN, R. L. 2011. Pharmacokinetics of oral gabapentin in greyhound dogs. *Vet J,* 187**,** 133-5.

KUKANICH, B. & PAPICH, M. G. 2004. Pharmacokinetics of tramadol and the metabolite O-desmethyltramadol in dogs. *J Vet Pharmacol Ther,* 27**,** 239-46.

LANDA, L., !`ULCOVÁ, A. & GBELEC, P. 2016. The use of cannabinoids in animals and therapeutic implications for veterinary medicine: a review. *Veterinarni Medicina,* 61**,** 111-122.

LASCELLES, B. D. X., GAYNOR, J. S., SMITH, E. S., ROE, S. C., MARCELLIN-LITTLE, D. J., DAVIDSON, G., BOLAND, E. & CARR, J. 2008. Amantadine in a Multimodal Analgesic Regimen for Alleviation of Refractory Osteoarthritis Pain in Dogs. *Journal of Veterinary Internal Medicine,* 22**,** 53-59.

LASCELLES, B. D. X., KNAZOVICKY, D., CASE, B., FREIRE, M., INNES, J. F., DREW, A. C. & GEARING, D. P. 2015. A canine-specific anti-nerve growth factor antibody alleviates pain and improves mobility and function in dogs with degenerative joint disease-associated pain. *BMC Veterinary Research,* 11**,** 101.

LEES, P., PELLIGAND, L., ELLIOTT, J., TOUTAIN, P. L., MICHELS, G. & STEGEMANN, M. 2015. Pharmacokinetics, pharmacodynamics, toxicology and therapeutics of mavacoxib in the dog: a review. *J Vet Pharmacol Ther,* 38**,** 1-14.

LI, J. J., HOSSEINI-BEHESHTI, E., GRAU, G. E., ZREIQAT, H. & LITTLE, C. B. 2019. Stem Cell-Derived Extracellular Vesicles for Treating Joint Injury and Osteoarthritis. *Nanomaterials (Basel, Switzerland),* 9**,** 261.

LIN, C.-R., AMAYA, F., BARRETT, L., WANG, H., TAKADA, J., SAMAD, T. A. & WOOLF, C. J. 2006. Prostaglandin E2 receptor EP4 contributes to inflammatory pain hypersensitivity. *Journal of Pharmacology and Experimental Therapeutics,* 319**,** 1096-1103.

LOESER, R. F., GOLDRING, S. R., SCANZELLO, C. R. & GOLDRING, M. B. 2012. Osteoarthritis: a disease of the joint as an organ. *Arthritis and rheumatism,* 64**,** 1697-1707.

MACHADO, G. C., MAHER, C. G., FERREIRA, P. H., PINHEIRO, M. B., LIN, C.-W. C., DAY, R. O., MCLACHLAN, A. J. & FERREIRA, M. L. 2015. Efficacy and safety of paracetamol for spinal pain and osteoarthritis: systematic review and meta-analysis of randomised placebo controlled trials. *BMJ : British Medical Journal,* 350**,** h1225.

MALEK, S., SAMPLE, S. J., SCHWARTZ, Z., NEMKE, B., JACOBSON, P. B., COZZI, E. M., SCHAEFER, S. L., BLEEDORN, J. A., HOLZMAN, G. & MUIR, P. 2012. Effect of analgesic therapy on clinical outcome measures in a randomized controlled trial using client-owned dogs with hip osteoarthritis. *BMC Vet Res,* 8**,** 185.

MARX, C., SILVEIRA, M. D., SELBACH, I., DA SILVA, A. S., BRAGA, L. M. G. D. M., CAMASSOLA, M. & NARDI, N. B. 2014. Acupoint Injection of Autologous Stromal Vascular Fraction and Allogeneic Adipose-Derived Stem Cells to Treat Hip Dysplasia in Dogs. *Stem Cells International,* 2014**,** 391274.

MCKELLAR, Q. A., PEARSON, T., GALBRAITH, E. A., BOYLE, J. & BELL, G. 1991. Pharmacokinetics and clinical efficacy of a cinchophen and prednisolone combination in the dog. *Journal of Small Animal Practice,* 32**,** 53-58.

MEJIA, S., DUERR, F. M., GRIFFENHAGEN, G. & MCGRATH, S. 2021. Evaluation of the Effect of Cannabidiol on Naturally Occurring Osteoarthritis-Associated Pain: A Pilot Study in Dogs. *Journal of the American Animal Hospital Association,* 57**,** 81-90.

MILES, J., BØJESEN, J., CHRISTENSEN, P., ANDERSEN-RANBERG, E., VITGER, A., POULSEN, H. H. & NIELSEN, L. N. 2020. Tramadol and gabapentin improve peak vertical force in osteoarthritic dogs already receiving non-steroidal anti-inflammatory drugs. *BSAVA Congress Proceedings 2020.* British Small Animal Veterinary Association.

MLACNIK, E., BOCKSTAHLER, B. A., MÜLLER, M., TETRICK, M. A., NAP, R. C. & ZENTEK, J. 2006. Effects of caloric restriction and a moderate or intense physiotherapy program for treatment of lameness in overweight dogs with osteoarthritis. *Journal of the American Veterinary Medical Association,* 229**,** 1756-1760.

MOCCHI, M., DOTTI, S., BUE, M. D., VILLA, R., BARI, E., PERTEGHELLA, S., TORRE, M. L. & GROLLI, S. 2020. Veterinary Regenerative Medicine for Musculoskeletal Disorders: Can Mesenchymal Stem/Stromal Cells and Their Secretome Be the New Frontier? *Cells,* 9.

MOHORIC, L., ZORKO, B., CEH, K. & MAJDIC, G. 2016. Blinded placebo study of bilateral osteoarthritis treatment using adipose derived mesenchymal stem cells. *Slovenian Vet Res,* 53.

MONTEIRO-STEAGALL, B. P., STEAGALL, P. V. M. & LASCELLES, B. D. X. 2013. Systematic Review of Nonsteroidal Anti-Inflammatory Drug-Induced Adverse Effects in Dogs. *Journal of Veterinary Internal Medicine,* 27**,** 1011-1019.

MONTEIRO, B. & STEAGALL, P. V. 2019. Antiinflammatory Drugs. *Veterinary Clinics of North America: Small Animal Practice,* 49**,** 993-1011.

MONTEIRO, B. P., LAMBERT, C., BIANCHI, E., GENEVOIS, J. P., SOLDANI, G. & TRONCY, E. 2019. Safety and efficacy of reduced dosage ketoprofen with or without tramadol for long-term treatment of osteoarthritis in dogs: a randomized clinical trial. *BMC Vet Res,* 15**,** 213.

MURPHY, D. J., TODHUNTER, R. J., FUBINI, S. L., VERNIER-SINGER, M., STRAUBINGER, R. K. & LUST, G. 2000. The effects of methylprednisolone on normal and monocyte-conditioned medium-treated articular cartilage from dogs and horses. *Vet Surg,* 29**,** 546-57.

NAJM, A., ALUNNO, A., GWINNUTT, J. M., WEILL, C. & BERENBAUM, F. 2021. Efficacy of intra-articular corticosteroid injections in knee osteoarthritis: A systematic review and meta-analysis of randomized controlled trials. *Joint Bone Spine,* 88**,** 105198.

NAKAO, K., MURASE, A., OHSHIRO, H., OKUMURA, T., TANIGUCHI, K., MURATA, Y., MASUDA, M., KATO, T., OKUMURA, Y. & TAKADA, J. 2007. CJ-023,423, a novel, potent and selective prostaglandin EP4 receptor antagonist with antihyperalgesic properties. *Journal of Pharmacology and Experimental Therapeutics,* 322**,** 686-694.

NATIONAL OFFICE OF ANIMAL HEALTH. 2021a. *NOAH compendium, Datasheet for Daxocox tablets for dogs (Animalcare Limited).* [Online]. Available: <https://www.noahcompendium.co.uk/?id=-479652> [Accessed 11/10/21 2021].

NATIONAL OFFICE OF ANIMAL HEALTH. 2021b. *NOAH Compendium, Datasheet for Galliprant (Elanco) tablets for dogs* [Online]. Available: <https://www.noahcompendium.co.uk/?id=-472523> [Accessed 08/10/21 2021].

NATIONAL OFFICE OF ANIMAL HEALTH. 2021c. NOAH Compendium, Datasheet for Librela® solution for injection for dogs by Zoetis UK Ltd. Available on <https://www.noahcompendium.co.uk/?id=-478735> [accessed 25/05/21].

O'NEILL, D. G., CHURCH, D. B., MCGREEVY, P. D., THOMSON, P. C. & BRODBELT, D. C. 2014a. Approaches to canine health surveillance. *Canine Genetics and Epidemiology,* 1**,** 2.

O'NEILL, D. G., CHURCH, D. B., MCGREEVY, P. D., THOMSON, P. C. & BRODBELT, D. C. 2014b. Prevalence of disorders recorded in dogs attending primary-care veterinary practices in England. *PloS one,* 9**,** e90501-e90501.

OLIVEIRA, R. L., CHAGASTELLES, P. C., SESTERHEIM, P. & PRANKE, P. 2017. In Vivo Immunogenic Response to Allogeneic Mesenchymal Stem Cells and the Role of Preactivated Mesenchymal Stem Cells Cotransplanted with Allogeneic Islets. *Stem Cells Int,* 2017**,** 9824698.

OLSSON, D. C., TEIXEIRA, B. L., JEREMIAS, T. D. S., RÉUS, J. C., DE LUCA CANTO, G., PORPORATTI, A. L. & TRENTIN, A. G. 2021. Administration of mesenchymal stem cells from adipose tissue at the hip joint of dogs with osteoarthritis: A systematic review. *Research in Veterinary Science,* 135**,** 495-503.

ONAKPOYA, I. J. 2020. Paracetamol as first line for treatment of knee and hip osteoarthritis. *BMJ Evid Based Med,* 25**,** 40.

PAYNE-JOHNSON, M., BECSKEI, C., CHAUDHRY, Y. & STEGEMANN, M. R. 2015. Comparative efficacy and safety of mavacoxib and carprofen in the treatment of canine osteoarthritis. *Veterinary Record,* 176**,** 284-284.

PEREGO, R., SPADA, E., BAGGIANI, L., MARTINO, P. A. & PROVERBIO, D. 2020. Efficacy of a Semi Automated Commercial Closed System for Autologous Leukocyte- and Platelet-Rich Plasma (l-prp) Production in Dogs: A Preliminary Study. *Animals (Basel),* 10.

PEREZ JIMENEZ, T. E., MEALEY, K. L., GRUBB, T. L., GREENE, S. A. & COURT, M. H. 2016. Tramadol metabolism to O-desmethyl tramadol (M1) and N-desmethyl tramadol (M2) by dog liver microsomes: Species comparison and identification of responsible canine cytochrome P-450s (CYPs). *Drug Metab Dispos,* 44**,** 1963-1972.

PET FOOD MANUFACTURERS ASSOCIATION. 2021. *Pet Population 2021* [Online]. Available: <https://www.pfma.org.uk/pet-population-2021> [Accessed 08/10/21 2021].

PETTITT, R. A. & GERMAN, A. J. 2015. Investigation and management of canine osteoarthritis. *In Practice,* 37**,** 1-8.

PHINNEY, D. G. & PITTENGER, M. F. 2017. Concise Review: MSC-Derived Exosomes for Cell-Free Therapy. *Stem Cells,* 35**,** 851-858.

PIRAS, L. A., MANCUSI, D., OLIMPO, M., GASTALDI, L., ROSSO, V., PANERO, E., STAFFIERI, F. & PEIRONE, B. 2021. Post-operative analgesia following TPLO surgery: A comparison between cimicoxib and tramadol. *Res Vet Sci,* 136**,** 351-359.

PRZYBYŁA, G. W., SZYCHOWSKI, K. A. & GMIŃSKI, J. 2021. Paracetamol – An old drug with new mechanisms of action. *Clinical and Experimental Pharmacology and Physiology,* 48**,** 3-19.

RAUSCH-DERRA, L., HUEBNER, M., WOFFORD, J. & RHODES, L. 2016. A prospective, randomized, masked, placebo-controlled multisite clinical study of grapiprant, an EP4 prostaglandin receptor antagonist (PRA), in dogs with osteoarthritis. *Journal of Veterinary Internal Medicine,* 30**,** 756-763.

RAUSCH-DERRA, L. C., HUEBNER, M. & RHODES, L. 2015. Evaluation of the safety of long-term, daily oral administration of grapiprant, a novel drug for treatment of osteoarthritic pain and inflammation, in healthy dogs. *American Journal of Veterinary Research,* 76**,** 853-859.

READ, R. A., CULLIS-HILL, D. & JONES, M. P. 1996. Systemic use of pentosan polysulphate in the treatment of osteoarthritis. *J Small Anim Pract,* 37**,** 108-14.

REDONDO, J. I., RUBIO, M., SOLER, G., SERRA, I., SOLER, C. & GÓMEZ-VILLAMANDOS, R. J. 2007. Normal Values and Incidence of Cardiorespiratory Complications in Dogs During General Anaesthesia. A Review of 1281 Cases. *Journal of Veterinary Medicine Series A,* 54**,** 470-477.

REYMOND, N., SPERANZA, C., GRUET, P., SEEWALD, W. & KING, J. 2012. Robenacoxib vs. carprofen for the treatment of canine osteoarthritis; a randomized, noninferiority clinical trial. *Journal of veterinary pharmacology and therapeutics,* 35**,** 175-183.

RICHARDSON, J. D. 2000. Cannabinoids modulate pain by multiple mechanisms of action. *The Journal of Pain,* 1**,** 2-14.

ROSSI, L. A., MURRAY, I. R., CHU, C. R., MUSCHLER, G. F., RODEO, S. A. & PIUZZI, N. S. 2019. Classification systems for platelet-rich plasma. *The Bone & Joint Journal,* 101-B**,** 891-896.

SALAZAR, V., DEWEY, C. W., SCHWARK, W., BADGLEY, B. L., GLEED, R. D., HORNE, W. & LUDDERS, J. W. 2009. Pharmacokinetics of single-dose oral pregabalin administration in normal dogs. *Vet Anaesth Analg,* 36**,** 574-80.

SALICHS, M., BADIELLA, L., SARASOLA, P. & HOMEDES, J. 2021. Efficacy and safety of enflicoxib for treatment of canine osteoarthritis: A 6-week randomised, controlled, blind, multicentre clinical trial. *Veterinary Record,* n/a**,** e949.

SANDERSOLN, R. O., BEATA, C., FLIPO, R. M., GENEVOIS, J. P., MACIAS, C., TACKE, S., VEZZONI, A. & INNES, J. F. 2009. Systematic review of the management of canine osteoarthritis. *Veterinary Record,* 164**,** 418-424.

SASAKI, A., MIZUNO, M., MOCHIZUKI, M. & SEKIYA, I. 2019. Mesenchymal stem cells for cartilage regeneration in dogs. *World J Stem Cells,* 11**,** 254-269.

SCHARSTUHL, A., SCHEWE, B., BENZ, K., GAISSMAIER, C., BÜHRING, H. J. & STOOP, R. 2007. Chondrogenic potential of human adult mesenchymal stem cells is independent of age or osteoarthritis etiology. *Stem Cells,* 25**,** 3244-51.

SCHENCK, E. G. & AREND, I. 1978. [The effect of tramadol in an open clinical trial (author's transl)]. *Arzneimittelforschung,* 28**,** 209-12.

SCHNEIDER, B. M., DODMAN, N. H. & MARANDA, L. 2009. Use of memantine in treatment of canine compulsive disorders. *Journal of Veterinary Behavior,* 4**,** 118-126.

SCHULZE-TANZIL, G. 2021. Experimental therapeutics for the treatment of osteoarthritis. *Journal of Experimental Pharmacology,* 13**,** 101-125.

SCREVEN, R., KENYON, E., MYERS, M. J., YANCY, H. F., SKASKO, M., BOXER, L., BIGLEY, E. C., BORJESSON, D. L. & ZHU, M. 2014. Immunophenotype and gene expression profile of mesenchymal stem cells derived from canine adipose tissue and bone marrow. *Veterinary Immunology and Immunopathology,* 161**,** 21-31.

SEVALLA, K., TODHUNTER, R. J., VERNIER-SINGER, M. & BUDSBERG, S. C. 2000. Effect of Polysulfated Glycosaminoglycan on DNA Content and Proteoglycan Metabolism in Normal and Osteoarthritic Canine Articular Cartilage Explants. *Veterinary Surgery,* 29**,** 407-414.

SHAH, K., DRURY, T., ROIC, I., HANSEN, P., MALIN, M., BOYD, R., SUMER, H. & FERGUSON, R. 2018. Outcome of Allogeneic Adult Stem Cell Therapy in Dogs Suffering from Osteoarthritis and Other Joint Defects. *Stem cells international,* 2018**,** 7309201-7309201.

SHLOPOV, B. V., SMITH JR, G. N., COLE, A. A. & HASTY, K. A. 1999. Differential patterns of response to doxycycline and transforming growth factor β1 in the down‐regulation of collagenases in osteoarthritic and normal human chondrocytes. *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology,* 42**,** 719-727.

SILLS, G. J. 2006. The mechanisms of action of gabapentin and pregabalin. *Curr Opin Pharmacol,* 6**,** 108-13.

SMITH, J., HANNON, R., BRUNNBERG, L., GEBSKI, V. & CULLIS-HIII, D. 2001. A Randomised double blind comparator clinical study of the efficacy of sodium pentosan Polysulfate injection and carprofen capsules in arthritic dogs. *Journal of the Osteoarthritis Research Society International,* 9**,** S21-S22.

SOKOLOVE, J. & LEPUS, C. M. 2013. Role of inflammation in the pathogenesis of osteoarthritis: latest findings and interpretations. *Therapeutic advances in musculoskeletal disease,* 5**,** 77-94.

SRZENTIĆ DRAŽILOV, S., MRKOVAČKI, J., SPASOVSKI, V., FAZLAGIĆ, A., PAVLOVIĆ, S. & NIKČEVIĆ, G. 2018. The use of canine mesenchymal stem cells for the autologous treatment of osteoarthritis. *Acta Veterinaria Hungarica,* 66**,** 376-389.

SUNAGA, T., OH, N., HOSOYA, K., TAKAGI, S. & OKUMURA, M. 2012. Inhibitory effects of pentosan polysulfate sodium on MAP-kinase pathway and NF-κB nuclear translocation in canine chondrocytes in vitro. *J Vet Med Sci,* 74**,** 707-11.

TOFIÑO-VIAN, M., GUILLÉN, M. I. & ALCARAZ, M. J. 2018. Extracellular vesicles: A new therapeutic strategy for joint conditions. *Biochem Pharmacol,* 153**,** 134-146.

TOUTAIN, C. E., BROSSARD, P., KING, S. B. & HELBIG, R. 2018. Six-month safety evaluation of robenacoxib tablets (Onsior™) in dogs after daily oral administrations. *BMC Veterinary Research,* 14**,** 242.

UPCHURCH, D. A., RENBERG, W. C., ROUSH, J. K., MILLIKEN, G. A. & WEISS, M. L. 2016. Effects of administration of adipose-derived stromal vascular fraction and platelet-rich plasma to dogs with osteoarthritis of the hip joints. *Am J Vet Res,* 77**,** 940-51.

VENATOR, K. P., FRYE, C. W., GAMBLE, L. J. & WAKSHLAG, J. J. 2020. Assessment of a Single Intra-Articular Stifle Injection of Pure Platelet Rich Plasma on Symmetry Indices in Dogs with Unilateral or Bilateral Stifle Osteoarthritis from Long-Term Medically Managed Cranial Cruciate Ligament Disease. *Vet Med (Auckl),* 11**,** 31-38.

VERHEUL, H. M., HOEKMAN, K., LUYKX-DE BAKKER, S., EEKMAN, C. A., FOLMAN, C. C., BROXTERMAN, H. J. & PINEDO, H. M. 1997. Platelet: transporter of vascular endothelial growth factor. *Clin Cancer Res,* 3**,** 2187-90.

VERRICO, C. D., WESSON, S., KONDURI, V., HOFFEREK, C. J., VAZQUEZ-PEREZ, J., BLAIR, E., DUNNER, K., JR., SALIMPOUR, P., DECKER, W. K. & HALPERT, M. M. 2020. A randomized, double-blind, placebo-controlled study of daily cannabidiol for the treatment of canine osteoarthritis pain. *Pain,* 161**,** 2191-2202.

VETERINARY MEDICINE DIRECTORATE. 2018. *Guidance: Animal Test Certificates* [Online]. Available: <https://www.gov.uk/guidance/animal-test-certificates> [Accessed 22/10/21 2021].

VILAR, J. M., BATISTA, M., MORALES, M., SANTANA, A., CUERVO, B., RUBIO, M., CUGAT, R., SOPENA, J. & CARRILLO, J. M. 2014. Assessment of the effect of intraarticular injection of autologous adipose-derived mesenchymal stem cells in osteoarthritic dogs using a double blinded force platform analysis. *BMC Veterinary Research,* 10**,** 143.

VILAR, J. M., CUERVO, B., RUBIO, M., SOPENA, J., DOMÍNGUEZ, J. M., SANTANA, A. & CARRILLO, J. M. 2016. Effect of intraarticular inoculation of mesenchymal stem cells in dogs with hip osteoarthritis by means of objective force platform gait analysis: concordance with numeric subjective scoring scales. *BMC veterinary research,* 12**,** 1-10.

VILLATORO, A. J., ALCOHOLADO, C., MARTÍN-ASTORGA, M. C., FERNÁNDEZ, V., CIFUENTES, M. & BECERRA, J. 2019. Comparative analysis and characterization of soluble factors and exosomes from cultured adipose tissue and bone marrow mesenchymal stem cells in canine species. *Vet Immunol Immunopathol,* 208**,** 6-15.

VOGA, M., ADAMIC, N., VENGUST, M. & MAJDIC, G. 2020. Stem Cells in Veterinary Medicine-Current State and Treatment Options. *Frontiers in veterinary science,* 7**,** 278-278.

WALTON, M. B., COWDEROY, E. C., WUSTEFELD-JANSSENS, B., LASCELLES, B. D. X. & INNES, J. F. 2014. Mavacoxib and meloxicam for canine osteoarthritis: a randomised clinical comparator trial. *Veterinary Record,* 175**,** 280-280.

WATKINS, L. R., CHAVEZ, R. A., LANDRY, R., FRY, M., GREEN-FULGHAM, S. M., COULSON, J. D., COLLINS, S. D., GLOVER, D. K., RIEGER, J. & FORSAYETH, J. R. 2020. Targeted interleukin-10 plasmid DNA therapy in the treatment of osteoarthritis: Toxicology and pain efficacy assessments. *Brain, Behavior, and Immunity,* 90**,** 155-166.

WEBSTER, R. P., ANDERSON, G. I. & GEARING, D. P. 2014. Canine Brief Pain Inventory scores for dogs with osteoarthritis before and after administration of a monoclonal antibody against nerve growth factor. *Am J Vet Res,* 75**,** 532-5.

WESSMANN, A., STALIN, K. & KISIELEWICZ, C. 2016. Use of cannabidiol (CBD) in dogs and cats. *Scientific information sheets.* British Small Animal Veterinary Association.

WHITE, K. & HUNT, J. 2019. Chronic and osteoarthritic pain. *BSAVA Guide to Pain Management in Small Animal Practice.* British Small Animal Veterinary Association.

YAMAOKA, T. T. & AUCKBURALLY, A. 2014. Adjunctive analgesics. *The Veterinary Nurse,* 5**,** 474-481.

ZHU, Y., LIU, T., SONG, K., FAN, X., MA, X. & CUI, Z. 2008. Adipose‐derived stem cell: a better stem cell than BMSC. *Cell Biochemistry and Function: Cellular biochemistry and its modulation by active agents or disease,* 26**,** 664-675.

Figure Legends

Figure 1: Illustration of an osteoarthritic knee joint showing the four main stages of the pain pathway in osteoarthritis and the targets of certain drugs. Transduction is the conversion of a nociceptive stimulus into electrical impulse. Transmission is the electrical impulse transmitted from peripheral sensory nerves to the central nervous system (CNS). Modulation is how the nociceptive stimulus is processed by the CNS and includes the endogenous opioid system, as well as ascending input and descending inhibitory pathways. Perception is how the brain (particularly the somatosensory cortex) interprets nociceptive inputs resulting in conscious perception of pain. Adapted from Yamaoka and Auckburally (2014)

Figure 2: Targets of corticosteroids, COX inhibiting NSAIDs and piprants on the pathway of arachidonic acid metabolism. Corticosteroids inhibit phospholipase A2, preventing the conversion of phospholipids to arachidonic acid. NSAIDs inhibit cyclooxygenase, therefore preventing the production of prostaglandins from arachidonic acid. Piprants selectively antagonise the prostaglandin-E2 EP4 receptor. Adapted from Monteiro and Steagall (2019).

Conflict of Interest Statement

No conflicts of interests have been declared.