

## Understanding human immunity in idiopathic Recurrent Pregnancy Loss

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### Abstract

Miscarriage, defined as the loss of a pregnancy before a viable gestation, affects 1 in 6 couples. Recurrent pregnancy loss (RPL), defined as two or more miscarriages, affects up to 1.9% of couples. The physical, psychological, and financial impact of miscarriage can be substantial. However, despite its multifactorial etiology, for up to 50% of couples a reason behind this condition cannot be identified, termed 'idiopathic RPL'. Much recent research has strived to understand this, with immune dysregulation being a source of particular interest. In this short review we summarize the current evidence on the complex role of the immune system both pre- and early post-conception in RPL. A key question is whether systemic peripheral blood markers, in particular natural killer cell and T cells, may be utilized to accurately predict and/ or diagnose those pregnancies at high risk of loss. Given the invasive nature of endometrial testing, identification of reliable peripheral immune biomarkers is particularly appealing. Clinical trials using potent immunomodulatory agents, including intravenous immunoglobulin, donor leukocyte immunization, and tumor necrosis factor (TNF)- $\alpha$  inhibitors, have been undertaken with the primary objective of preventing miscarriage in women with RPL. Standardisation of both diagnostic and prognostic immune cell testing assays is required to permit accurate identification of those women who may benefit from immunomodulation. Prompt clarification is required to meet the increasing expectation from couples and clinicians, as without these advancements women are at risk of exposure to potent immune-therapies and subsequent studies are at risk of failure, generating further controversy regarding the role of immune dysregulation in women with RPL. Through this review we highlight clear gaps in our current knowledge on immune activity in RPL.

**Keywords:** Recurrent pregnancy loss, miscarriage, pregnancy, immune system, natural killer cells

## Introduction

Miscarriage is defined in the UK as the spontaneous loss of a fetus prior to the 24<sup>th</sup> gestational week(1, 2). Recurrent pregnancy loss (RPL), also termed recurrent miscarriage (RM), defined by the European Society of Human Reproduction and Embryology as two previous losses, affects 0.7%-1.9% of women(2, 3). Importantly, women with are are at increased risk of serious obstetric complications, including fetal growth restriction, preterm birth, and stillbirth(2). The potentially serious psychological impact of this condition is also highly significant, with significant risk of post-traumatic stress disorder, anxiety, and depression following RPL(2, 4, 5).

RPL is multi-factorial, with a range of contributory factors now understood as summarized in Table 1. For ~50% of women, the underlying pathogenesis of RPL is however unknown and the condition is termed 'idiopathic' (6, 7). Furthermore, even though 'idiopathic' RPL is defined by the lack of an identifiable underlining reason, it remains unclear whether idiopathic and non-idiopathic cases are exclusively underpinned by discrete pathogenic mechanisms. For the purpose of this review, 'idiopathic RPL' describes couples for which underlying maternal comorbidities and/or genetic aberration has been excluded prior to, or alongside immunological investigation. We discuss current revised concepts regarding idiopathic RPL, with immune dysregulation at the forefront of much recent research activity.

Table 1. Factors contributing to the etiology of recurrent pregnancy loss (RPL)

Factors contributing to RPL etiology
Maternal demographics (age>35 years, higher order loss, and obesity)(8, 9)
Environmental (smoking, alcohol consumption, high caffeine intake, cocaine use, organic solvent exposure, vitamin D insufficiency)(10-13)
Maternal comorbidities (e.g. hereditary and acquired thrombophilia, including antiphospholipid syndrome, thyroid disease, diabetes mellitus, and infections, such as <i>chlamydia trachomatis</i> , <i>ureaplasma</i> , and <i>mycoplasma</i> , which can potentially induce chronic endometritis)(14-20)
Genetic abnormalities (parental chromosomal abnormalities in 5.7% of couples with RPL and fetal aneuploidy in cases of early spontaneous miscarriages)(21, 22)
Male factors (microdeletions of the Y chromosome and sperm DNA fragmentation may be involved in the etiology of RPL, although routine analysis is not currently recommended)(1, 3, 23)

In human pregnancy, immune cell activity appears highly dynamic with a plethora of pro- and anti-inflammatory responses contributing to many critical reproductive processes including implantation, trophoblast invasion, spiral artery remodeling, fetal growth, and parturition(24, 25). The "implantation checkpoint hypothesis" suggests that multiple immune checkpoints exist at distinct menstrual cycle stages, highlighting the need for an endometrium that is both receptive and selective for embryo implantation(7). Aberrant immune activity in RPL may result in an excessively receptive, but poorly selective, endometrium, thereby permitting implantation of "low-quality" embryos with impaired growth and survival potential(7, 26). On the other hand, improper decidualization, potentially due to aberrant progesterone signalling, could also result to an immune environment unable to support "high-quality" embryo implantation and development(7, 27).

The immunological dialogue occurring at the fetal-maternal interface within the endometrium and later the maternal decidua is fascinatingly complex involving a diverse array of immune cells and mediators. Immune cell recruitment appears important, with the maternal decidua enriched with a unique innate and adaptive leukocyte infiltrate; comprising 70% uterine Natural Killer cells (uNKs), 20%-30% macrophages, and 3%-10% T Cells(28-30). Decidual immune cells actively interact with invasive fetal-derived extravillous trophoblast cells (EVTs)(28), which are negative for Human Leukocyte Antigen (HLA)-A, -B, and -D and HLA class II, which likely permits protection from T cell-mediated cytotoxicity. Positive expression of HLA-C, HLA-E, and HLA-G, thereby also facilitates protection from NK cell-mediated attack through inhibitory Killer-cell Immunoglobulin-like (KIR), Natural Killer Group (NKG)2, and Immunoglobulin-Like Transcript (ILT) receptor-mediated interactions(28, 31-37). Importantly, fetal-maternal interactions are measurable in peripheral maternal blood, with HLA-negative fetal syncytiotrophoblast cells shown to interact with circulating maternal immune subsets during hemochorial placentation(38-40). Although these cells are possibly subject to NK cell killing, defensive mechanisms such as soluble HLA-G secretion have been postulated to control peripheral NK (pNK) responses(41, 42).

Given the difficulties of invasive uterine sampling, a key question is whether dysregulated uterine immune cell activity is reflected by peripheral immune cell frequencies and function pre-conception and/or during early pregnancy. The relationship between endometrial and decidual uNK subsets and their peripheral counterparts remains of great interest to those seeking reliable biomarkers/prognostic tests. Different theories regarding uNK origin, including pNK recruitment to the decidua and de novo development of decidual NK (dNK) cells from endometrium-resident progenitors, exist(43). This is similarly true for adaptive subsets, with modifications in CD4<sup>+</sup> and CD8<sup>+</sup> T cell populations measured in the uterus compared to peripheral blood(44). Whether cytokine secretion and activity differ between the uterus and periphery has again generated much interest, with significant efforts to characterise differences in both healthy and high-risk reproductive cohorts(45, 46). As shall be discussed, high variation in the methodology and eligibility criteria in previous reports has almost certainly contributed to conflicting data, thereby generating questions regarding the significance of immunological alterations observed RPL. In this review, we aim to present current emerging concepts regarding the phenotype and function of specific innate and adaptive immune cell subsets in normal pregnancy and RPL, as summarised in text and tabular forms. This includes evidence regarding more novel immune subsets which may generate future interest. We also present a current overview of immunological tests and potential therapeutic strategies.

## **Innate immunity in RPL**

### NK cells

#### *NK cell numbers*

Due to their high prevalence in the uterine lymphocyte population, reaching up to 90%, NK cells have been classically associated with reproductive success(47), with key roles in early uterine artery remodeling, trophoblast invasion and placentation identified(28). It is however postulated that uncontrolled NK cell endometrial recruitment and/or failed CD56<sup>dim</sup> cell conversion to less cytotoxic CD56<sup>bright</sup> cells may occur in women with RPL. Increased anti-fetal-placental cytotoxicity and more intense induction of angiogenesis may confer a deleterious, high oxidative stress environment(48). Although an increase in both pNK and uNK cells, in particular cytotoxic CD56<sup>dim</sup> ones, has been linked to RPL, the predictive accuracy of measuring NK cell titers remains unclear with significantly heterogenic findings obtained (Table 2). These differences could simply reflect cohort or methodological inconsistencies, but it is also possible that NK cell elevation is only present in some women with idiopathic RPL. A systematic review on the prognostic value of pNK and uNK cell testing in regards to pregnancy outcome in women with infertility and RPL concluded that lack of a consensus on the definition of “normal”/“abnormal” NK cell test results is a primary factor contributing to these heterogenic findings(49).

#### *NK cell cytotoxicity*

NK cell activity also appears dysregulated in women with RPL, although current evidence is conflicting with both reduced and increased cytotoxicity reported(50, 51)(Table 2). Elevated cytotoxicity could reflect a local anti-fetal immune activity but also a systemic response to increasing circulating volumes of fetal-derived trophoblast microparticles. Conversely, reduced pNK and uNK cytotoxicity could indicate a possible association between early pregnancy loss and defective decidual CD56<sup>+</sup> leukocyte activity(52). The cytotoxic activity of uNK cells in women with RPL has not been thoroughly examined, possibly due to difficulties in isolating these cells without affecting their activation status. Additional experiments and alternative markers to more conclusively define NK cell cytotoxicity are required to establish the validity of these heterogenic findings.

#### *NK cell cytokine production*

Although most studies have utilized NK cell cytotoxicity as a marker of NK cell activity, cytokine production may also be important. During pregnancy, a local and peripheral anti-inflammatory, pro-regulatory shift in NK cell cytokine production is described and likely contributes to pregnancy maintenance. In women with RPL, even prior to conception, NK cytokine secretion appears dysregulated (Table 2). Of particular interest are IFN- $\gamma$  and TNF- $\alpha$ , both of which may exert key roles in human pregnancy. IFN- $\gamma$  is considered important for pregnancy establishment, being involved in the decidual vessel remodeling, while a protective role of TNF- $\alpha$  has also been displayed(53), with control of trophoblast invasion described(54). Albeit important during initial implantation, excessive or imbalanced cytokine production may be deleterious, with dysregulated levels postulated as important within the context of RPL. Overall, evidence supporting the use of cytokine profiling in women with RPL is highly heterogenic and is not routinely recommended in current clinical practice(55).

Table 2. Evidence of the role of Natural Killer (NK) cells in human pregnancy and recurrent pregnancy loss (RPL). Theories regarding the alterations in NK cell numbers, cytotoxicity, and cytokine production in normal pregnancy and RPL are displayed separately in the respective sections of the table. CD: cluster of differentiation, EVT: extravillous trophoblast cells, IFN- $\gamma$ : interferon-gamma, IL: interleukin, pNK: peripheral NK, TGF- $\beta$ : transforming growth factor-beta, TNF- $\alpha$ : tumor necrosis factor-alpha, uNK: uterine NK

<b>Natural Killer (NK) cells</b>	
NK cells accumulate in the luteal phase endometrium and early-pregnancy decidua, progressively disappearing from mid-gestation onwards, being absent at term(28, 56) → potential role in preparation for and during early pregnancy	
Two main subpopulations present in both peripheral blood and uterus(57-60): • CD56 <sup>dim</sup> CD16 <sup>+</sup> : Major circulatory subset; primarily cytolytic activity • CD56 <sup>bright</sup> CD16 <sup>-</sup> : Most prevalent in uterus; lower cytotoxic ability and a higher capacity for cytokine production	
<b>Normal pregnancy:</b>	
<ul style="list-style-type: none"> <li>• Dense uNK cell infiltrate surrounds fetal trophoblasts, coinciding with trophoblast invasion(28, 56, 61)</li> <li>• uNK cells do not kill EVTs(62, 63)</li> <li>• NK cells' activating receptor interaction with their ligands on trophoblasts results in the production of growth and angiogenic factors, cytokines, chemokines, and matrix metalloproteinases(64-67)</li> </ul>	
Key roles in: A) angiogenesis and spiral artery remodeling, B) trophoblast growth, differentiation, and invasion, C) placental tissue building and remodeling <u>But</u> NK cell function in pregnancy remains unclear	
<b>NK cell numbers</b>	
<b>Normal pregnancy:</b>	<b>Women with RPL:</b>
<ul style="list-style-type: none"> <li>• 1<sup>st</sup> and 2<sup>nd</sup> trimester compared to non-pregnant: No difference in pNK cell numbers(68)</li> </ul>	<ul style="list-style-type: none"> <li>• Pre-conception and during pregnancy: <math>\uparrow</math> pNK cells(69-73)</li> <li>• In pregnant and non-pregnant (incl. idiopathic): <math>\uparrow</math> CD56<sup>dim</sup> and thus <math>\downarrow</math> CD56<sup>bright</sup>/CD56<sup>dim</sup> pNK cell ratio(71, 72)</li> <li>• <u>But</u> unaltered pNK cell numbers and CD56<sup>bright</sup>/CD56<sup>dim</sup> NK ratio(74)</li> </ul>
	$\uparrow$ Peripheral blood CD56 <sup>+</sup> , CD56 <sup>dim</sup> , and CD56 <sup>bright</sup> cell levels measured as predictive biomarker for subsequent loss(71, 75-77)
<ul style="list-style-type: none"> <li>• Early pregnancy: significant NK cell infiltration in the uterus(28, 61)</li> </ul>	<ul style="list-style-type: none"> <li>• In pregnant and non-pregnant (incl. idiopathic): <math>\uparrow</math> uNK cell infiltration(78, 79)</li> <li>• In non-pregnant women with idiopathic miscarriage: <math>\uparrow</math> CD56<sup>dim</sup> and <math>\downarrow</math> CD56<sup>bright</sup> uNK cell frequencies(78, 80)</li> <li>• <u>But</u> meta-analysis found no significant change in uNK cell frequencies(69)</li> </ul>
<b>NK cell cytotoxicity</b>	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
1 <sup>st</sup> to 3 <sup>rd</sup> trimester: $\uparrow$ activated CD56 <sup>bright</sup> pNK cell percentage(81)	<ul style="list-style-type: none"> <li>• Pregnant and non-pregnant: <math>\uparrow</math> pNK cell cytotoxicity(50, 70, 73)</li> <li>• <math>\uparrow</math> Activated pNK cell levels(72, 82, 83)</li> <li>• <u>But</u> in pregnant and non-pregnant: unaltered(84) or <math>\downarrow</math>(85) pNK cell cytotoxicity</li> </ul>
	$\uparrow$ Pre-conception pNK cell activity suggested as a prognostic miscarriage marker(86)
	• In pregnant: $\uparrow$ activated CD56 <sup>+</sup> uterine cell levels(87)
<b>NK cell cytokine production</b>	
NK cells secrete Type-1 (TNF- $\alpha$ and IFN- $\gamma$ ), Type-2 (IL-5 and IL-13), and immunosuppressive (IL-10 and TGF- $\beta$ ) cytokines(88): • Type-1 cytokine-secreting pNK cells are the major subpopulation at the non-pregnant state	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
<ul style="list-style-type: none"> <li>• <math>\downarrow</math> IFN-<math>\gamma</math> production by pNK cells(89)</li> <li>• <math>\uparrow</math> IL-10<sup>+</sup> CD56<sup>dim</sup> and CD56<sup>bright</sup> pNK cell levels(88)</li> </ul>	<ul style="list-style-type: none"> <li>• In non-pregnant women with idiopathic RPL: <math>\uparrow</math> IFN-<math>\gamma</math>+TNF-<math>\alpha</math> and <math>\downarrow</math> IL-4<sup>+</sup>IL-10<sup>+</sup> CD56<sup>bright</sup> pNK cells(74)</li> </ul>
<ul style="list-style-type: none"> <li>• High levels of TGF-<math>\beta</math><sup>+</sup> CD56<sup>dim</sup> and CD56<sup>bright</sup> uNK cells(88)</li> </ul>	<ul style="list-style-type: none"> <li>• <u>But</u> in non-pregnant women with idiopathic RPL: <math>\downarrow</math> prevalence of TNF-<math>\alpha</math><sup>+</sup> and/or IFN-<math>\gamma</math><sup>+</sup> uNK cells(90)</li> </ul>

### *NK cell memory*

Even though NK cells are considered an innate immune subset, an ability to form immunological memory (mNK) has been displayed [Reviewed in (91)]. mNK cells have been most extensively studied in the context of cytomegalovirus (CMV) infection, where they persist after initial infection and display an enhanced antigen-specific response upon secondary challenge(92-94). Antigen-specific memory responses have been exhibited by liver-resident NK cells(95), while cytokine-induced mNK cells were also described(96).

It is now understood that the risk of a subsequent miscarriage increases with each subsequent loss(2). It has been hypothesised that during an index pregnancy, NK cell “priming” occurs in the decidua, resulting in a more effective response in any subsequent pregnancy(97). Pregnancy-specific mNK cell generation independent of CMV-induced alterations has been suggested(98). Even though the origin and role of these cells remains to be elucidated, pregnancy-trained mNKs may be generated in response to signals in the uterine microenvironment during the first pregnancy, with more effective production of VEGF $\alpha$  and IFN- $\gamma$  to support local spiral artery remodelling and angiogenesis in subsequent pregnancies(65, 99, 100). Whether aberrant mNK responses are linked to pregnancy loss and RPL is yet to be elicited but of certain interest.

### Antigen-presenting cells

Dendritic Cells (DCs) and macrophages, have a key role in antigen recognition and presentation and are involved in the orchestration of lymphocyte-mediated immunity. There is now evidence to suggest peripheral and decidual DCs may support pregnancy success (Table 3). For instance, prior to trophoblast invasion peripheral DCs may promote the overall inflammatory stage required for pregnancy establishment without inducing anti-fetal immunity. Due to their very low frequency in the uterus, the role of DCs in reproductive success has not been extensively studied and thus any potential role in RPL is not known.

Monocytes and macrophages are also gaining increased research interest. Macrophages, which mainly arise from circulating monocytes upon tissue migration(101), represent one of the major effector immune populations. Using a three-dimensional *in vitro* system, monocyte/macrophage recruitment in response to trophoblast signals has now been displayed, indicating a possible mechanism for monocyte migration from the peripheral blood to the uterus during pregnancy, enhancing the local macrophage population(24). Characteristically, macrophages are recognised for their ability to phagocytose dead and potentially harmful cells, bacteria, and debris(101). During menstruation, key roles in tissue degradation, clearance of local debris, and possibly endometrial regeneration and angiogenesis have been shown(102). Although the exact function of macrophages in pregnancy appears complex, with gestation-specific pro- and anti-inflammatory functions reported, establishment of early blastocyst implantation is amongst those actions described(27, 103)(Table 3). Dysregulated macrophage activity has been reported in other malplacental disorders, like preeclampsia and preterm birth(104). Whether alterations in monocyte or macrophage numbers or activity contribute to immune dysregulation in idiopathic RPL remains to be explored.

Table 3. Evidence of the role of antigen-presenting cells (APCs) in human pregnancy and recurrent pregnancy loss (RPL), including information on the numbers and activity of dendritic cells (DCs) and macrophages (Mφs), displayed in the respective sections of the table. CCR2: C-C chemokine receptor type 2, CD: cluster of differentiation, EVT: extravillous trophoblast cells, FasL: Fas ligand, HLA: human leukocyte antigen, IL: interleukin, MMPs: matrix metalloproteinases, uNK cells: uterine Natural Killer cells

<b>Antigen-presenting cells (APCs)</b>	
<b>Dendritic cells (DCs)</b>	
Low frequency in endometrium and early pregnancy decidua (~1.7% of decidual leukocytes)(105, 106)	
<b>Normal pregnancy:</b>	<b>In the peripheral blood of women with RPL:</b>
1 <sup>st</sup> and 2 <sup>nd</sup> trimester: ↑ peripheral blood DCs compared to the non-pregnant state(107, 108)	In women with miscarriage(107): • ↓ DC capacity for cytokine production • ↑ Numbers of mature DCs
Peripheral blood DCs(107, 108): • ↑ Mature and activated DCs • <u>But</u> a state of incomplete DC activation has been suggested, potentially leading to defects in T cell activation through antigen presentation	During the later stage of fetal growth DC activity could promote anti-fetal T cell-mediated responses, through antigen presentation, and thus pregnancy loss(109)
Decidual DCs possibly adopt an immature tolerogenic profile, possessing a decreased T cell stimulatory capacity(106, 110)	<b>In the decidua of women with RPL:</b>
	1 <sup>st</sup> trimester: mature DC decrease observed during the 7 <sup>th</sup> -11 <sup>th</sup> week of healthy pregnancy not detected, suggesting a potential association between RPL and mature DC activity(111) Defective senescent DC clearance by uNK cells during decidualization could be associated with RPL(112)
<b>Macrophages (Mφs)</b>	
<b>Normal pregnancy:</b>	<b>In the uterus of women with RPL:</b>
Second largest immune cell population in early pregnancy decidua (20%-30% of leukocytes)(30)	In idiopathic spontaneous miscarriage(113) but not RPL(114): ↑ decidual Mφs
Prior to EVT invasion: Mφs may cooperate with uNK cells, mediating the disruption and disorganization of endothelial and vascular smooth muscle cells, and thus contributing to vascular remodeling through the secretion of MMPs and the phagocytosis of apoptotic cells(115)	Endometrial Mφs ↑ in women with idiopathic RPL and subsequent miscarriage(78)
• M2 phenotype is considered as essential during pregnancy(104, 116), with immunomodulatory agents like IL-10 being expressed by decidual Mφs(97). • <u>But</u> M1/M2 dichotomy appears an oversimplification, with the balance of different pro- and anti-inflammatory Mφs likely dependent upon the stage of pregnancy(117). Shifts in Mφ activity during pregnancy(104, 117): a) Peri-implantation period: pro-inflammatory M1 activity b) Placental development: anti-inflammatory M2 predominance c) Prior to parturition: pro-inflammatory M1 predominance	Polarization towards the pro-inflammatory M1 phenotype may promote pregnancy complications(104, 116)
Tolerogenic M2 Mφ recruitment and polarization is thought to be influenced by hormones like estrogen and progesterone(116)	
FasL expression by Mφs possibly mediates apoptosis of Fas <sup>+</sup> activated T cells, eliminating T cell-mediated anti-fetal immunity and controlling trophoblast invasion(113)	↑ FasL expression by Mφs could promote excessive trophoblast apoptosis, leading to pregnancy loss(113, 118)

## **Adaptive immunity in RPL**

### T cells

T cells constitute a central component of adaptive immunity with both cytotoxic and immunomodulatory potential. In the peripheral blood and decidua, T cells are thought to acquire an important role in fetal antigen recognition and modulation of local immunity (Table 4). As shall be discussed, enhanced activity of these cells and a shift in the balance between T cell subsets may be involved in miscarriage pathogenesis.

#### *Cytotoxic T cells*

Cytotoxic T (Tc) cells are central in the defense against intracellular pathogens. They are capable of killing infected and malignant cells through multiple mechanisms, including cytokine secretion, cytotoxic granule release, and death receptor interactions(119). The exact role of Tc cells in pregnancy is unclear. Although the absence of HLA-A, -B, and -D expression on EVT's highlights a potential mechanism to evade Tc-mediated cytotoxicity, HLA-C expression possibly permits trophoblast recognition by Tc cells(120). Tc cells at the fetal-maternal interface could be less cytotoxic, facilitating local inflammatory processes such as implantation, without attacking the fetus. Heterogenic data has to date been collected in regards to Tc cell involvement in RPL (Table 4), hence, the value of measuring T cell frequencies and activity in RPL currently remains limited with further research required.



Table 4. Evidence of the role of total and CD8<sup>+</sup> cytotoxic T (Tc) cells in human pregnancy and recurrent pregnancy loss (RPL). Information about total and cytotoxic T cells are displayed separately, in the respective table sections. APCs: antigen-presenting cells, CD: cluster of differentiation, EVT: extravillous trophoblast cells, GM-CSF: granulocyte-macrophage colony-stimulating factor, HLA: human leukocyte antigen, NK cell: Natural Killer cell, PD-1: programmed cell death protein 1, Tim-3: T-cell immunoglobulin and mucin-domain containing-3

<b>T cells</b>	
Abundant in endometrium(46) with constant numbers throughout menstruation(121)	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
Unaltered prevalence in peripheral blood(122)	<ul style="list-style-type: none"> <li>• ↑ Peripheral T cell activation(123, 124)</li> <li>• <u>But</u> no significant alterations in T cell levels in the peripheral blood, endometrium, or decidua(78, 114, 125)</li> </ul>
In uterus during early pregnancy: constant absolute numbers but ↓ prevalence due to NK cell surge(121)	Predictive value: <ul style="list-style-type: none"> <li>• ↓ Peripheral T cell numbers in women with subsequent miscarriage(127)</li> <li>• ↑ Endometrial CD4<sup>+</sup>/CD8<sup>+</sup> T cell ratio in women with idiopathic RPL and subsequent miscarriage(80)</li> </ul>
↑ in uterus throughout pregnancy(126): <ul style="list-style-type: none"> <li>• 1<sup>st</sup> trimester: 5%-20% of lymphocytes</li> <li>• Late pregnancy: 40%-80% of lymphocytes</li> </ul>	
<b>CD8<sup>+</sup> cytotoxic T (Tc) cells</b>	
Major immune effector subset, being capable of killing infected and tumor cells through a variety of mechanisms, including cytolytic granule formation, cytokine secretion, and death receptor interactions	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
<ul style="list-style-type: none"> <li>• Activation marker expression by uterine Tc cells suggests fetal antigen recognition(128)</li> <li>• Fetal antigen-specific Tc cells detected in maternal periphery(129)</li> <li>• Recognition of the fetus directly through HLA-C expression on fetal EVTs and indirectly through maternal APCs(120)</li> </ul>	<ul style="list-style-type: none"> <li>• ↑ Peripheral Tc cell levels and ↓ CD4<sup>+</sup>/CD8<sup>+</sup> T cell ratio(130)</li> <li>• <u>But</u> ↓ peripheral Tc cell levels and unchanged CD4<sup>+</sup>/CD8<sup>+</sup> cell ratio(73)</li> </ul>
Trophoblast antigen-specific tolerance may be induced via upregulation of death receptors on maternal Tc cells(120, 131): <ul style="list-style-type: none"> <li>• ↑ Tim-3<sup>+</sup>PD-1<sup>+</sup> Tc prevalence in decidua compared to peripheral blood implying partial local suppression of Tc cell activity</li> </ul>	<ul style="list-style-type: none"> <li>• Non-pregnant women with idiopathic RPL: ↑ activated Tc prevalence in peripheral blood(132)</li> <li>• <u>But</u> in non-pregnant women with idiopathic RPL: unaltered peripheral Tc activation status(133)</li> </ul>
<u>But</u> Tc lymphocytes may contribute to implantation via the secretion of colony-stimulating factors (incl. GM-CSF)(134)	

### T helper cell cytokine profiling

The balance between the T helper (Th)1 and Th2 cytokines is considered important in guiding the immune responses during pregnancy. Direct effects of Th cell-associated cytokines in fetal development and survival have been described. Most cytokines are growth and/or differentiation factors, and thus could encourage fetal growth(135). As summarized in Table 5, a “Type-1/Type-2” hypothesis in both healthy pregnancy and miscarriage has been suggested, with a dominant Type-2 anti-inflammatory cytokine production contributing to pregnancy success, and a pro-inflammatory Type-1 ‘anti-fetal’ shift contributing to pregnancy loss, with overproduction of IFN-γ and TNF-α shown to induce aberrant trophoblast apoptosis(54, 136). This appears an over-simplification however, since Th1 cytokines, including IFN-γ and TNF-α, also have key roles in local spiral artery remodeling (53, 100), and fetal protection (137). Further large, well-designed studies will be required to understand the significance of this shift (Table 5). It is possible a dynamic cytokine balance may be important, with the effects of individual pro- and anti-inflammatory cytokines dependent upon their relative concentration and the gestation stage(54, 138, 139). Strategies to modulate cytokine balance, like progesterone, could represent a valuable option to protect against RPL(140), but mechanistic data is required to confirm this.

### *Regulatory T cells*

Regulatory T cells (Tregs) comprise a Th cell subpopulation, considered as a central immunosuppressive component of the adaptive immune system. A vital role for Tregs in pregnancy, particularly in the second trimester, has been suggested(141)(Table 5). Tregs may regulate excessive inflammation and induce fetal antigen-specific tolerance from early pregnancy(120). Experimental evidence indicates that defects in peripheral and decidual Treg numbers or activity could be linked to the pathogenesis of RPL, although the role of individual Treg subtypes requires further examination(142-144).

Table 5. Evidence of the role of CD4<sup>+</sup> T helper (Th) cells, including regulatory T cells (Tregs), in human pregnancy and recurrent pregnancy loss (RPL). Information about the different T cell subtypes are displayed in the respective table sections. CD: cluster of differentiation, GM-CSF: granulocyte-macrophage colony-stimulating factor, HLA: human leukocyte antigen, IFN- $\gamma$ : interferon- $\gamma$ , IL: interleukin, M $\phi$ : macrophage, NK cell: Natural Killer cell, TGF- $\beta$ : transforming growth factor- $\beta$ , TNF: tumor necrosis factor

<b>T cells</b>	
<b>CD4<sup>+</sup> T helper (Th) cell cytokine profiling</b>	
Four major subsets(45, 135, 145, 146): <ul style="list-style-type: none"> <li>• Th1: secrete pro-inflammatory cytokines (incl. IL-1, IL-2, IFN-<math>\gamma</math>, TNF-<math>\alpha</math>, and TNF-<math>\beta</math>) and stimulate cellular immunity</li> <li>• Th2: secrete anti-inflammatory cytokines (incl. IL-4, IL-5, IL-10, IL-12, and IL-13) and activate humoral immunity</li> <li>• Th17: secrete pro-inflammatory cytokines (incl. IL-17, GM-CSF, IL-21, and IL-22)</li> <li>• Tregs: characterized by anti-inflammatory IL-10 secretion (discussed separately)</li> </ul>	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
Unaltered prevalence in peripheral blood (~40% of lymphocytes)(142)	Idiopathic RPL(132, 133): <ul style="list-style-type: none"> <li>• <math>\uparrow</math> Peripheral activated Th cell prevalence</li> <li>• <u>But</u> appeared anergic</li> </ul>
	Th shift in the endometrium: ~11% of local cells in women with RPL, while ~3% in healthy women(78, 80)
<ul style="list-style-type: none"> <li>• "Th1/Th2 hypothesis": Shift towards Th2-mediated responses, accompanied by <math>\downarrow</math> Th1 cell function, reflected by a cytokine expression shift in the periphery and placental tissue(135, 147, 148)</li> <li>• Extended into the "Type-1/Type-2 hypothesis" representing the balance between pro-inflammatory Th1 and Th17 (Type-1) and anti-inflammatory Th2 and Treg (Type-2) activity</li> </ul>	"Th1/Th2 hypothesis": shift towards Th1 responses, possibly directed against paternal trophoblast antigens(54, 147-151) <ul style="list-style-type: none"> <li>• In peripheral blood of pregnant and non-pregnant: <math>\uparrow</math> Type-1 cytokine production (incl. IL-1<math>\beta</math>, IL-12, IL-17, IL-18, IL-23, IFN-<math>\gamma</math>, TNF-<math>\beta</math>, and TNF-<math>\alpha</math>)(147, 148, 150-153)</li> <li>• In the peripheral blood and decidua: <math>\downarrow</math> Type-2 cytokine secretion, including low IL-10(148-152)</li> </ul>
	Association with IL-10 promoter polymorphisms and polymorphisms linked to $\uparrow$ IFN- $\gamma$ and TNF- $\alpha$ production(154)
Peripheral blood(153): <ul style="list-style-type: none"> <li>• <math>\uparrow</math> IL17<sup>+</sup> Th cell prevalence in the 1<sup>st</sup>, compared to the 2<sup>nd</sup> trimester</li> <li>• <u>But</u> unaltered IFN-<math>\gamma</math><sup>+</sup> Th cell prevalence throughout pregnancy</li> </ul>	<ul style="list-style-type: none"> <li>• Pre-pregnancy: <math>\uparrow</math> TNF-<math>\alpha</math><sup>+</sup>/IL-10<sup>+</sup>, TNF-<math>\alpha</math><sup>+</sup>/IL-4<sup>+</sup>, and IFN-<math>\gamma</math><sup>+</sup>/IL-4<sup>+</sup> ratios(155)</li> <li>• During the 1st trimester and at the time of miscarriage: pro-inflammatory shift in Type-1/Type-2 cytokines ratios(150, 151)</li> <li>• <u>But</u> in other similarly powered studies: no significant alterations in IL-12, IL-4, IL-13, TGF-<math>\beta</math>, and IFN-<math>\gamma</math> concentrations(122, 152)</li> </ul>
Peripheral blood: $\downarrow$ IFN- $\gamma$ and TNF- $\alpha$ and $\uparrow$ IL-10 level linked to pregnancy success(156)	$\uparrow$ Type-1 cytokine production may promote T cell-, NK cell-, and M $\phi$ -mediated cytotoxicity against fetal cells(45, 157, 158). This can be controlled by Type-2 cytokines(45, 54).
<b>Regulatory T cells (Tregs)</b>	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
From the early pregnancy: Tregs may regulate excessive inflammation and induce fetal antigen-specific tolerance(120)	During miscarriage: $\downarrow$ decidual Treg concentrations(142-144)
Early pregnancy: $\uparrow$ peripheral blood Treg prevalence(141-143)	In healthy early pregnancy but not idiopathic RPL: Treg levels $\uparrow$ in decidua compared to peripheral blood(143, 144)
2 <sup>nd</sup> compared to 1 <sup>st</sup> trimester: $\uparrow$ peripheral blood Treg prevalence(141)	Pre-conception, 1 <sup>st</sup> trimester, and during miscarriage: $\downarrow$ peripheral blood Treg levels(142-144)
Treg recruitment and local differentiation is promoted by trophoblasts(159)	Predictive value: low peripheral blood Treg levels during early pregnancy linked to miscarriage(160)
Post-partum: $\downarrow$ Treg levels, agreeing with withdrawal of fetal-induced stimulation(141)	In peripheral blood CD4 <sup>+</sup> cell cultures of women with idiopathic RPL: $\downarrow$ Treg-mediated suppression of IL-17 production(161)

## T cell memory

Following the primary immunological response towards a pathogenic stimulus, a small percentage of T cells persists and differentiates into memory cells(162). As summarised in Table 6, central ( $T_{CM}$ ) and effector ( $T_{EM}$ ) T cell memory appears important in the establishment of tolerance to fetal-derived antigens(163). Further studies are required to understand the potential implication of T cell memory in healthy pregnancy and pregnancy-associated complications. It would be valuable to access the paternal antigen-specificity of these cells in first and subsequent pregnancies of the same and of different paternal backgrounds as well as examine their pro- or anti-inflammatory properties.

*Table 6. Evidence of the role of T cell memory in human pregnancy and recurrent pregnancy loss (RPL). CCR7: C-C chemokine receptor type 7, CD: cluster of differentiation, CTLA-4: cytotoxic T-lymphocyte-associated protein 4, FoxP3: forkhead box protein P3, IFN- $\gamma$ : interferon- $\gamma$ , IL: interleukin, LAG-3: lymphocyte-activation gene 3, PD-1: programmed cell death protein 1, Th cell: T helper cell, Tim-3: T-cell immunoglobulin and mucin-domain containing-3*

<b>T cells</b>	
<b>T cell memory</b>	
<ul style="list-style-type: none"> <li>• Central memory T (<math>T_{CM}</math>) cells: reside to the lymph nodes, exhibit limited effector function, and upon secondary antigen exposure they proliferate and differentiate into Th1 or Th2 cells under local cytokine influence(44, 162, 164)</li> <li>• Effector memory T (<math>T_{EM}</math>) cells: reside in peripheral tissues and upon secondary antigen exposure they rapidly respond through the secretion of perforin, IFN-<math>\gamma</math>, IL-4, and IL-5, displaying limited proliferative ability(44, 162, 164)</li> </ul>	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
Paternal antigen-specific memory T cell generation: possibly contribute to tolerance generation, although their exact contribution to pregnancy is unclear(129, 165)[Reviewed in (163)]	Unchanged memory-like CD4 <sup>+</sup> and CD8 <sup>+</sup> T cell frequencies in the peripheral blood(124)
Unaltered peripheral blood CD4 <sup>+</sup> $T_{CM}$ population(166-168) <ul style="list-style-type: none"> <li>• <math>\uparrow</math> after parturition, suggesting persistence of cells(167)</li> <li>• <math>\uparrow</math> activation during and after pregnancy(167)</li> </ul>	Non-pregnant: $\uparrow$ total and CD4 <sup>+</sup> $T_{CM}$ cells in peripheral blood(123, 169)
Unaltered peripheral blood CD8 <sup>+</sup> $T_{CM}$ population(166-168) <ul style="list-style-type: none"> <li>• Unaltered activation during and after pregnancy(167)</li> </ul>	
Peripheral blood CD4 <sup>+</sup> $T_{EM}$ population: <ul style="list-style-type: none"> <li>• <math>\uparrow</math> at the 2<sup>nd</sup> and 3<sup>rd</sup> trimesters and after parturition(166, 167)</li> <li>• <u>But</u> in one study found to be unaltered at the 3<sup>rd</sup> trimester(168)</li> <li>• <math>\uparrow</math> activation and reduced susceptibility to apoptosis(166, 167)</li> </ul>	Non-pregnant: $\uparrow$ total, but not CD4 <sup>+</sup> , $T_{EM}$ cells in peripheral blood(169)
Peripheral blood CD8 <sup>+</sup> $T_{EM}$ population: <ul style="list-style-type: none"> <li>• Unaltered during pregnancy but increased after parturition(166-168)</li> <li>• Unaltered activation and susceptibility to apoptosis(166, 167)</li> </ul>	
In pregnant women with a male fetus: $T_{EM}$ cells dominate the peripheral blood fetal HY-specific CD8 <sup>+</sup> T cell population, exhibiting a functional inflammatory response to HY peptide presentation(129)	
In the decidua: CD8 <sup>+</sup> cells compose the major T cell population(121) with $T_{EM}$ cells constituting the major CD8 <sup>+</sup> T cell subset(165)	Endometrial CD8 <sup>+</sup> memory T cells(170): <ul style="list-style-type: none"> <li>• Unaltered prevalence</li> <li>• Unaltered IFN-<math>\gamma</math> production upon stimulation</li> </ul>
Decidual $T_{EM}$ cells: <ul style="list-style-type: none"> <li>• Unusual cytokine profile (IFN-<math>\gamma^{high}</math>IL-4<sup>high</sup>) and <math>\downarrow</math> perforin and granzyme B expression compared to peripheral blood(44, 165)</li> <li>• High proliferative activity(131)</li> <li>• <math>\uparrow</math> CTLA-4, Tim-3, LAG-3, and PD-1 checkpoint protein expression(44, 165), indicating a strict control of local activity</li> </ul>	

## B cells

Despite the small uterine B cell numbers, dysregulation of humoral immunity may be associated with RPL. B cells are thought to contribute to pregnancy success through the downregulation of poly-reactive natural antibody secretion and the production of protective blocking asymmetric antibodies. B cells with regulatory activity and distinct memory subsets have also been described, with potential roles in reproductive success presented. Due to the small number of uterine B cell, the properties of these B cells have not been extensively studied (Table 7).

Table 7. Evidence of the role of B cells in human pregnancy and recurrent pregnancy loss (RPL), including the role of different subsets, antibodies, and B cell memory. Information about total B cells, B cell subtypes and their antibody production, and B cell memory are displayed in the respective table sections. AAb: asymmetric Ab, Ab: antibody, Bregs: regulatory B cells, CD: cluster of differentiation, IL: interleukin, NAb: natural antibody, RA: rheumatoid arthritis, SLE: systemic lupus erythematosus, Th: T helper

<b>B cells</b>	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
During pregnancy cell-mediated autoimmunity, like RA, is ameliorated, while antibody-mediated autoimmunity, such as SLE, is often aggravated(46, 171, 172), suggesting a role of humoral immunity in pregnancy	Pre-conception B cell frequencies in the peripheral blood were unaltered(123)
Endometrium and decidual: low B cell levels(173, 174)	↑ Endometrial CD20 <sup>+</sup> B cell levels(80)
<ul style="list-style-type: none"> <li>• The role of B cells in early pregnancy is unclear</li> <li>• In the 3<sup>rd</sup> trimester and during parturition peripheral B cell lymphopenia is reported(175), indicating an absence of B cell involvement in the later pregnancy stages</li> </ul>	↑ Endometrial B cell prevalence in women with RPL and a following 1 <sup>st</sup> -trimester miscarriage, compared to those with an intact conceptus at least until the 36 <sup>th</sup> week of their following pregnancy(80)
<b>B cell subsets and antibody (Ab) production</b>	
3 major B cell subsets(176-178):	
<ul style="list-style-type: none"> <li>• B-1 cells detected in peripheral and umbilical cord blood(179): produce low-affinity polyreactive, often autoreactive, NABs in the absence of exogenous immunization(177, 179, 180). Auto-Ab production is thought to be mediated by CD5<sup>+</sup> B-1 cells(181, 182), although CD5<sup>-</sup> NAb-producing B cells were also detected(183)</li> <li>• B-2 "conventional" cells: upon antigen recognition can differentiate into high-affinity Ab-secreting plasma or memory cells(184)</li> <li>• Bregs: mainly characterized by their IL-10 secretion(176, 185). IL-10-producing Bregs inhibit Th1 activation and Th17 differentiation and promote CD4<sup>+</sup> T cell conversion into Tregs, promoting tolerance(178, 186)</li> </ul>	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
B cell subsets have been poorly defined in the context of pregnancy	B cell subset number alterations not associated with miscarriage
↓ Circulating CD5 <sup>+</sup> B cell prevalence is during pregnancy, indicating a possible protective mechanism through ↓ NAb production(187)	Failure to limit NAb production has been linked to pregnancy complications but not RPL(176, 177)
Protective effect of maternally-derived anti-paternal Abs during pregnancy suggested(177): <ul style="list-style-type: none"> <li>• Pregnancy-protective Ab production by human term placenta-derived B cells upon stimulation reported(173).</li> <li>• Anti-paternal Ab AAbs, binding antigen but lacking the capacity for immune effector activation(176), could acquire a protective blocking role(188).</li> </ul>	<ul style="list-style-type: none"> <li>• Anti-paternal or anti-firstborn HLA Ab presence was linked to ↓ likelihood of subsequent live birth(189).</li> <li>• ↓ AAb production detected in the placenta and maternal serum in RPL, compared to normal pregnancy(177, 190-192)</li> </ul>
Bregs may contribute to the maintenance of tolerance during pregnancy through their IL-10 secretion(176).	Human Bregs not studied in the context of RPL
<b>B cell memory</b>	
Upon antigenic stimulation, a small B cell percentage forms a persisting memory population that upon secondary stimulation with the same antigen secretes large amounts of high-affinity, class-switched, monospecific antibodies(176). B cell memory is attributed to the B-2 subset(176, 184).	
<b>Normal pregnancy:</b>	<b>In women with RPL:</b>
<ul style="list-style-type: none"> <li>• 3<sup>rd</sup> trimester and at the day of delivery: ↓ peripheral blood memory B cell counts, compared to non-pregnant controls(175).</li> <li>• 6 weeks post-partum: ↑ memory B cell percentages in the peripheral blood, compared to the 3<sup>rd</sup> trimester(175).</li> </ul>	B cell memory not studied in the context of RPL

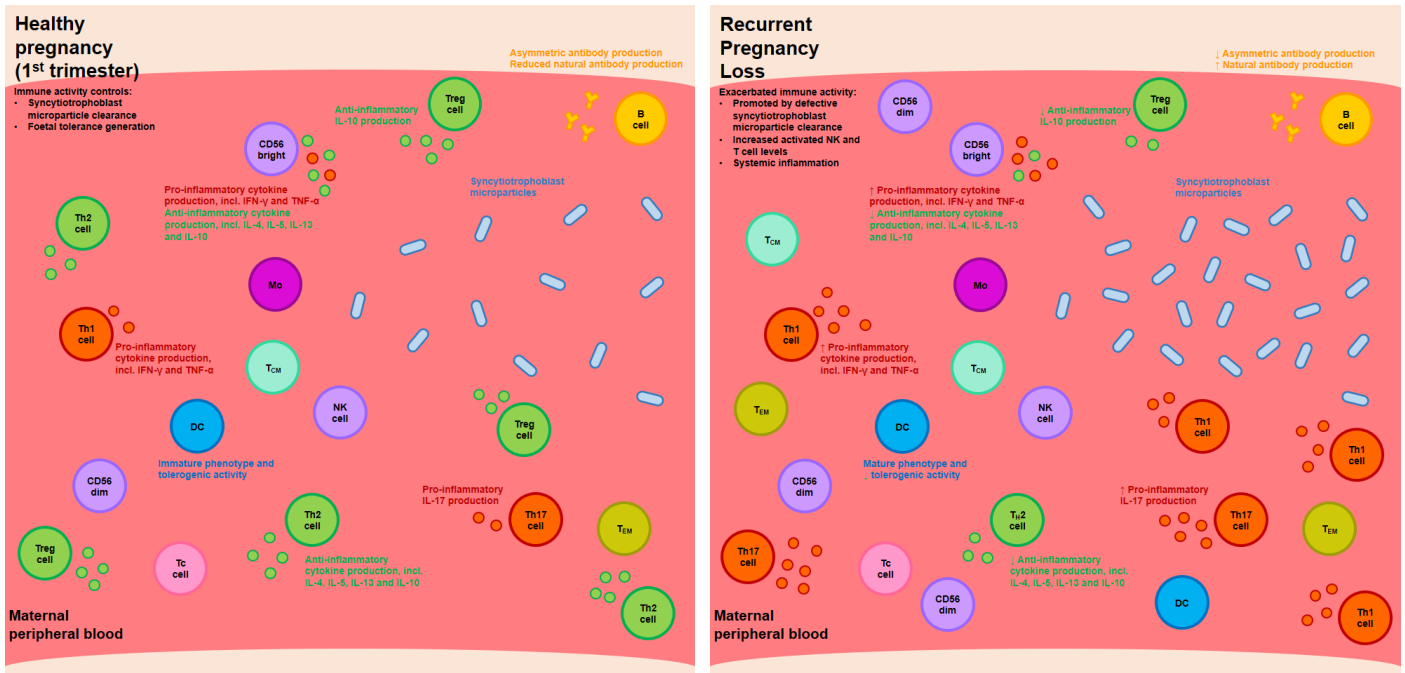


Figure 1. Overview of peripheral immunity during the first trimester in women with healthy pregnancy and women with recurrent pregnancy loss. CD: cluster of differentiation, DC: dendritic cell, IFN: interferon, IL: interleukin, Mo: monocyte, NK cell: natural killer cell, Tc cell: cytotoxic T cell, T<sub>CM</sub>: central memory T cell, T<sub>EM</sub> cell: effector

(coloured in print)

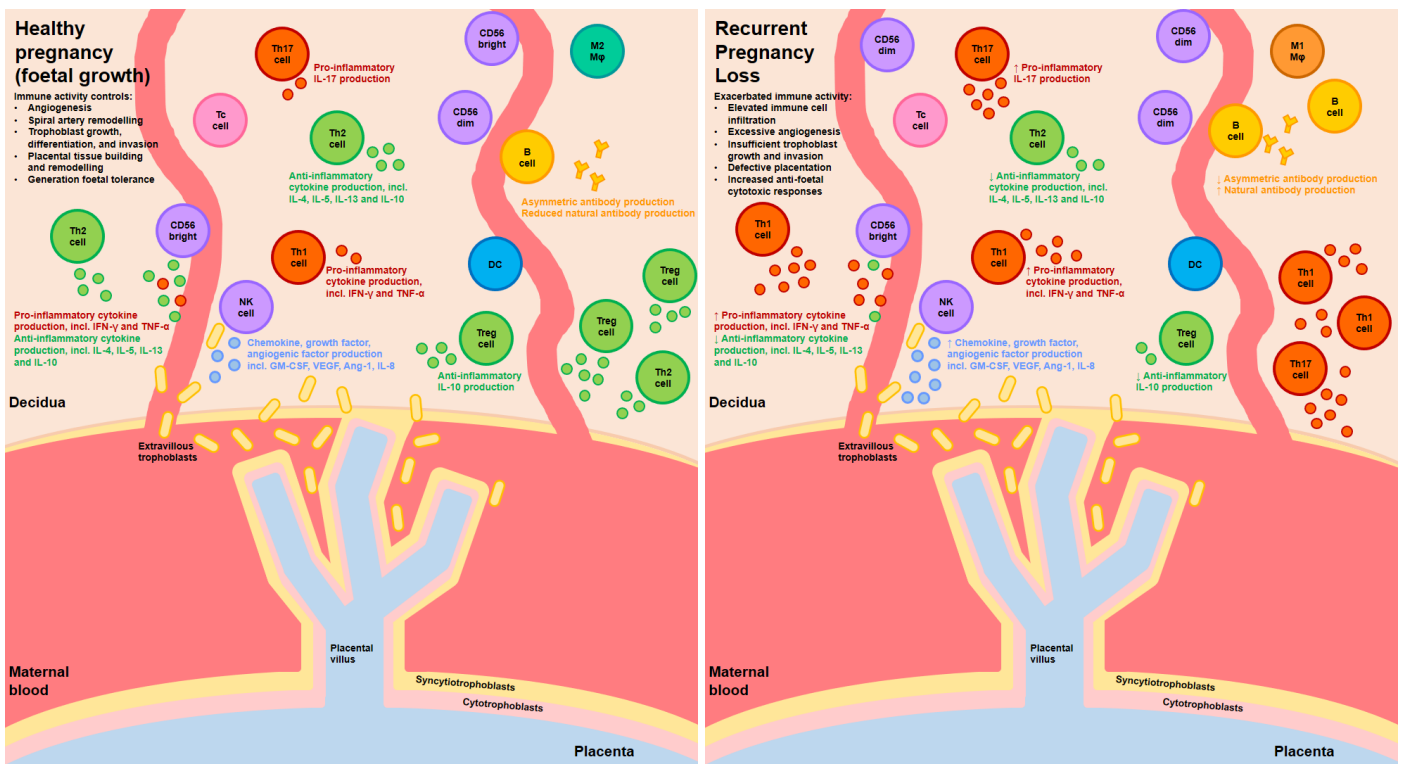


Figure 2. Overview of decidual immunity during healthy pregnancy, at the stage of foetal growth, and in women with recurrent pregnancy loss. Ang-1: angiopoietin 1, CD: cluster of differentiation, DC: dendritic cell, GM-CSF: granulocyte-macrophage colony-stimulating factor, IFN: interferon, IL: interleukin Mφ: macrophage, NK cell: natural killer cell, Tc cell: cytotoxic T cell, Th cell: T helper cell, TNF: tumor necrosis factor, Treg cell: regulatory T cell, VEGF: vascular endothelial growth factor

(coloured in print)

## **Immunological testing in idiopathic RPL - progress and challenges**

Immune cell testing is without question an expanding field with the exciting potential to offer important diagnostic and prognostic information for couples with RM. For reasons clearly outlined within this review, at present no accepted immune-based tests exist outside of the research setting(3). Enumeration of circulating cytotoxic and helper T, B, and NK cells and assays evaluating the Th cell-associated cytokine production and the cytotoxic activity of NK cells may provide important prognostic markers, however taking these tests forward from 'bench to bedside' requires further validation. The comparative value of these tests also remains to be elucidated(55, 64). Albeit beyond the scope of this review, we suggest that only then should targeted immunomodulatory treatment trials, such as human immunoglobulins (IVIgG), paternal/donor leukocyte immunization, and/ or TNF- $\alpha$  inhibitors, be performed(193, 194).

NK cell testing has gained particular interest. Although commercially available, international consensus regarding "normal" NK cell ranges is required. High-quality prospective case-control studies with sufficient power to account for confounding factors such as age(195) are required in both healthy controls and women with RPL. The key role of NK cells in implantation and vessel remodeling has been highlighted in the present review. However, the suggested changes in their numbers and activity in RPL remain uncertain. Hence, the role of these cells in RPL should be fully elucidated prior to targeting immune activity for therapeutic purposes. Similarly, the value of cytokine testing therefore remains contentious due to data heterogeneity(156, 196) and lack of a universal definition of "increased" Type-1/Type-2 ratios.

An important factor contributing to experimental finding variation is the methodological differences between studies. Apart from the lack of a universal RPL definition, leading to diverse inclusion criteria, the timing of immunological testing remains inconsistent. Immune cell variation during the menstrual cycle and upon pregnancy establishment has been observed, possibly due to the influence of hormone level fluctuation, including progesterone, estrogen, and human chorionic gonadotropin(197, 198). Studies accounting for menstrual cycle phase may also be informative in order to accurately identify pre-conception immune cell markers in women with RPL.

Variation can also be seen between studies examining the total RPL population and those focusing on women with the idiopathic form of this condition. The pathogenesis of non-idiopathic RPL may include co-morbidities like antiphospholipid syndrome and infection, which are underlined by specific patterns of immune dysregulation. Thus, inclusion of women with non-idiopathic RPL may predispose towards the detection of immune abnormalities. To the contrary, it remains uncertain whether idiopathic and non-idiopathic cases are underlined by the same immunological pathogenesis (7). In addition, RPL may be underlined by a multifactorial pathogenic mechanism, with different factors including ethnicity, BMI, and male parameters likely contributing to the reproductive outcome. Moving forward it is important to investigate the total RPL population, with well-powered idiopathic and non-idiopathic subpopulations analysed separately. This would also help determine the optimal timing of immunological testing in clinical practice, i.e. in relation to other diagnostic tests performed.

Sampling methods are also of certain importance and interest. Although uterine endometrial/decidual sampling poses the opportunity to characterise the local immune environment, the introduction of an invasive



endometrial/decidual procedure is not without risk. Peripheral blood tests offer a less invasive, favourable option, however their suitability remains to be fully validated in terms of RPL pathogenesis and prognostic accuracy. Moving forward, paired blood-uterus samples would help bridge this knowledge gap.

Given the potential psychological impact of miscarriage, the impact of stress upon circulating immune cell types, including NK and T cells, is of probable significance(4, 5, 199, 200). The effectiveness of supportive care for women with idiopathic RPL is already shown(201). Studies measuring the value of supportive care upon immune cell function in women with RPL remain to be established.

### Final Remarks

Immune dysfunction is associated with idiopathic RPL. Prospective, well-powered studies to elucidate the exact mechanisms underlying this association and evaluate the diagnostic and prognostic capacity of immunological testing are urgently required. We anticipate this will inform potential future targeted immunotherapies. Albeit beyond the scope of this review, given the potential side-effects and risks of some of these therapies, well conducted pre- and post-conception research studies should precede their routine clinical implementation in the management of idiopathic RPL(202). Current trials, examining the use of antibiotics, steroids, and biologics are expected to offer a new insight on the management of RPL(203-205).

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## References:

- 1.Royal College of Obstetricians & Gynaecologists. The Investigation and Treatment of Couples with Recurrent First-trimester and Second-trimester Miscarriage. Green-top Guideline. 2011;17:1-18.
- 2.Quenby S, Gallos ID, Dhillon-Smith RK, Podesek M, Stephenson MD, Fisher J, et al. Miscarriage matters: the epidemiological, physical, psychological, and economic costs of early pregnancy loss. *Lancet*. 2021;397(10285):1658-67.
- 3.The ESHRE Guideline Group on RPL, Atik RB, Christiansen OB, Elson J, Kolte AM, Lewis S, et al. ESHRE guideline: recurrent pregnancy loss. *Hum Reprod Open*. 2018;2018(2):hoy004.
- 4.Serrano F, Lima ML. Recurrent miscarriage: psychological and relational consequences for couples. *Psychol Psychother*. 2006;79(Pt 4):585-94.
- 5.Kolte AM, Olsen LR, Mikkelsen EM, Christiansen OB, Nielsen HS. Depression and emotional stress is highly prevalent among women with recurrent pregnancy loss. *Hum Reprod*. 2015;30(4):777-82.
- 6.Stray-Pedersen B, Stray-Pedersen S. Recurrent abortion: the role of psychotherapy. In: Sharp F, Beard RW, editors. *Early Pregnancy Loss*. London: Springer; 1988. p. 433-40.
- 7.Ewington LJ, Tewary S, Brosens JJ. New insights into the mechanisms underlying recurrent pregnancy loss. *J Obstet Gynaecol Res*. 2019;45(2):258-65.
- 8.Knuudsen UB, Hansen V, Juul S, Secher NJ. Prognosis of a new pregnancy following previous spontaneous abortions. *Eur J Obstet Gynecol Reprod Biol*. 1991;39(1):31-6.
- 9.Lashen H, Fear K, Sturdee DW. Obesity is associated with increased risk of first trimester and recurrent miscarriage: matched case-control study. *Hum Reprod*. 2004;19(7):1644-6.
- 10.Ness RB, Grisso JA, Hirschinger N, Markovic N, Shaw LM, Day NL, et al. Cocaine and tobacco use and the risk of spontaneous abortion. *N Engl J Med*. 1999;340(5):333-9.
- 11.Khattak S, K-Moghtader G, McMartin K, Barrera M, Kennedy D, Koren G. Pregnancy outcome following gestational exposure to organic solvents: a prospective controlled study. *JAMA*. 1999;281(12):1106-9.
- 12.Rasch V. Cigarette, alcohol, and caffeine consumption: risk factors for spontaneous abortion. *Acta Obstet Gynecol Scand*. 2003;82(2):182-8.
- 13.Andersen LB, Jørgensen JS, Jensen TK, Dalgård C, Barington T, Nielsen J, et al. Vitamin D insufficiency is associated with increased risk of first-trimester miscarriage in the Odense Child Cohort. *Am J Clin Nutr*. 2015;102(3):633-8.
- 14.Mills JL, Simpson JL, Driscoll SG, Jovanovic-Peterson L, Van Allen M, Aarons JH, et al. Incidence of spontaneous abortion among normal women and insulin-dependent diabetic women whose pregnancies were identified within 21 days of conception. *N Engl J Med*. 1988;319(25):1617-23.
- 15.Regan L, Rai R. Epidemiology and the medical causes of miscarriage. *Best Pract Res Clin Obstet Gynaecol*. 2000;14(5):839-54.
- 16.Ford HB, Schust DJ. Recurrent Pregnancy Loss: Etiology, Diagnosis, and Therapy. *Rev Obstet Gynecol*. 2009;2(2):76-83.
- 17.Rai RS, Regan L, Clifford K, Pickering W, Dave M, Mackie I, et al. Antiphospholipid antibodies and beta 2-glycoprotein-I in 500 women with recurrent miscarriage: results of a comprehensive screening approach. *Hum Reprod*. 1995;10(8):2001-5.
- 18.Baud D, Goy G, Jaton K, Osterheld MC, Blumer S, Borel N, et al. Role of Chlamydia trachomatis in Miscarriage. *Emerg Infect Dis*. 2011;17(9):1630-5.
- 19.Cao CJ, Wang YF, Fang DM, Hu Y. Relation between mycoplasma infection and recurrent spontaneous abortion. *Eur Rev Med Pharmacol Sci*. 2018;22(8):2207-11.
- 20.McQueen DB, Bernardi LA, Stephenson MD. Chronic endometritis in women with recurrent early pregnancy loss and/or fetal demise. *Fertil Steril*. 2014;101(4):1026-30.
- 21.Öcak Z, Özlü T, Özyurt O. Association of recurrent pregnancy loss with chromosomal abnormalities and hereditary thrombophilias. *Afr Health Sci*. 2013;13(2):447-52.
- 22.Hyde KJ, Schust DJ. Genetic considerations in recurrent pregnancy loss. *Cold Spring Harb Perspect Med*. 2015;5(3):a023119.
- 23.Ibrahim Y, Johnstone E. The male contribution to recurrent pregnancy loss. *Transl Androl Urol*. 2018;7(Suppl 3):S317-S27.
- 24.Mor G, Cardenas I, Abrahams V, Guller S. Inflammation and pregnancy: the role of the immune system at the implantation site. *Ann N Y Acad Sci*. 2011;1221:80-7.
- 25.Mor G, Aldo P, Alvero AB. The unique immunological and microbial aspects of pregnancy. *Nat Rev Immunol*. 2017;17(8):469-82.
- 26.Quenby S, Vince G, Farquharson R, Aplin J. Recurrent Miscarriage: A Defect in Nature's Quality Control? *Hum Reprod*. 2002;17(8):1959-63.
- 27.Ticconi C, Pietropolli A, Di Simone N, Piccione E, Fazleabas A. Endometrial Immune Dysfunction in Recurrent Pregnancy Loss. *Int J Mol Sci*. 2019;20(21):5332.
- 28.Moffett-King A. Natural killer cells and pregnancy. *Nat Rev Immunol*. 2002;2(9):656-63.
- 29.Ander SE, Diamond MS, Coyne CB. Immune responses at the maternal-fetal interface. *Sci Immunol*. 2019;4(31):eaat6114.
- 30.Liu S, Diao L, Huang C, Li Y, Zeng Y, Kwak-Kim JYH. The role of decidual immune cells on human pregnancy. *J Reprod Immunol*. 2017;124:44-53.
- 31.Braud VM, Allan DS, O'Callaghan CA, Söderström K, D'Andrea A, Ogg GS, et al. HLA-E binds to natural killer cell receptors CD94/NKG2A, B and C. *Nature*. 1998;391(6669):795-9.
- 32.Long EO, Barber DF, Burshtyn DN, Faure M, Peterson M, Rajagopalan S, et al. Inhibition of natural killer cell activation signals by killer cell immunoglobulin-like receptors (CD158). *Immunol Rev*. 2001;181:223-33.
- 33.Apps R, Murphy SP, Fernando R, Gardner L, Ahad T, Moffett A. Human leucocyte antigen (HLA) expression of primary trophoblast cells and placental cell lines, determined using single antigen beads to characterize allotype specificities of anti-HLA antibodies. *Immunology*. 2009;127(1):26-39.
- 34.Apps R, Gardner L, Moffett A. A critical look at HLA-G. *Trends Immunol*. 2008;29(7):313-21.
- 35.King A, Boocock C, Sharkey AM, Gardner L, Beretta A, Siccardi AG, et al. Evidence for the expression of HLA-A-C class I mRNA and protein by human first trimester trophoblast. *J Immunol*. 1996;156(6):2068-76.
- 36.King A, Allan DS, Bowen M, Powis SJ, Joseph S, Verma S, et al. HLA-E is expressed on trophoblast and interacts with CD94/NKG2 receptors on decidual NK cells. *Eur J Immunol*. 2000;30(6):1623-31.
- 37.Hutter H, Hammer A, Dohr G, Hunt JS. HLA expression at the maternal-fetal interface. *Dev Immunol*. 1998;6(3-4):197-204.
- 38.Sargent IL, Borzychowski AM, Redman CW. NK cells and human pregnancy – an inflammatory view. *Trends Immunol*. 2006;27(9):399-404.
- 39.Hunt JS, Fishback JL, Chumbley G, Loke YW. Identification of class I MHC mRNA in human first trimester trophoblast cells by in situ hybridization. *J Immunol*. 1990;144(11):4420-5.
- 40.Jones CJP, Aplin JD. Glycosylation at the Fetomaternal Interface: Does the Glycocode Play a Critical Role in Implantation? *Glycoconj J*. 2009;26(3):359-66.
- 41.Marchal-Bras-Goncalves R, Rouas-Freiss N, Connan F, Choppin J, Dausset J, Carosella ED, et al. A soluble HLA-G protein that inhibits natural killer cell-mediated cytotoxicity. *Transplant Proc*. 2001;33(3):2355-9.
- 42.Solier C, Aguerre-Girr M, Lenfant F, Campan A, Berrebi A, Rebmann V, et al. Secretion of pro-apoptotic intron 4-retaining soluble HLA-G1 by human villous trophoblast. *Eur J Immunol*. 2002;32(12):3576-86.
- 43.Manaster I, Mandelboim O. The unique properties of uterine NK cells. *Am J Reprod Immunol*. 2010;63(6):434-44.
- 44.Powell RM, Lissauer D, Tamblin J, Beggs A, Cox P, Moss P, et al. Decidual T Cells Exhibit a Highly Differentiated Phenotype and Demonstrate Potential Fetal Specificity and a Strong Transcriptional Response to IFN. *J Immunol*. 2017;199(10):3406-17.
- 45.Saini V, Arora S, Yadav A, Bhattacharjee J. Cytokines in recurrent pregnancy loss. *Clin Chim Acta*. 2011;412(9-10):702-8.
- 46.Laird SM, Tuckerman EM, Cork BA, Linjawi S, Blakemore AI, Li TC. A review of immune cells and molecules in women with recurrent miscarriage. *Hum Reprod Update*. 2003;9(2):163-74.
- 47.Fukui A, Funamizu A, Yokota M, Yamada K, Nakamura R, Fukuhara R, et al. Uterine and circulating natural killer cells and their roles in women with recurrent pregnancy loss, implantation failure and preeclampsia. *J Reprod Immunol*. 2011;90(1):105-10.
- 48.Rodrigues MN, Favaron PO, Dombrowski JG, de Souza RM, Migliano MA. Role of natural killer (NK) cells during pregnancy: A review. *Open J Anim Sci*. 2013;3(2):138-44.
- 49.Tang AW, Alfirevic Z, Quenby S. Natural killer cells and pregnancy outcomes in women with recurrent miscarriage and infertility: a systematic review. *Hum Reprod*. 2011;26(8):1971-80.
- 50.Higuchi K, Aoki K, Kimbara T, Hosoi N, Yamamoto T, Okada H. Suppression of natural killer cell activity by monocytes following immunotherapy for recurrent spontaneous aborters. *Am J Reprod Immunol*. 1995;33(3):221-7.
- 51.Hidaka Y, Amino N, Iwatani Y, Kaneda T, Mitsuda N, Morimoto Y, et al. Changes in natural killer cell activity in normal pregnant and postpartum women: increases in the first trimester and postpartum period and decrease in late pregnancy. *J Reprod Immunol*. 1991;20(1):73-83.
- 52.Vassiliadou N, Bulmer JN. Functional studies of human decidual in spontaneous early pregnancy loss: effect of soluble factors and purified CD56+ lymphocytes on killing of natural killer- and lymphokine-activated killer-sensitive targets. *Biol Reprod*. 1998;58(4):982-7.
- 53.Ashkar AA, Croy BA. Functions of uterine natural killer cells are mediated by interferon gamma production during murine pregnancy. *Semin Immunol*. 2001;13(4):235-41.
- 54.Carp H. Cytokines in recurrent miscarriage. *Lupus*. 2004;13(9):630-4.
- 55.ESHRE Early Pregnancy Guideline development Group. Recurrent Pregnancy Loss. European Society of Human Reproduction and Embryology; 2017. p. 1-153.
- 56.Tuckerman E, Laird SM, Prakash A, Li TC. Prognostic value of the measurement of uterine natural killer cells in the endometrium of women with recurrent miscarriage. *Hum Reprod*. 2007;22(8):2208-13.
- 57.Tamblin JA, Jeffery LE, Susarla R, Lissauer DM, Coort SL, Garcia AM, et al. Transcriptomic analysis of vitamin D responses in uterine and peripheral NK cells. *Reproduction*. 2019;158(2):211-21.
- 58.Cooper MA, Fehniger TA, Turner SC, Chen KS, Ghaheri BA, Ghayur T, et al. Human natural killer cells: a unique innate immunoregulatory role for the CD56(bright) subset. *Blood*. 2001;97(10):3146-51.
- 59.Jacobs R, Hintzen G, Kemper A, Beul K, Kempf S, Behrens G, et al. CD56bright cells differ in their KIR repertoire and cytotoxic features from CD56dim NK cells. *Eur J Immunol*. 2001;31(10):3121-7.
- 60.Lanier LL, Le AM, Civin CI, Loken MR, Phillips JH. The relationship of CD16 (Leu-11) and Leu-19 (NKH-1) antigen expression on human peripheral blood NK cells and cytotoxic T lymphocytes. *J Immunol*. 1986;136(12):4480-6.
- 61.Moffett A, Regan L, Braude P. Natural killer cells, miscarriage, and infertility. *BMJ*. 2004;329(7477):1283-5.
- 62.King A, Birkby C, Loke YW. Early human decidual cells exhibit NK activity against the K562 cell line but not against first trimester trophoblast. *Cell Immunol*. 1989;118(2):337-44.
- 63.Abadia-Molina AC, Ruiz C, Montes MJ, King A, Loke YW, Olivares EG. Immune phenotype and cytotoxic activity of lymphocytes from human term decidua against trophoblast. *J Reprod Immunol*. 1996;31(1-2):109-23.
- 64.Laird SM, Lash GE, Li TC, Bulmer JN. The Role of Natural Killer Cells in Human Fertility. *Royal College of Obstetricians and Gynaecologists*. 2016;Scientific Impact Paper No. 53:1-11.
- 65.Hanna J, Goldman-Wohl D, Hamani Y, Avraham I, Greenfield C, Natanson-Yaron S, et al. Decidual NK cells regulate key developmental processes at the human fetal-maternal interface. *Nat Med*. 2006;12(9):1065-74.
- 66.Lash GE, Schiessl B, Kirkley M, Innes BA, Cooper A, Searle RF, et al. Expression of angiogenic growth factors by uterine natural killer cells during early pregnancy. *J Leukoc Biol*. 2006;80(5):572-80.
- 67.Saito S, Nishikawa K, Morii T, Enomoto M, Narita N, Motoyoshi K, et al. Cytokine production by CD16-CD56bright natural killer cells in the human early pregnancy decidua. *Int Immunol*. 1993;5(5):559-63.
- 68.Perricone R, Di Muzio G, Perricone C, Giacomelli R, De Nardo D, Fontana L, et al. High levels of peripheral blood NK cells in women suffering from recurrent spontaneous abortion are reverted from high-dose intravenous immunoglobulins. *Am J Reprod Immunol*. 2006;55(3):232-9.
- 69.Seshadri S, Sunkara SK. Natural killer cells in female infertility and recurrent miscarriage: a systematic review and meta-analysis. *Hum Reprod Update*. 2014;20(3):429-38.
- 70.Shakhar K, Ben-Eliyahu S, Loewenthal R, Rosenne E, Carp H. Differences in number and activity of peripheral natural killer cells in primary versus secondary recurrent miscarriage. *Fertil Steril*. 2003;80(2):368-75.
- 71.Beer AE, Kwak JY, Ruiz JE. Immunophenotypic profiles of peripheral blood lymphocytes in women with recurrent pregnancy losses and in infertile women with multiple failed in vitro fertilization cycles. *Am J Reprod Immunol*. 1996;35(4):376-82.
- 72.King K, Smith S, Chapman M, Sacks G. Detailed analysis of peripheral blood natural killer (NK) cells in women with recurrent miscarriage. *Hum Reprod*. 2010;25(1):52-8.
- 73.Yoo JH, Kwak-Kim J, Han AR, Ahn H, Cha SH, Koong MK, et al. Peripheral blood NK cell cytotoxicities are negatively correlated with CD8(+) T cells in fertile women but not in women with a history of recurrent pregnancy loss. *Am J Reprod Immunol*. 2012;68(1):38-46.
- 74.Fukui A, Kwak-Kim J, Ntrivalas E, Gilman-Sachs A, Lee SK, Beaman K. Intracellular cytokine expression of peripheral blood natural killer cell subsets in women with recurrent spontaneous abortions and implantation failures. *Fertil Steril*. 2008;89(1):157-65.

75. Coulam CB, Goodman C, Roussev RG, Thomason EJ, Beaman KD. Systemic CD56+ cells can predict pregnancy outcome. *Am J Reprod Immunol*. 1995;33(1):40-6.
76. Adib Rad H, Basirat Z, Mostafazadeh A, Faramarzi M, Bijani A, Nouri HR, et al. Evaluation of peripheral blood NK cell subsets and cytokines in unexplained recurrent miscarriage. *J Chin Med Assoc*. 2018;81(12):1065-70.
77. Emmer PM, Veerhoek M, Nelen WL, Steegers EA, Joosten I. Natural killer cell reactivity and HLA-G in recurrent spontaneous abortion. *Transplant Proc*. 1999;31(4):1838-40.
78. Quenby S, Bates M, Doig T, Brewster J, Lewis-Jones DI, Johnson PM, et al. Pre-implantation endometrial leukocytes in women with recurrent miscarriage. *Hum Reprod*. 1999;14(9):2386-91.
79. Kwak JY, Beer AE, Kim SH, Mantouvalos NP. Immunopathology of the implantation site utilizing monoclonal antibodies to natural killer cells in women with recurrent pregnancy losses. *Am J Reprod Immunol*. 1999;41(1):91-8.
80. Lachapelle MH, Miron P, Hemmings R, Roy DC. Endometrial T, B, and NK cells in patients with recurrent spontaneous abortion. Altered profile and pregnancy outcome. *J Immunol*. 1996;156(10):4027-34.
81. Mosimann B, Wagner M, Shehata H, Poon LC, Ford B, Nicolaidis KH, et al. Natural killer cells and their activation status in normal pregnancy. *Int J Reprod Med*. 2013;2013:906813.
82. Coulam CB, Roussev RG. Correlation of NK cell activation and inhibition markers with NK cytotoxicity among women experiencing immunologic implantation failure after in vitro fertilization and embryo transfer. *J Assist Reprod Genet*. 2003;20(2):58-62.
83. Ntrivalas EI, Kwak-Kim JY, Gilman-Sachs A, Chung-Bang H, Ng SC, Beaman KD, et al. Status of peripheral blood natural killer cells in women with recurrent spontaneous abortions and infertility of unknown aetiology. *Hum Reprod*. 2001;16(5):855-61.
84. Emmer PM, Nelen WL, Steegers E, Hendriks JC, Veerhoek M, Joosten I. Peripheral natural killer cytotoxicity and CD56(pos)CD16(pos) cells increase during early pregnancy in women with a history of recurrent spontaneous abortion. *Hum Reprod*. 2000;15(5):1163-9.
85. Souza SS, Ferriani RA, Santos CM, Voltarelli JC. Immunological evaluation of patients with recurrent abortion. *J Reprod Immunol*. 2002;56(1-2):111-21.
86. Aoki K, Kajjura S, Matsumoto Y, Ogasawara M, Okada S, Yagami Y, et al. Preconceptional natural-killer-cell activity as a predictor of miscarriage. *Lancet*. 1995;345(8961):1340-2.
87. Kodama T, Hara T, Okamoto E, Kusunoki Y, Ohama K. Characteristic changes of large granular lymphocytes that strongly express CD56 in endometrium during the menstrual cycle and early pregnancy. *Hum Reprod*. 1998;13(4):1036-43.
88. Higuma-Myojo S, Sasaki Y, Miyazaki S, Sakai M, Siozaki A, Miwa N, et al. Cytokine profile of natural killer cells in early human pregnancy. *Am J Reprod Immunol*. 2005;54(1):21-9.
89. Veenstra van Nieuwenhoven AL, Bouman A, Moes H, Heineman MJ, de Leij LF, Santema J, et al. Cytokine production in natural killer cells and lymphocytes in pregnant women compared with women in the follicular phase of the ovarian cycle. *Fertil Steril*. 2002;77(5):1032-7.
90. Fukui A, Funamizu A, Fukuhara R, Shibahara H. Expression of natural cytotoxicity receptors and cytokine production on endometrial natural killer cells in women with recurrent pregnancy loss or implantation failure, and the expression of natural cytotoxicity receptors on peripheral blood natural killer cells in pregnant women with a history of recurrent pregnancy loss. *J Obstet Gynaecol Res*. 2017;43(11):1678-86.
91. Cerwenka A, Lanier LL. Natural Killer Cell Memory in Infection, Inflammation and Cancer. *Nat Rev Immunol*. 2016;16(2):112-23.
92. Sun JC, Beilke JN, Lanier LL. Adaptive Immune Features of Natural Killer Cells. *Nature*. 2009;457(7229):557-61.
93. Tesi B, Schlums H, Cichocki F, Bryceson YT. Epigenetic Regulation of Adaptive NK Cell Diversification. *Trends Immunol*. 2016;37(7):451-61.
94. Schlums H, Cichocki F, Tesi B, Theorell J, Beziat V, Holmes TD, et al. Cytomegalovirus Infection Drives Adaptive Epigenetic Diversification of NK Cells With Altered Signaling and Effector Function. *Immunity*. 2015;42(3):443-56.
95. Béziat V, Dalgaard O, Asselah T, Halfon P, Bedossa P, Boudifa A, et al. CMV Drives Clonal Expansion of NKG2C+ NK Cells Expressing Self-Specific KIRs in Chronic Hepatitis Patients. *Eur J Immunol*. 2012;42(2):447-57.
96. Romee R, Rosario M, Berrien-Elliott MM, Wagner JA, Jewell BA, Schappe T, et al. Cytokine-induced Memory-Like Natural Killer Cells Exhibit Enhanced Responses Against Myeloid Leukemia. *Sci Transl Med*. 2016;8(357):357ra123.
97. Vento-Tormo R, Efremova M, Botting RA, Turco MY, Vento-Tormo M, Meyer KB, et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature*. 2018;563(7731):347-53.
98. Feyaerts D, van der Meer A, Joosten I, van der Molen RG. Selective Expansion and CMV-dependency in Pregnancy Trained Human Endometrial NK Cells. *Cell Mol Immunol*. 2019;16(4):410-1.
99. Croy BA, He H, Esadeg S, Wei Q, McCartney D, Zhang J, et al. Uterine Natural Killer Cells: Insights Into Their Cellular and Molecular Biology From Mouse Modelling. *Reproduction*. 2003;126(2):149-60.
100. Ashkar AA, Di Santo JP, Croy BA. Interferon gamma contributes to initiation of uterine vascular modification, decidual integrity, and uterine natural killer cell maturation during normal murine pregnancy. *J Exp Med*. 2000;192(2):259-70.
101. Hirayama D, Iida T, Nakase H. The Phagocytic Function of Macrophage-Enforcing Innate Immunity and Tissue Homeostasis. *Int J Mol Sci*. 2018;19(1):92.
102. Jensen AL, Collins J, Shipman EP, Wira CR, Guyre PM, Pioli PA. A subset of human uterine endometrial macrophages is alternatively activated. *Am J Reprod Immunol*. 2012;68(5):374-86.
103. Tang MX, Hu XH, Liu ZZ, Kwak-Kim J, Liao AH. What are the roles of macrophages and monocytes in human pregnancy? *J Reprod Immunol*. 2015;112:73-80.
104. Yao Y, Xu XH, Jin L. Macrophage Polarization in Physiological and Pathological Pregnancy. *Front Immunol*. 2019;10:792.
105. Rieger L, Honig A, Sütterlin M, Kapp M, Dietl J, Ruck P, et al. Antigen-presenting cells in human endometrium during the menstrual cycle compared to early pregnancy. *J Soc Gynecol Investig*. 2004;11(7):488-93.
106. Gardner L, Moffett A. Dendritic cells in the human decidua. *Biol Reprod*. 2003;69(4):1438-46.
107. Ehrentraut S, Sauss K, Neumeister R, Luley L, Oettel A, Fette F, et al. Human Miscarriage Is Associated With Dysregulations in Peripheral Blood-Derived Myeloid Dendritic Cell Subsets. *Front Immunol*. 2019;10:2440.
108. Della Bella S, Giannelli S, Cozzi V, Signorelli V, Cappelletti M, Cetin C, et al. Incomplete activation of peripheral blood dendritic cells during healthy human pregnancy. *Clin Exp Immunol*. 2011;164(2):180-92.
109. Steinman RM. The dendritic cell system and its role in immunogenicity. *Annu Rev Immunol*. 1991;9:271-96.
110. Kämmerer U, Eggert AO, Kapp M, McLellan AD, Geijtenbeek TBH, Dietl J, et al. Unique appearance of proliferating antigen-presenting cells expressing DC-SIGN (CD209) in the decidua of early human pregnancy. *Am J Pathol*. 2003;162(3):887-96.
111. Askelund K, Liddell HS, Zanderigo AM, Fernando NS, Khong TY, Stone PR, et al. CD83+ dendritic cells in the decidua of women with recurrent miscarriage and normal pregnancy. *Placenta*. 2004;25(2-3):140-5.
112. Lucas ES, Vrljicak P, Muter J, Diniz-da-Costa MM, Brighton PJ, Kong CS, et al. Recurrent pregnancy loss is associated with a pro-senescent decidual response during the peri-implantation window. *Commun Biol*. 2020;3(1):37.
113. Guenther S, Vrekoussis T, Heublein S, Bayer B, Anz D, Knabl J, et al. Decidual macrophages are significantly increased in spontaneous miscarriages and over-express FasL: a potential role for macrophages in trophoblast apoptosis. *Int J Mol Sci*. 2012;13(7):9069-80.
114. Quack KC, Vassiliadou N, Pudney J, Anderson DJ, Hill JA. Leukocyte activation in the decidua of chromosomally normal and abnormal fetuses from women with recurrent abortion. *Hum Reprod*. 2001;16(5):949-55.
115. Smith SD, Dunk CE, Aplin JD, Harris LK, Jones RL. Evidence for Immune Cell Involvement in Decidual Spiral Arteriole Remodeling in Early Human Pregnancy. *Am J Pathol*. 2009;174(5):1959-71.
116. Tsao FY, Wu MY, Chang YL, Wu CT, Ho HN. M1 macrophages decrease in the deciduae from normal pregnancies but not from spontaneous abortions or unexplained recurrent spontaneous abortions. *J Formos Med Assoc*. 2018;117(3):204-11.
117. Brown MB, von Chamier M, Allam AB, Reyes L. M1/M2 Macrophage Polarity in Normal and Complicated Pregnancy. *Front Immunol*. 2014;5:606.
118. Hutter S, Heublein S, Knabl J, Andergassen U, Vrekoussis T, Makrigiannakis A, et al. Macrophages: are they involved in endometriosis, abortion and preeclampsia and how? *J Nippon Med Sch*. 2013;80(2):97-103.
119. Wissinger E. CD8+ T Cells. *Bitesized immunology: British Society for Immunology*.
120. Tsuda S, Nakashima A, Shima T, Saito S. New Paradigm in the Role of Regulatory T Cells During Pregnancy. *Front Immunol*. 2019;10:573.
121. Bulmer JN, Morrison L, Longfellow M, Ritson A, Pace D. Granulated lymphocytes in human endometrium: histochemical and immunohistochemical studies. *Hum Reprod*. 1991;6(6):791-8.
122. Ng SC, Gilman-Sachs A, Thaker P, Beaman KD, Beer AE, Kwak-Kim J. Expression of intracellular Th1 and Th2 cytokines in women with recurrent spontaneous abortion, implantation failures after IVF/ET or normal pregnancy. *Am J Reprod Immunol*. 2002;48(2):77-86.
123. Carbone J, Sarmiento E, Gallego A, Lanio N, Navarro J, García S, et al. Peripheral blood T- and B-cell immunophenotypic abnormalities in selected women with unexplained recurrent miscarriage. *J Reprod Immunol*. 2016;113:50-3.
124. Prado-Drayer A, Teppa J, Sánchez P, Camejo MI. Immunophenotype of peripheral T lymphocytes, NK cells and expression of CD69 activation marker in patients with recurrent spontaneous abortions, during the mid-luteal phase. *Am J Reprod Immunol*. 2008;60(1):66-74.
125. Yahata T, Kurabayashi T, Honda A, Takakuwa K, Tanaka K, Abo T. Decrease in the proportion of granulated CD56+ T-cells in patients with a history of recurrent abortion. *J Reprod Immunol*. 1998;38(1):63-73.
126. Tilburgs T, Claas FHJ, Scherjon SA. Elsevier Trophoblast Research Award Lecture: Unique Properties of Decidual T Cells and Their Role in Immune Regulation During Human Pregnancy. *Placenta*. 2010;31 Suppl:S82-6.
127. Kwak JY, Beaman KD, Gilman-Sachs A, Ruiz JE, Schewitz D, Beer AE. Up-regulated expression of CD56+, CD56+/CD16+, and CD19+ cells in peripheral blood lymphocytes in pregnant women with recurrent pregnancy losses. *Am J Reprod Immunol*. 1995;34(2):93-9.
128. Saito S, Nishikawa K, Morii T, Narita N, Enomoto M, Ichijo M. Expression of activation antigens CD69, HLA-DR, interleukin-2 receptor-alpha (IL-2R alpha) and IL-2R beta on T cells of human decidua at an early stage of pregnancy. *Immunology*. 1992;75(4):710-2.
129. Lissauer D, Piper K, Goodyear O, Kilby MD, Moss PA. Fetal-specific CD8+ cytotoxic T cell responses develop during normal human pregnancy and exhibit broad functional capacity. *J Immunol*. 2012;189(2):1072-80.
130. Ghafourian M, Abuhamid A, Karami N. Increase of peripheral blood TCD8+ cells in women with recurrent miscarriage. *J Obstet Gynaecol*. 2014;34(1):36-9.
131. Wang SC, Li YH, Piao HL, Hong XW, Zhang D, Xu YY, et al. PD-1 and Tim-3 pathways are associated with regulatory CD8+ T-cell function in decidua and maintenance of normal pregnancy. *Cell Death Dis*. 2015;6:e1738.
132. Kuon RJ, Schaumann J, Goeggl T, Strowitzki T, Sadeghi M, Opelz G, et al. Patients with idiopathic recurrent miscarriage show higher levels of DR+ activated T-cells that are less responsive to mitogens. *J Reprod Immunol*. 2015;112:82-7.
133. Yang KM, Ntrivalas E, Cho HJ, Kim NY, Beaman K, Gilman-Sachs A, et al. Women With Multiple Implantation Failures and Recurrent Pregnancy Losses Have Increased Peripheral Blood T Cell Activation. *Am J Reprod Immunol*. 2010;63(5):370-8.
134. Clark DA, Chaouat G, Mogil R, Wegmann TG. Prevention of spontaneous abortion in DBA/2-mated CBA/J mice by GM-CSF involves CD8+ T cell-dependent suppression of natural effector cell cytotoxicity against trophoblast target cells. *Cell Immunol*. 1994;154(1):143-52.
135. Calleja-Agius J, Brincat MP. Recurrent miscarriages: What is the role of cytokines? *Gynecol Endocrinol*. 2008;24(12):663-8.
136. Yui J, Garcia-Lloret M, Wegmann TG, Guilbert LJ. Cytotoxicity of tumour necrosis factor-alpha and gamma-interferon against primary human placental trophoblasts. *Placenta*. 1994;15(8):819-35.
137. Torchinsky A, Shepshelovich J, Orenstein H, Zaslavsky Z, Savion S, Carp H, et al. TNF-alpha protects embryos exposed to developmental toxicants. *Am J Reprod Immunol*. 2003;49(3):159-68.
138. Guilbert LJ. There is a bias against type 1 (inflammatory) cytokine expression and function in pregnancy. *J Reprod Immunol*. 1996;32(2):105-10.
139. Mor G, Cardenas I. The immune system in pregnancy: a unique complexity. *Am J Reprod Immunol*. 2010;63(6):425-33.
140. Lissauer D, Eldershaw SA, Inman CF, Coomarasamy A, Moss PAH, Kilby MD. Progesterone promotes maternal-fetal tolerance by reducing human maternal T-cell polyfunctionality and inducing a specific cytokine profile. *Eur J Immunol*. 2015;45(10):2858-72.
141. Somerset DA, Zheng Y, Kilby MD, Sansom DM, Drayton MT. Normal human pregnancy is associated with an elevation in the immune suppressive CD25+ CD4+ regulatory T-cell subset. *Immunology*. 2004;112(1):38-43.
142. Jin LP, Chen QY, Zhang T, Guo PF, Li DJ. The CD4+CD25+ bright regulatory T cells and CTLA-4 expression in peripheral and decidual lymphocytes are down-regulated in human miscarriage. *Clin Immunol*. 2009;133(3):402-10.

143. Mei S, Tan J, Chen H, Chen Y, Zhang J. Changes of CD4+CD25high regulatory T cells and FOXP3 expression in unexplained recurrent spontaneous abortion patients. *Fertil Steril*. 2010;94(6):2244-7.
144. Yang H, Qiu L, Chen G, Ye Z, Lü C, Lin Q. Proportional change of CD4+CD25+ regulatory T cells in decidua and peripheral blood in unexplained recurrent spontaneous abortion patients. *Fertil Steril*. 2008;89(3):656-61.
145. Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 Cells. *Annu Rev Immunol*. 2009;27:485-517.
146. Moore KW, de Waal Malefyt R, Coffman RL, O'Garra A. Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol*. 2001;19:683-765.
147. Raghupathy R, Makhseed M, Azizieh F, Hassan N, Al-Azemi M, Al-Shamali E. Maternal Th1- and Th2-type reactivity to placental antigens in normal human pregnancy and unexplained recurrent spontaneous abortions. *Cell Immunol*. 1999;196(2):122-30.
148. Marzi M, Vigano A, Trabattoni D, Villa ML, Salvaggio A, Clerici E, et al. Characterization of type 1 and type 2 cytokine production profile in physiologic and pathologic human pregnancy. *Clin Exp Immunol*. 1996;106(1):127-33.
149. Piccinni MP, Beloni L, Livi C, Maggi E, Scarselli G, Romagnani S. Defective production of both leukemia inhibitory factor and type 2 T-helper cytokines by decidual T cells in unexplained recurrent abortions. *Nat Med*. 1998;4(9):1020-4.
150. Makhseed M, Raghupathy R, Azizieh F, Al-Azemi MM, Hassan NA, Bandar A. Mitogen-induced cytokine responses of maternal peripheral blood lymphocytes indicate a differential Th-type bias in normal pregnancy and pregnancy failure. *Am J Reprod Immunol*. 1999;42(5):273-81.
151. Raghupathy R, Makhseed M, Azizieh F, Omu A, Gupta M, Farhat R. Cytokine production by maternal lymphocytes during normal human pregnancy and in unexplained recurrent spontaneous abortion. *Hum Reprod*. 2000;15(3):713-8.
152. Hossein H, Mahroo M, Abbas A, Firouzeh A, Nadia H. Cytokine production by peripheral blood mononuclear cells in recurrent miscarriage. *Cytokine*. 2004;28(2):83-6.
153. Lissauer D, Goodyear O, Khanum R, Moss PA, Kilby MD. Profile of maternal CD4 T-cell effector function during normal pregnancy and in women with a history of recurrent miscarriage. *Clin Sci*. 2014;126(5):347-54.
154. Daher S, Shulzhenko N, Morgun A, Mattar R, Rampim GF, Camano L, et al. Associations between cytokine gene polymorphisms and recurrent pregnancy loss. *J Reprod Immunol*. 2003;58(1):69-77.
155. Kwak-Kim JY, Chung-Bang HS, Ng SC, Ntrivalas EI, Mangubat CP, Beaman KD, et al. Increased T helper 1 cytokine responses by circulating T cells are present in women with recurrent pregnancy losses and in infertile women with multiple implantation failures after IVF. *Hum Reprod*. 2003;18(4):767-73.
156. Jenkins C, Roberts J, Wilson R, MacLean MA, Shilito J, Walker JJ. Evidence of a T(H) 1 type response associated with recurrent miscarriage. *Fertil Steril*. 2000;73(6):1206-8.
157. Bansal AS. Joining the immunological dots in recurrent miscarriage. *Am J Reprod Immunol*. 2010;64(5):307-15.
158. Albers B, Johnson A, Lewis J, Raff M, Roberts K, Walter P. Helper T Cells and Lymphocyte Activation. *New York: Garland Science*; 2002. 1410-23 p.
159. Ramhorst R, Fraccaroli L, Aldo P, Alvero AB, Cardenas I, Leirós CP, et al. Modulation and recruitment of inducible regulatory T cells by first trimester trophoblast cells. *Am J Reprod Immunol*. 2012;67(1):17-27.
160. Winger EE, Reed JL. Low circulating CD4(+) CD25(+) Foxp3(+) T regulatory cell levels predict miscarriage risk in newly pregnant women with a history of failure. *Am J Reprod Immunol*. 2011;66(4):320-8.
161. Wang WJ, Hao CF, Qu QL, Wang X, Qiu LH, Lin QD. The deregulation of regulatory T cells on interleukin-17-producing T helper cells in patients with unexplained early recurrent miscarriage. *Hum Reprod*. 2010;25(10):2591-6.
162. Mueller SN, Gebhardt T, Carbone FR, Heath WR. Memory T Cell Subsets, Migration Patterns, and Tissue Residence. *Annu Rev Immunol*. 2013;31:137-61.
163. Kieffer TEC, Laskewitz A, Scherjon SA, Faas MM, Prins JR. Memory T Cells in Pregnancy. *Front Immunol*. 2019;10:625.
164. Lanzavecchia A, Sallusto F. Understanding the Generation and Function of Memory T Cell Subsets. *Curr Opin Immunol*. 2005;17(3):326-32.
165. Tilburgs T, Schonkeren D, Eikmans M, Nagtzaam NM, Datema G, Swings GM, et al. Human decidua tissue contains differentiated CD8+ effector-memory T cells with unique properties. *J Immunol*. 2010;185(7):4470-7.
166. Mohan Shah N, Herasimtschuk AA, Boasso A, Benlahrech A, Fuchs D, Imami N, et al. Changes in T Cell and Dendritic Cell Phenotype From Mid to Late Pregnancy Are Indicative of a Shift From Immune Tolerance to Immune Activation. *Front Immunol*. 2017;8:1138.
167. Kieffer TEC, Faas MM, Scherjon SA, Prins JR. Pregnancy Persistently Affects Memory T Cell Populations. *J Reprod Immunol*. 2017;119:1-8.
168. Loewendorf AI, Nguyen TA, Yesayan MN, Kahn DA. Normal Human Pregnancy Results in Maternal Immune Activation in the Periphery and at the Uteroplacental Interface. *PLoS One*. 2014;9(5):e96723.
169. Ramhorst R, Garcia V, Agriello E, Corigliano A, Etchepareborda E, Irigoyen M, et al. Intracellular Expression of CD69 in Endometrial and Peripheral T Cells Represents a Useful Marker in Women With Recurrent Miscarriage: Modulation After Allogeneic Leukocyte Immunotherapy. *Am J Reprod Immunol*. 2003;49(3):149-58.
170. Southcombe JH, Mounce G, McGee K, Elghajji A, Broens JJ, Quenby S, et al. An Altered Endometrial CD8 Tissue Resident Memory T Cell Population in Recurrent Miscarriage. *Sci Rep*. 2017;7:41335.
171. Ruiz-Irastorza G, Lima F, Alves J, Khamashta MA, Simpson J, Hughes GR, et al. Increased rate of lupus flare during pregnancy and the puerperium: a prospective study of 78 pregnancies. *Br J Rheumatol*. 1996;35(2):133-8.
172. Nelson JL, Ostensen M. Pregnancy and rheumatoid arthritis. *Rheum Dis Clin North Am*. 1997;23(1):195-212.
173. Kamat BR, Isaacson PG. The immunocytochemical distribution of leukocytic subpopulations in human endometrium. *Am J Pathol*. 1987;127(1):66-73.
174. Vargas ML, Santos JL, Ruiz C, Montes MJ, Alemán P, Garcia-Tortosa C, et al. Comparison of the proportions of leukocytes in early and term human decidua. *Am J Reprod Immunol*. 1993;29(3):135-40.
175. Lima J, Martins C, Leandro MJ, Nunes G, Sousa MJ, Branco JC, et al. Characterization of B cells in healthy pregnant women from late pregnancy to post-partum: a prospective observational study. *BMC Pregnancy Childbirth*. 2016;16(1):139.
176. Fettke F, Schumacher A, Costa SD, Zenclussen AC. B cells: the old new players in reproductive immunology. *Front Immunol*. 2014;5:285.
177. Muzzio D, Zenclussen AC, Jensen F. The role of B cells in pregnancy: the good and the bad. *Am J Reprod Immunol*. 2013;69(4):408-12.
178. Blair PA, Noreña LY, Flores-Borja F, Rawlings DJ, Isenberg DA, Ehrenstein MR, et al. CD19(+)/CD24(hi)/CD38(hi) B cells exhibit regulatory capacity in healthy individuals but are functionally impaired in systemic Lupus Erythematosus patients. *Immunity*. 2010;32(1):129-40.
179. Griffin DO, Holdick NE, Rothstein TL. Human B1 cells in umbilical cord and adult peripheral blood express the novel phenotype CD20+ CD27+ CD43+ CD70-. *J Exp Med*. 2011;208(1):67-80.
180. Casali P, Schettino EW. Structure and function of natural antibodies. *Curr Top Microbiol Immunol*. 1996;210:167-79.
181. Duan B, Morel L. Role of B-1a cells in autoimmunity. *Autoimmun Rev*. 2006;5(6):403-8.
182. Mantovani L, Wilder RL, Casali P. Human rheumatoid B-1a (CD5+ B) cells make somatically hypermutated high affinity IgM rheumatoid factors. *J Immunol*. 1993;151(1):473-88.
183. Kasaian MT, Ikematsu H, Casali P. Identification and analysis of a novel human surface CD5- B lymphocyte subset producing natural antibodies. *J Immunol*. 1992;148(9):2690-702.
184. Parra D, Takizawa F, Sunyer JO. Evolution of B cell immunity. *Annu Rev Anim Biosci*. 2013;1:65-97.
185. Mizoguchi A, Bhan AK. A case for regulatory B cells. *J Immunol*. 2006;176(2):705-10.
186. Flores-Borja F, Bosma A, Ng D, Reddy V, Ehrenstein MR, Isenberg DA, et al. CD19+CD24hiCD38hi B cells maintain regulatory T cells while limiting TH1 and TH17 differentiation. *Sci Transl Med*. 2013;5(173):173ra23.
187. Bhat NM, Mithal A, Bieber MM, Herzenberg LA, Teng NN. Human CD5+ B lymphocytes (B-1 cells) decrease in peripheral blood during pregnancy. *J Reprod Immunol*. 1995;28(1):53-60.
188. Gentile T, Borel IM, Angelucci J, Miranda S, Margni RA. Preferential synthesis of asymmetric antibodies in rats immunized with paternal particulate antigens. Effect on pregnancy. *J Reprod Immunol*. 1992;22(2):173-83.
189. Nielsen HS, Witvliet MD, Steffensen R, Haasnoot GW, Goulmy E, Christiansen OB, et al. The presence of HLA-antibodies in recurrent miscarriage patients is associated with a reduced chance of a live birth. *J Reprod Immunol*. 2010;87(1-2):67-73.
190. Malan Borel I, Gentile T, Angelucci J, Pividori J, Guala MC, Binaghi RA, et al. IgG asymmetric molecules with antipaternal activity isolated from sera and placenta of pregnant human. *J Reprod Immunol*. 1991;20(2):129-40.
191. Eblen AC, Gercel-Taylor C, Shields LB, Sanfilippo JS, Nakajima ST, Taylor DD. Alterations in humoral immune responses associated with recurrent pregnancy loss. *Fertil Steril*. 2000;73(2):305-13.
192. Zenclussen AC, Gentile T, Kortebani G, Mazzolli A, Margni R. Asymmetric antibodies and pregnancy. *Am J Reprod Immunol*. 2001;45(5):289-94.
193. Wong LF, Porter TF, Scott JM. Immunotherapy for recurrent miscarriage. *Cochrane Database Syst Rev*. 2014;2014(10):CD000112.
194. Fu J, Li L, Qi L, Zhao L. A randomized controlled trial of etanercept in the treatment of refractory recurrent spontaneous abortion with innate immune disorders. *Taiwan J Obstet Gynecol*. 2019;58(5):621-5.
195. Gounder SS, Abdullah BJJ, Radzuanb NEIBM, Zain FDBMZ, Sait NBM, Chua C, et al. Effect of Abing on NK Cell Population and Their Proliferation at Ex Vivo Culture Condition. *Anal Cell Pathol*. 2018;2018:7871814.
196. Makhseed M, Raghupathy R, Azizieh F, Farhat R, Hassan N, Bandar A. Circulating cytokines and CD30 in normal human pregnancy and recurrent spontaneous abortions. *Hum Reprod*. 2000;15(9):2011-7.
197. Lee S, Kim J, Jang B, Hur S, Jung U, Kil K, et al. Fluctuation of peripheral blood T, B, and NK cells during a menstrual cycle of normal healthy women. *J Immunol*. 2010;185(1):756-62.
198. Park DW, Yang KM. Hormonal regulation of uterine chemokines and immune cells. *Clin Exp Reprod Med*. 2011;38(4):179-85.
199. Landmann RM, Müller FB, Perini C, Wesp M, Erne P, Bühler FR. Changes of immunoregulatory cells induced by psychological and physical stress: relationship to plasma catecholamines. *Clin Exp Immunol*. 1984;58(1):127-35.
200. Espersen GT, Elbaek A, Ernst E, Toft E, Kaalund S, Jersild C, et al. Effect of physical exercise on cytokines and lymphocyte subpopulations in human peripheral blood. *APMIS*. 1990;98(5):395-400.
201. Regan L, Rai R, Saravelos S, Li TC, Gynaecologists RCoO. Investigation and Treatment of Recurrent First-Trimester and one or more Second-Trimester Miscarriage. *Green-top Guideline*. 2021;17 (Peer Review Draft):1-48.
202. Tommy's. Why did I miscarry and was it my fault? 2020 [updated December 14th, 2019].
203. Warwick Clinical Trials Unit. CERM Overview 2020 [Available from: <https://warwick.ac.uk/fac/sci/med/research/ctu/trials/cerm/>].
204. ClinicalTrials.gov. Low Dose Prednisone Therapy in Women With Recurrent Pregnancy Loss. *ClinicalTrials.gov Identifier: NCT04558268*.
205. ClinicalTrials.gov. Low-dose Interleukin-2 in Women With Unexplained Miscarriages (FaCIL-2). *ClinicalTrials.gov Identifier: NCT03970954*.