**An expanded database of Southern Hemisphere surface sediment dinoflagellate cyst assemblages and their oceanographic affinities**

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**Abstract.** Dinoflagellate cyst assemblages present a valuable proxy to infer paleoceanographic conditions, yet factors influencing geographic distributions of species remain largely unknown, especially in the Southern Ocean. Strong lateral transport, sea-ice dynamics and a sparse and uneven geographic distribution of surface sediment samples have limited the use of dinocyst assemblages as a quantitative proxy for paleo-environmental conditions such as sea-surface temperature (SST), nutrient concentrations, salinity and sea ice (presence). In this study we present a new set of surface sediment samples (n=66) from around Antarctica, doubling the number of Antarctic-proximal samples to 100 (dataset wsi\_100) and increasing the total number of Southern Hemisphere samples to 655 (dataset sh\_655). Additionally, we use modelled ocean conditions and apply Lagrangian techniques on all Southern Hemisphere sample stations to quantify and evaluate the influence of lateral transport on the sinking trajectory of microplankton and through that, to the inferred ocean conditions. *K*-means cluster analysis on the wsi\_100 dataset demonstrates the strong affinity of *Selenopemphix antarctica* to sea-ice presence, and *Islandinium* spp. to low-salinity conditions. For the entire Southern Hemisphere, the *k*-means cluster analysis identifies nine clusters with a characteristic assemblage. In most clusters a single dinocyst species dominates the assemblage. These clusters correspond to well-defined oceanic conditions in specific Southern Ocean zones or along the ocean fronts. We find that when lateral transport is predominantly zonal, the environmental parameters inferred from the sea floor assemblages mostly correspond to those of the overlying ocean surface. In this case, the transport factor can thus be neglected and will not represent a bias in the reconstructions. Yet, for some individual sites, e.g., deep water sites or sites under strong current regimes, lateral transport can play a large role. The results of our study further constrain environmental conditions represented by dinocyst assemblages and the location of Southern Ocean frontal systems.

# 1. Introduction

Dinoflagellate cyst assemblages have been increasingly used as a proxy to reconstruct southern high-latitude oceanographic conditions for the late Paleogene to recent (34–0 million years ago; Esper and Zonneveld, 2007; Prebble et al., 2016; Sangiorgi et al., 2018; Bijl et al., 2018a; Hoem et al., 2021a, b; Marschalek et al., 2021). Such reconstructions rely on an accurate understanding of the modern ecological affinities of taxa. For extant species, modern biogeography is statistically linked to surface oceanographic conditions (sea-surface temperature, nutrients, salinity, sea-ice; de Vernal et al., 2005; Zonneveld et al., 2013; Prebble et al., 2013; Mudie et al., 2017; Marret et al., 2020; see Table 1 for an overview). They all assume that the assemblages in a surface sediment sample at a certain location represent the parameters of (and derive from) the sea water directly above the sediment. Transfer function techniques then offer a quantitative approach to directly translate (down-core) assemblages into values of past environmental parameters (e.g., Marret et al., 2001; de Vernal et al., 2005; Esper and Zonneveld, 2007). This requires a sufficiently large training set, with a large number of geographically widespread surface sediment samples, so that the full variety of surface oceanographic conditions is represented.

Constraining oceanographic affinities of dinocyst species has been hindered by the low number and uneven geographic distribution of surface sediment samples from the Southern Ocean, particularly when compared to that in the Northern Hemisphere high latitudes (de Vernal et al., 2001; 2020). Notably scarce are samples from the Antarctic margin, i.e., the region south of the PF (Marret et al., 2020). Among the most dominant species found close to the Antarctic margin, *Selenopemphix antarctica* is generally an environmentally significant taxon, that is commonly associated with (past) Southern Ocean sea-ice presence (Harland and Pudsey, 1999; Marret et al., 2001; Houben et al., 2013). Sporadic occurrences of *S. antarctica* north of the winter sea-ice edge (Esper and Zonneveld 2007) may be linked to northward transport by surface and deep currents from the sea-ice zone (Nooteboom et al., 2019). However, the low number of samples analysed from close to the Antarctic margin makes the sea-ice affinity of *S. antarctica* poorly constrained. Specifically, it is unknown what the regional applicability of *S. antarctica* as a sea-ice indicator may be, and whether whole dinocyst assemblages from sea-ice regions may be more suitable than a single species to constrain past sea-ice conditions.

Previous attempts to apply transfer functions with surface sediment samples from the Southern Ocean focused on sea-surface temperature (SST) reconstructions (Marret et al., 2001; Esper and Zonneveld, 2007). Pleistocene dinocyst assemblages from the subtropical front in the Pacific Ocean yield realistic SST reconstructions for interglacial time intervals that are in agreement with other proxies, but SST reconstructions that are colder than those obtained with other proxies for glacial intervals (Marret et al., 2001). This is largely due to the occurrence of *S. antarctica* in glacial phases, which results in a cold bias in the transfer function output (Prebble et al., 2016). The extensive surface sediment sample set from around New Zealand (Prebble et al., 2013) demonstrated that *S. antarctica* is a rare member of dinocyst assemblages in the subantarctic Pacific, and in that region its presence cannot be easily explained by lateral transport (Nooteboom et al., 2019). This raises questions about whether this species lives exclusively in regions covered with sea ice. Esper and Zonneveld (2007) showed overall good consistency with dinocyst-based SST reconstructions to those from other proxies for Southern Ocean sediment cores spanning the past 140 kyrs, with some potential bias due to selective degradation of dinocysts (Esper and Zonneveld, 2007).

Table 1. Dinocyst compilation efforts. DCA = Detrended correspondence analysis, CCA = Canonical Correspondence analysis, MAT = Modern Analogue Technique

|  |  |  |
| --- | --- | --- |
| **Source** | **# samples** | **Statistical method and aims** |
| Marret et al., 2001 | 81 (Southern Hemisphere) | Multivariate analysis (non-metric multidimensional scaling; MDS) – best analogue method |
| Marret and Zonneveld, 2003 | 835 (global) | DCA, CCA – to explore which environmental variables explain dinocyst assemblages most |
| Esper and Zonneveld, 2007 | 138 (Southern Hemisphere) | CCA, MAT – to explore environment controls on dinocyst distribution |
| Prebble et al., 2013 | 311 (Southern Hemisphere) | Cluster analysis (*k*-means clustering) – to explore environment controls on dinocyst distribution |
| Zonneveld et al., 2013 | 2405 (global) | - |
| Nooteboom et al., 2019 | 2405 (global) | Lagrangian particle trace simulations in ocean model – to demonstrate the effect of lateral transport on dinocyst distributions |
| Marret et al., 2020 | 3636 (global)  595 (Southern Hemisphere) | - |
| This study | 655 (Southern Hemisphere) | Cluster analysis (*k*-means clustering)­ – to explore environment controls on dinocyst distribution |

Lateral transport of sinking particles creates a deviation from the first-order approximation that plankton (or other particles) found in the sediments relate to oceanographic conditions in the surface waters directly overlying the site (e.g., Zonneveld et al., 2013). For instance, Nooteboom et al. (2019) showed that lateral transport can affect the location of deposition, under realistic assumptions on sinking speed of pelagic particles in their descent towards the ocean floor. The effect of lateral transport on sinking particles is large in places with deep (>1 km) waters and strong currents, where currents flow meridionally (because that is the main direction of environmental gradients), and also where turbulence occurs during sinking (Nooteboom et al., 2019). Thus, the potential for lateral transport of pelagic particles should be considered when using sediment to assess surface water conditions at the site.

In this study, we present dinocyst assemblage data from a new surface sediment sample set predominantly from the Antarctic margin. This sample set fills a clear gap in previously underrepresented ice-proximal Southern Hemisphere surface sediment samples. Moreover, the wide geographic spread of these new samples allows investigation of longitudinal differences in ice-proximal dinocyst assemblages. Adding these to existing surface sediment samples of the Southern Hemisphere allows an improved assessment of biogeographic and oceanographic affinities of Southern Hemisphere dinocyst assemblages. We compare dinocyst assemblages to the following oceanographic parameters: SST, salinity, nutrients (nitrate and silicate concentrations), and sea ice cover. We assess these by analysing model output data, specifically for the surface water directly overlying the sites, and for the surface water in the location of origin of simulated particles that descended to those sites. This allows an evaluation of the extent to which lateral transport affected the paleoceanographic affinities of sedimentary dinocyst assemblages.

# 2. Modern oceanographic conditions

## 2.1 The Southern Ocean

The dominant oceanographic feature in the Southern Ocean is the Antarctic Circumpolar Current (ACC; Marshall and Speer, 2012). Without land barriers and (partly) driven by strong Southern Hemisphere Westerly Winds, the ACC flows clockwise around Antarctica and is the world’s strongest ocean current (>140 Sv; Orsi et al., 1995, Park et al., 2019 and reference therein). The flow speed is the highest at the ocean fronts associated to the ACC, which introduces strong environmental gradients and creates quasi-latitudinal zones with characteristic environmental conditions (Fig. 1; Orsi et al., 1995). From north to south, the main fronts are the Subtropical Front (STF), the Subantarctic Front (SAF), and the Polar Front (PF; Fig. 1).

In general, the STF is considered the northern boundary of the ACC (Orsi et al., 1995). Surface water temperature changes by ~4-5°C and salinity changes by ~0.5 across the 0.5° latitudinal band that represents the STF, which separates warm, nutrient-poor Subtropical Surface Waters of the subtropical gyres to the north from colder and nutrient-rich Subantarctic Surface Waters of the Subantarctic Zone to the south (SAZ; see Fig. 1).The SAF separates the SAZ from the Polar Frontal Zone (PFZ), where Subantarctic Surface Waters are subducted to form Antarctic Intermediate Waters (Orsi et al., 1995). The PF separates these waters from colder (<2°C) Antarctic Surface Waters that form the Antarctic Zone (AZ). The typical high-nutrient-low-chlorophyll-signature of the AZ is caused by a scarcity of light and micro-nutrients which limit the utilization of available macro-nutrients. Overall, this leads to a dominance of silicate-based phytoplankton (e.g., diatoms) over calcareous species (e.g., coccolithophores). In the AZ, (seasonal) sea ice plays a major role as it can strongly modulate light and nutrient availability (Mitchell et al., 1991). Sea ice is mostly found in areas close to the Antarctic continent, the Ross and Weddell Seas, and Prydz Bay, where brine rejection promotes the formation of Antarctic Bottom Water, which then sinks to the deep ocean (Adkins, 2013; Talley, 2013, Solodoch et al., 2022).

Sea-ice dynamics differ between the Ross and Weddell Seas. The Weddell Sea has a strong, dynamic gyre transports sea ice and icebergs out of the Weddell Sea into the South Atlantic, an area known as Iceberg Alley (Orsi et al., 1995). The most prominent feature in the Ross Sea is the Ross Sea polynya, open patches in the sea ice, maintained by katabatic winds (Rhodes et al., 2009; Bromwich et al., 1992).

Figure 1: Compilation of maps of modern annual average environmental parameters in the Southern Ocean. a) Southern Ocean SST and approximate winter and summer sea ice extent (modified from Crosta et al., 2021 and Spreen et al., 2015–2021). b) Surface (1 meter) salinity (psu), nitrate and silicate concentrations (μmol/kg). Data from the World Ocean Atlas (Locarnini et al., 2018). Maps were produced using ggplot2 in R (Wickham, 2016).

## 2.2 Lateral transport

Taking (past) lateral transport into account in the interpretation of sediment proxies remains difficult, as the quantification of transport comes with large uncertainties and assumptions regarding (past) ocean current strengths and directions, particle aggregation and size and – connected to this – their sinking speed (Nooteboom et al., 2019). Recent efforts have attempted to better constrain the influence of lateral transport on sedimentary microplankton assemblages by applying Lagrangian particle trace experiments in ocean model simulations with a spatial resolution high enough to represent an eddying flow (0.25° and finer; Nooteboom et al., 2019). In the Southern Ocean especially, it is thought that the ACC strength and sea-ice dynamics can assert a large influence on particle transport. Here, recent modelling efforts could explain unexpected, rare occurrences of species (Nooteboom et al., 2019) outside their common habitat. More specifically, findings of *Selenopemphix antarctica* north of the SAF could be attributed to latitudinal transport from the AZ, thus helping to better constrain and interpret (past) ecological affinities of single species and explain their rare presence in unexpected locations. This demonstrated the potential influence lateral transport may play in shaping microplankton assemblages, and, through that, the inferred surface oceanographic affinities of microplankton species.

# 3. Methods

## 3.1 New surface sediment samples from around Antarctica

In total, we examined 66 new surface sediment samples located close to the Antarctic continent and 7 samples located north of the modern PF (Fig. 2; Table 2). For some new samples, existing age models confirm a modern or (late?) Holocene age (e.g., Hartman et al., 2018; Wilson et al., 2018; Armand et al., 2018; Behrens et al., 2019). However, for many samples, no absolute age determination was available, but the use of the box coring technique (Table 2) to retrieve the sediment-water interface together with the dinocyst assemblages and palynofacies found (e.g., presence of more labile organic matter, such as amorphous organic matter and chitin remains) suggest a tentative modern-to-late Holocene age. Previous studies of modern surface sediments used an age-cut off of 7 ka (Prebble et al., 2013). We are confident that the data added here have a modern age similar to those of the data we compare ours to. We do note that this surface sediment sample set averages out potential environmental changes that occurred in the Holocene.

All samples, but particularly those from the Antarctic continental margin, may have contributions of reworked palynomorphs from older sediments (Bijl et al., 2018b). Most reworking is easily recognized (and excluded from further analysis) because these species are extinct. For extant taxa, such as *Spiniferites ramosus, Operculodinium centrocarpum,* and *Nematosphaeropsis labyrinthus*, it is impossible to assess from their appearance whether they are reworked or in situ. This means that caution should be taken when interpreting rare occurrences of these taxa.

A total of 46 productive samples are from south of 60°S, and located in the Ross Sea (28 samples), the Sabrina coast (17 samples), Prydz Bay (6 samples) and Weddell Sea (4 samples), almost doubling the number of existing samples from Antarctic ice-proximal sites to 100. We also add the first dinocyst assemblages from south of 70°S (Ross Sea, Fig. 2), yet some ice-proximal areas remain uncovered, such as the Weddell Sea and offshore Dronning Maud Land, as well as most of the West Antarctic continental margin. The seven additional samples are from north of the PF, located close to Crozet Island, on the Kerguelen Plateau and along the west coast of South America (Fig. 2).

Most samples are in (relatively) close proximity to land, including the coast of New Zealand, the western coasts of Southern Africa and Australia and both sides of South America. The Atlantic has the highest spatial coverage, whereas the Pacific and Indian Oceans appear underrepresented.

Figure 2: Overview map of all available surface sediment data for the Southern Hemisphere. Blue dots represent samples that have been previously published (Marret et al., 2020), black dots are newly analysed samples from this study, and white dots are new samples that did not yield enough dinocysts to be included in further analyses. Also plotted are ACC frontal systems (dark green lines, STF = Subtropical Front, SAF = Subantarctic Front, PF = Polar Front). Winter and Summer sea ice is presented in light and dark grey areas, respectively, and is derived from Spreen et al. (2015–2021). Maps were produced using ggplot2 in R (Wickham, 2016).

Table 2. List of newly counted samples

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Station** | **Sample method** | **Latitude** | **Longitude** | **Adjusted Latitude in ocean model** | **Adjusted Longitude in ocean model** | **Water depth (m)** | **in SH\_655** | **in WSI\_100** |
| ANT 33\_RS78 | Box core | -78.6939 | -163.6669 | NA | NA |  | n | n |
| ANT 33\_A1-08 | Box core | -78.1719 | -165.7881 | -76.9000 | -165.7881 | 498 | y | y |
| ANT 33\_A1-07 | Box core | -78.1681 | -163.0389 | -76.9000 | -163.0389 | 679 | y | y |
| ANT 33\_A1-10 | Box core | -77.9681 | -171.3731 | NA | NA |  | n | n |
| ANT 33\_A1-20 | Box core | -77.6661 | 165.8850 | -76.9000 | 165.8850 | 590 | y | y |
| ANT 31\_JB01 | Box core | -77.5883 | 165.5694 | -76.9000 | 165.5694 | 774 | y | y |
| ANT 33\_A1-05 | Box core | -77.3961 | -162.6781 | -76.9000 | -162.6781 | 658 | y | y |
| ANT 32\_RB08B | Box core | -77.3089 | 179.8525 | -76.9000 | 179.8525 | 670 | y | y |
| ANT 32\_RB11B | Box core | -77.2694 | 174.5997 | -76.9000 | 174.5997 | 494 | y | y |
| ANT 33\_A1-15 | Box core | -77.1269 | 174.9639 | -76.9000 | 174.9639 | 400 | y | y |
| ANT 32\_RB07B | Box core | -77.0497 | -178.9003 | -76.9000 | -178.9003 | 628 | y | y |
| ANT 32\_RB06B | Box core | -76.7133 | -178.2403 |  |  | 619 | y | y |
| BC22 | Box core | -76.6930 | 169.0780 |  |  | 800 | y | y |
| ANT 33\_A1-18 | Box core | -76.4211 | 167.7239 |  |  | 743 | y | y |
| ANT 32\_RB05B | Box core | -76.4053 | -177.7194 |  |  | 606 | y | y |
| ANT 32\_RB03B | Box core | -75.7467 | -176.8697 |  |  | 610 | y | y |
| ANT 32\_RB02B | Box core | -75.4233 | -176.4858 |  |  |  | n | n |
| ANT 31\_JB04 | Box core | -75.3011 | 172.3725 |  |  | 511 | y | y |
| ANT 33\_I5 | Box core | -75.0869 | 165.0481 |  |  | 1174 | y | y |
| ANT 31\_R09 | Box core | -75.0033 | 165.9980 |  |  | 1032 | y | y |
| ANT 31\_R08 | Box core | -75.0030 | 165.0120 |  |  |  | n | n |
| ANT 31\_R10 | Box core | -75.0022 | 167.0019 |  |  | 636 | y | y |
| ANT 35\_R1-03 | Box core | -74.9953 | 168.3578 |  |  | 349 | y | y |
| ANT 31\_R11 | Box core | -74.9492 | 167.8056 |  |  | 449 | y | y |
| ANT 31\_R14 | Box core | -74.9350 | 164.8047 |  |  | 901 | y | y |
| ANT 31\_R18 | Box core | -74.9130 | 163.7650 |  |  |  | n | n |
| ANT 31\_R02 | Box core | -74.7850 | 165.1330 | -74.8022 | 165.1150 | 719 | y | y |
| ANT 31\_R05 | Box core | -74.7767 | 167.7775 |  |  | 586 | y | y |
| ANT 31\_JB05 | Box core | -74.7553 | 173.1881 |  |  | 497 | y | y |
| ANT 32\_RB16B | Box core | -74.5136 | 175.1217 |  |  | 478 | y | y |
| ANT 33\_A2-02 | Box core | -74.2061 | 170.1181 |  |  |  | n | n |
| ANTA02\_AV43 | Box core | -74.1410 | 166.0830 | -74.1325 | 166.1983 |  | y | y |
| ANT 33\_A2-03 | Box core | -73.7011 | 170.9789 |  |  | 588 | y | y |
| ANT 33\_A2-05 | Box core | -72.6050 | 172.4369 |  |  | 546 | y | y |
| ANT 31\_R19 | Box core | -72.2544 | 170.3944 |  |  |  | n | n |
| ANT 32\_AB18B | Box core | -71.8986 | -128.1450 |  |  | 3463 | y | y |
| ANT 30\_IS-01 | Box core | -69.2594 | 76.3775 |  |  | 260 | y | y |
| ANT 29\_P6-12 | Box core | -68.9095 | 75.4897 |  |  |  | n | n |
| ANT 29\_P7-16 | Box core | -68.3836 | 76.1972 |  |  | 558 | y | y |
| ANT 29\_P3-09 | Box core | -67.5123 | 68.0119 |  |  |  | n | n |
| ANT 29\_P7-14 | Box core | -67.4378 | 77.1840 |  |  | 311 | y | y |
| ANT 29\_PA-05 | Box core | -67.2357 | 76.1799 |  |  | 353 | y | y |
| ANT 29\_PA-01 | Box core | -67.2061 | 71.2824 |  |  | 487 | y | y |
| NBP1402\_KC13 | Kasten core | -66.8730 | 118.2400 |  |  | 646 | y | y |
| NBP1402\_KC14 | Kasten core | -66.8730 | 118.2370 |  |  | 643 | y | y |
| NBP1402\_KC42 | Kasten core | -66.4840 | 120.3320 |  |  | 610 | y | y |
| U1357B | Piston core | -66.4133 | 140.4262 |  |  | 1017 | y | y |
| NBP1402\_KC9 | Kasten core | -66.3610 | 119.8620 |  |  | 683 | y | y |
| NBP1402\_SMG51 | Kasten core | -66.3240 | 120.4650 |  |  | 450 | y | y |
| NBP1402\_27A | Kasten core | -66.1850 | 120.5040 |  |  | 544 | y | y |
| NBP1402\_KC27B | Kasten core | -66.1850 | 120.5040 |  |  | 547 | y | y |
| NBP1402\_JKC53 | Kasten core | -66.1840 | 120.5030 |  |  | 545 | y | y |
| NBP1402\_MC\_45 | Multicore | -66.1830 | 120.5010 |  |  | 537 | y | y |
| NBP1402\_KC57 | Kasten core | -66.1289 | 120.4640 |  |  |  | n | n |
| ANT 29\_P3-03 | Box core | -66.0000 | 67.8115 |  |  | 2689 | y | y |
| SABRINA\_KC11 | Kasten core | -65.1320 | 120.0490 |  |  | 2611 | y | y |
| SABRINA\_PC7 | Piston core | -65.1310 | 120.0490 |  |  | 2611 | y | y |
| SABRINA\_PC6 | Piston core | -64.8190 | 120.1430 |  |  | 3012 | y | y |
| SABRINA\_KC14 | Kasten core | -64.5390 | 116.6400 |  |  | 2100 | y | y |
| SABRINA\_KC2 | Kasten core | -64.4710 | 115.6230 |  |  | 2161 | y | y |
| SABRINA\_KC3 | Kasten core | -64.4630 | 115.0430 |  |  | 1862 | y | y |
| U1361A | Piston core | -64.4100 | 143.8900 |  |  | 3454 | y | y |
| ANT 30\_D2-10 | Box core | -62.9600 | -52.2667 |  |  | 974 | y | y |
| ANT 30\_D1-07 | Box core | -62.2428 | -56.5769 |  |  | 3548 | y | y |
| ANT 30\_D2-05 | Box core | -61.5850 | -54.1500 |  |  | 450 | y | y |
| ANT 30\_DA-02 | Box core | -60.8350 | -49.5817 |  |  | 2592 | y | y |
| MD11-3352CQ | Box core | -50.5672 | 68.3858 |  |  |  | n | n |
| MD12-3396CQ | Box core | -47.7313 | 86.6952 |  |  |  | n | n |
| MD04-2716CQ | Box core | -46.1650 | 52.9267 |  |  |  | n | n |
| MD12-3401CQ | Box core | -44.6788 | 80.3930 |  |  | 3445 | y | n |
| MD04-2714CQ | Box core | -43.3983 | 49.8200 |  |  | 2300 | y | n |
| MD07-3100 | Calypso | -41.6022 | -74.9567 |  |  | 1609 | y | n |
| MD07-3098 | Calypso | -40.9300 | -75.0303 |  |  | 3055 | y | n |

## 3.2 Palynology

Samples were processed for palynological analysis following standard procedures of the Laboratory of Paleobotany and Palynology (e.g., Bijl et al., 2018b). Sediment samples were freeze-dried, crushed, and weighed, before a tablet of a known amount (19855) of *Lycopodium clavatum* was added to quantify dinoflagellate cyst concentrations. Samples were then treated with 30% hydrochloric acid (HCl) and 38% cold hydrofluoric acid to respectively dissolve carbonates and silicates. A second round of 30% HCl treatment removed fluoric gels that might have formed, before the acid-sample mix was centrifuged and decanted. The (organic) residue was sieved at 250 and 10 μm and an ultrasonic bath assisted in disaggregating organic particles. The residue was mounted on glass sides in glycerin jelly. Dinoflagellate cysts were then counted under light-transmitted light microscopy.

Where possible, specimens were identified to species level at 400x magnification. Taxonomy followed that cited in Williams et al. (2017), and informal taxonomic descriptions in Esper and Zonneveld (2007) for some subspeciation. To facilitate the integration of our analyses with previously published datasets (Prebble et al., 2013; Marret et al., 2020), we grouped different dinocyst species the same way. This led to the following taxonomic groupings:

1. *Brigantedinium* spp.: *Brigantedinium cariacoense*, *Brigantedinium simplex*, and *Dubridinium capitatum*
2. *Protoperidiniacean* cysts: *Lejeunecysta* spp., cysts of *Protoperidinium stellatum*, *Quinquecuspis concreta*, and *Votadinium calvum*
3. *Nematosphaeropsis labyrinthus*: all *Nematosphaeropsis* species
4. *Spiniferites hyperacanthus* was combined with *Spiniferites mirabilis*
5. *Spiniferites bulloides* was combined with *Spiniferites ramosus*
6. *Spiniferites belerius* was combined with *Spiniferites membranaceus*
7. Cysts of *Protoperidinium nudum* were assigned to *Selenopemphix quanta*
8. All *Echinidinium* species were grouped
9. Cysts of *Polarella glacialis* were counted but not included in the dinocyst sum
10. *Selenopemphix* sp. 1 (Esper and Zonneveld, 2007) was counted separately from *Selenopemphix antarctica*, but these were grouped in the cluster analyses of wsi\_100 and sh\_655.
11. *Gymnodinium* spp.: all species of *Gymnodinium*

In general, up to 100 (200 when possible) cysts (excluding *Polarella*) were counted per sample. This number captured the full diversity in our samples (see Fig. S1 for rarefaction analysis). As minimal cutoff, 13 samples with total counts <25 were excluded from further analyses. In this paper, we describe dinocyst abundances qualitatively as follows: rare = <1% of assemblage, few = 1% to <10%, common = 10% to <25%, abundant = 25% to <50%, dominant = >50%.

We excluded the cysts of *Polarella glacialis* in further data analyses for the following reasons: (1) ambiguity about whether the absence of *Polarella* in other studies is the result of its real absence or is due to other reasons. For instance, a coarser than 10 microns mesh size often used for sieving samples could result in the loss of *Polarella* due to its small size. This obscures presentation of biogeographic patterns for this species. (2) The short stratigraphic range of the species (few hundred years, both in samples from the Ross Sea and from Wilkes Land; Sangiorgi, pers. obs. 2022, and Hartman pers. comm. 2022, respectively) in the cores analyzed so far could imply that its presence in the sediment is linked to preservation exclusively in relatively freshly deposited samples. It remains possible that *Polarella* abundance can be used as an indicator for sea ice if they are found in older sediments. With the existing knowledge, the potential of *Polarella* in paleoreconstructions is very limited. (3) This species is often the dominant taxon in modern samples but absent in older samples. Since our analyses are based on percent abundance, including *Polarella* dilutes the percent abundance of the other taxa in the modern dataset, thereby skewing the comparison with older samples that lack *Polarella*. Since this study is intended to improve dinocyst assemblage-based tools for paleoreconstructions, adding a modern cyst species that has no stratigraphic record does not improve this tool. The *Polarella* counts are available in the dataset on Github (Bijl, 2022), but not included in relative abundance calculations for the rest of this paper.

## 3.3 Datasets

To evaluate regional biogeographic consistency in ice-proximal assemblages, we created a data subset containing only sample locations with sea ice in the overlying waters. For this, we used modern modelling data and the criterion of Antarctic winter (June, July, August; JJA) sea-ice presence > 0 (see section 3.5 below). This data subset (“wsi\_100”) contains 100 samples. We then added all new samples (Table 2) to the Southern Hemisphere samples of the latest existing global compilation of dinocyst surface sediments (Marret et al., 2020). This yields 655 samples in total (dataset “sh\_655”).

Data from this paper are stored at Github (Bijl, 2022). The environmental data corresponding to locations of wsi\_100 and sh\_655 are obtained from the Nucleus for European Modelling of the Ocean (NEMO; Madec, 2016) ocean model, which is coupled to the MEDUSA-2.0 biogeochemical model (Yool et al., 2013), following the approach as detailed in Section 3.5.

## 3.4 Cluster Analyses

Cluster analysis of surface samples allowed us to explore spatial relationships between dinocyst assemblages and surface oceanographic conditions. These relationships in the modern day can be used to infer paleoceanographic conditions from fossil assemblages. Following the method of Prebble et al. (2013), we performed *k*-means cluster analyses on the relative abundance of dinocyst species for the two datasets. We ran 100 replicates for each cluster solution and chose the most stable clustering. The evaluation of the clustering of wsi\_100 and sh\_655 can be found in Fig. S2 and S3. All analyses were done in R using the package dbscan (Hahsler et al., 2019).

## 3.5 Surface oceanographic parameters: surface versus transport data – particle tracking

To identify environmental conditions associated with the different clusters, we analysed SST (°C), sea-ice presence (from 0–1, fraction of grid cell that is sea ice-covered), salinity (psu), nitrate (mmol/m3) and silicate (mmol/m3) in the surface waters at the location of the surface sediment samples. For this, we made use of the ORCA0083-N06 output from the NEMO ocean model (Madec, 2016), which is coupled to the MEDUSA-2.0 biogeochemical model (Yool et al., 2013). We note that the density of environmental observations at high latitudes is low, and as such the verification of the model output comes with an unknown degree of uncertainty. To evaluate the impact of particle transport, we compared the conditions in the overlying surface water to those of modelled backtracked surface water locations. The NEMO model provides a global three-dimensional flow field with 5-daily output and has 1/12° horizontal resolution with 75 unevenly distributed vertical layers, properly reflecting the transport of Lagrangian particles by mesoscale and sub-mesoscale eddies (Nooteboom et al., 2020; Qin et al., 2014). In the first run, we recorded the surface water conditions of the overlying surface sediment locations, every three days from 2000-01-03 until 2003-12-30. In the second run, we released virtual particles every three days for four years (from 2000-01-03 until 2003-12-30, using the OceanParcels Lagrangian Simulator; Lange and van Sebille, 2017; Delandmeter and van Sebille, 2019) at the sediment surface location and backtracked them to the ocean surface using a 25 m day-1 sinking speed. This sinking speed is faster than the sinking of individual dinoflagellate cysts, as we assume some degree of particle aggregation (Nooteboom et al., 2019).

The southern boundary of the NEMO model set up was at 77 °S. As a result we had to relocate 11 samples that were originally located more southward to a latitude of 76.9 °S for the model run (Table 2), assuming minimal effects on environmental and transport conditions. Similarly, nine sample locations were too close to land and, due to the model resolution, appeared on land in the model. Thus, their location was adjusted (Table 2) to the nearest ocean site by minimally shifting the latitude and/or longitude.

# 4. Results

## 4.1 New Southern Hemisphere surface samples

The 66 newly analyzed productive samples contain 24 different species (Fig. 3). Although we counted *Selenopemphix antarctica* and *Selenopemphix* sp. 1 (after Esper and Zonneveld, 2007) separately, the two species were not counted separately in previously published datasets (most of the data simply refer to *Selenopemphix antarctica)*. In our cluster analysis, the two species are hence combined as *Selenopemphix antarctica*. All new samples from Antarctic-proximal sites are dominated by heterotrophic peridinioid (P) cysts (Fig. 3). The most equatorward new samples (Fig. 3b, i) show abundant to dominant *Brigantedinium* spp., *Impagidinium* spp., *Nematosphaeropsis labyrinthus* and *Spiniferites* spp. Typical ice-proximal species *S. antarctica* and *Nucicla umbiliphora* (e.g., Hartman et al., 2018) are rare or absent.

In the samples from the Weddell Sea (Fig. 3b, ii), *Islandinium minutum* is dominant, with very low contributions from *Selenopemphix antarctica*, *Brigantedinium* spp. or *Impagidinium pallidum*. The samples from Prydz Bay (Fig. 3b, iii) have a rather diverse assemblage with abundant *Selenopemphix antarctica* and common *Selenopemphix* sp.1, *Nucicla umbiliphora*, *Islandinium minutum* and *Brigantedinium* spp. In the sample further north and outside the bay (ANT29\_P3\_03), *Impagidinium pallidum* dominates, with common *Operculodinium centrocarpum*. In the samples from Sabrina Coast (Fig. 3b, iv), *Selenopemphix antarctica* sp. 1 is abundant, with rare to few *Selenopemphix* sp.1, *Nucicla umbiliphora*, and *Islandinium minutum*. A striking distinction can be made in samples from Sabrina Coast, with one part having abundant *Brigantedinium* spp. and the other part having abundant *Impagidinium pallidum* (>30%), common *Operculodinium centrocarpum*, and few *Pyxidinopsis reticulata* and *Spiniferites mirabilis.* Most of the Ross Sea samples contain a quite uniform assemblage (Fig. 3b, v), with dominant *Selenopemphix antarctica* sp. 2 and/or *Nucicla umbiliphora*. In most samples, one of the species is clearly dominant. *Selenopemphix antarctica*, *Islandinium minutum*, *Brigantedinium* spp. and *Operculodinium centrocarpum* are rare. In one sample located outside of the Ross Sea (ANTA02\_AV43), *Brigantedinium* spp. dominates, with common *Impagidinium pallidum* and *Selenopemphix antarctica*.

Figure 3: Overview of new counted samples. a) Map of locations of new samples. Map was produced using ggplot2 in R (Wickham, 2016). b) dinocyst assemblages and absolute abundances (in cysts/gram dry weight) for the new samples from the subantarctic/subtropical zone (i), Antarctic Peninsula (ii), Prydz Bay (iii), Sabrina/Wilkes Coast (iv) and Ross Sea (v).

## 4.3 Clustering of the wsi\_100 dataset: the Antarctic proximal sites

*K*-means clustering of the dinocyst assemblages in the surface sediment samples allows grouping of samples with similar dinocyst assemblages. The most stable solution for the wsi\_100 dataset has 5 clusters (Fig. S2; Fig. 4).

1. **Cluster 1** (*n* = 41) is dominated by *Selenopemphix antarctica*, with few *Brigantedinium* spp., *Operculodinium centrocarpum*, *Nucicla umbiliphora*, *Cryodinium meridianum* and *Islandinium minutum*. Most samples are from the Ross Sea, others come from along the Sabrina coast and in West Antarctica (Fig. 4b, dark blue).
2. **Cluster 2** (*n* = 9) consists of nine samples and is strongly dominated by *Islandinium minutum* (Fig. 4a), making up almost 100% of the average assemblage in this cluster. Only very few other species appear in this cluster, most often *Brigantedinium* spp. and *Selenopemphix antarctica*. Samples in this cluster are located in the Weddell Sea (Fig. 4b, blue).
3. **Cluster 3** (*n* = 13) has samples dominated by *Brigantedinium* spp. Other species, such as *Selenopemphix antarctica*, *Cryodinium meridianum*, *Impagidinium pallidum* and *Islandinium minutum* are rare. Except for one sample in the Ross Sea, Cluster 3 samples are located the furthest away from the Antarctic continent and north of 70°S, mainly appearing in the Pacific Ocean sector (Fig. 4b, green).
4. **Cluster 4** (*n* = 8) has abundant *Impagidinium pallidum*, common *Selenopemphix antarctica* and few *Brigantedinium* spp., *Operculodinium centrocarpum*, and *Pyxidinopsis reticulata*. Samples from this cluster appear everywhere around Antarctica with no clear restrictions or affinities (Fig. 4b, bright green). They are located within the Ross and Weddell Seas but also further away from the Antarctic continent.
5. **Cluster 5** (*n* = 29) has abundant to dominant *Selenopemphix antarctica*. *Brigantedinium* spp. is common, as are *Nucicla umbiliphora*, *Cryodinium meridianum* and *Islandinium minutum*. Samples in this cluster are located in the Ross Sea, Prydz Bay and along Sabrina Coast, mostly close to the coast (Fig. 4b, yellow).

Figure 4: 5-cluster solution for the wsi\_100 dataset (considering all sample locations that experience surface winter (JJA) sea ice). A) The dinocyst assemblages for each cluster (showing only the main 12 dinocyst species). Black horizontal line depicts the median, the coloured box the 25% and 75% quantile, and the whiskers the 95% confidence interval. b) locations of samples color-coded by cluster. Also plotted are frontal systems (blue lines, STF = Subtropical Front, SAF = Subantarctic Front, PF = Polar Front). Map was produced using ggplot2 in R (Wickham, 2016).

## 4.2 Clustering of the sh\_655 dataset

The 9-cluster result of our *k*-means cluster analyses offers the most stable solution for the sh\_655 dataset (Fig. 5; Fig. S3). Below is the median dinocyst assemblage for each cluster.

1. **Sant Cluster (***n* = 64): *Selenopemphix antarctica* dominates (Fig. 5a). *Nucicla umbiliphora*, *Islandinium minutum*, *Cryodinium meridianum* and *Brigantedinium* spp. are common. *Impagidinium pallidum* and *Operculodinium centrocarpum* are rare. All 64 samples in this cluster are located near the Antarctic continent, such as in the Ross Sea, offshore the Sabrina Coast and in Prydz Bay (Fig. 5b).
2. **Imin Cluster (***n* = 9): Dinocyst assemblages in this cluster are strongly dominated by *Islandinium minutum* (Fig. 5a). Other species such as *Brigantedinium* spp. are rare. Although only very few samples (9) belong to this cluster, all located in the Weddell Sea (Fig. 5b). These samples cluster consistently separate from the other samples in all cluster solutions (see supplementary figures). This cluster identical to Cluster 2 in the wsi\_100 dataset (Fig. 4).
3. **high-Brig Cluster (***n* = 86): This cluster is dominated by *Brigantedinium* spp. Assemblages are quite diverse, with few *Selenopemphix quanta*, various *Impagidinium* species, *Nematosphaeropsis labyrinthus*, *Operculodinium centrocarpum* and *Spiniferites* species.
4. **Nlab Cluster (***n* = 72): *Nematosphaeropsis labyrinthus* is abundant, with common *Brigantedinium* spp. Several other species, such as *Impagidinium* spp., *Selenopemphix quanta*, *Selenopemphix* *nephroides*, *Operculodinium centrocarpum*, *Pyxidinopsis reticulata, Dalella chathamensis* and *Spiniferites mirabilis* occur in low abundance. Samples are located between 40 and 60°S, most prominently south(-west) of Tasmania and New Zealand, but also in the Indian Ocean and along the South American west coast.
5. **low-Brig Cluster (***n* = 120): *Brigantedinium* spp. is common in this cluster. Samples also contain few to common *Operculodinium centrocarpum*, *Nematosphaeropsis labyrinthus, Impagidinium* and *Spiniferites* species as well as *Selenopemphix quanta* and *Selenopemphix* *nephroides*. Other P-cysts such as *Selenopemphix antarctica*, *Nucicla umbiliphora*, *Islandinium minutum* and *Cryodinium meridianum* are rare. The sample locations have a wide geographic spread, but many are located around New Zealand (closer to the coast than samples from other clusters), south of Australia and around Tasmania (but in low numbers), in mid-latitudes along the west coast of Africa, and along the west coast of South America. Samples can occasionally be found in the open ocean, for example the Indian Ocean or close to Antarctica.
6. **low-Ocen Cluster (***n* = 115): no single species clearly dominates the assemblage, yet *O. centrocarpum* is common to abundant. Samples in this cluster are diverse. Samples contain few *Brigantedinium* spp., while other species have strongly variable abundance, such as *Impagidinium pallidum*, other *Impagidinium* species, *Nematosphaeropsis labyrinthus* and *Spiniferites* spp. The geographic distribution of samples within this cluster is scattered; samples are from the Ross Sea and all along ice-proximal sites around Antarctica, but also from mid- and low latitudes, coastal areas and open ocean conditions. No clear geographical distribution is apparent.
7. **Iacu Cluster (***n* = 58): *Impagidinium aculeatum* is abundant. Samples also contain few other *Impagidinium* species, *Nematosphaeropsis labyrinthus*, *Operculodinium centrocarpum*, and *Brigantedinium* and *Spiniferites* species. The samples of this cluster are mostly found between 40° and 20°S, although some samples are located even closer to the equator (in the Atlantic Ocean).
8. **High-Ocen Cluster (***n* = 72): This cluster is dominated by *Operculodinium centrocarpum* with few *Brigantedinium* spp., *Nematosphaeropsis labyrinthus*, *Spiniferites mirabilis* and *Spiniferites* spp. Many other species occur in low abundance. Most samples of this cluster can be found close to the continental coasts of South Africa, Australia, and South America.
9. **Spin Cluster (***n* = 59): *Spiniferites* species are abundant, with few *Brigantedinium* spp., *Pyxidinopsis reticulata*, *Impagidinium* species (*I. aculeatum*, *I. patulum* and *I. sphaericum*) and *Selenopemphix quanta*. Most samples are located along the north-west coast of Australia and in low latitudes in the eastern Indian Ocean.

## 4.4 Comparison between clusters and environmental parameters

### 4.4.1 wsi\_100: directly overlying water

Overall, differences in oceanographic conditions are relatively small when considering the samples from the Antarctic proximal sites: nutrients are overall high, winter sea-ice presence is almost 100% and sea-surface temperature is low (average -1.5°C), but some differences between the clusters can be identified (Fig. 6). Salinity is generally between 33.6 and 34 psu in samples from all clusters, except for Cluster 2 that has a lower median of 32.3 psu. Cluster 3 includes samples within the largest SST range (-1.4 – 0.2°C; Fig. 6) and lowest sea-ice presence (median <0.7) compared to the other clusters. Cluster 1, 4 and 5 have overall very similar conditions. This means that the common *Impagidinium pallidum* in samples from Cluster 4 cannot be directly associated with any environmental parameters considered here.

Figure 5: 9-cluster solution for the sh\_655 dataset. A) The dinocyst assemblages for each cluster (showing only the main 20 dinocyst species). Line, box and whiskers as in Fig. 3. b) locations of samples color-coded by different clusters and frontal systems (blue lines, STF = Subtropical Front, SAF = Subantarctic Front, PF = Polar Front). Map was produced using ggplot2 in R (Wickham, 2016).

Figure 6: Environmental parameters for the 5-cluster solution for the wsi\_100 dataset. The median (black line) of all observations per cluster (1–5), with the 25% and 75% quantile as box and whiskers as 95% confidence interval. Single points represent outliers.

4.4.2 sh\_655 – assessing the effect of lateral transport on environmental affinities

To better attribute environmental parameters to the clusters in sh\_655, we compare the overlying surface water conditions of each location with that of modelled lateral particle (dinocyst) transport area due to ocean currents. Taking lateral transport into account, the source region for particles (dinocysts) is, for some samples, non-randomly offset from the directly overlying water. In the Southern Ocean south of the Subtropical Front, most transport is zonal and not meridional, because of prevailing zonal direction of currents. Regardless of their assigned cluster, all samples that are located between 40 and 60°S have a westward-displaced surface water source region (Fig. 7). For the Imin cluster, the source regions from the lateral transport are confined to the Weddell Sea. For the Sant cluster, the source region of samples in the Ross Sea are not displaced much when lateral transport is accounted for. Dinocysts in the samples from Sabrina Coast and Prydz Bay are originated slightly more eastwards. In general, for the Iacu cluster, the source regions are large, but do not show a clear bias to latitudinal or longitudinal transport. These source regions reflect ocean-current dictated provincialism (Fig. 7) as recently indicated by similar particle trace experiments (Nooteboom et al., 2022).

Figure 7: Location and source regions of samples. Overview map of sample locations (filled dots) color-coded by cluster with modelled backtracked transport surface-ocean origin (open dots). Also plotted are frontal systems (blue lines, STF = Subtropical Front, SAF = Subantarctic Front, PF = Polar Front). Map was produced using ggplot2 in R (Wickham, 2016).

We compare the differences in oceanographic conditions in the median overlying surface water for each cluster in the sh\_655 dataset. This can be further compared to the median conditions of the wider region after taking lateral transport into account (Fig. 8). Most clusters have distinct differences in oceanographic conditions of the directly overlying water. The differences between the oceanographic conditions in the overlying water and the lateral transport region are overall small for most clusters and parameters, but some peculiarities do appear:

* In the Sant cluster, there are hardly any differences between the directly overlying water and the lateral transport area. The average SST is -1.7°C, and the median sea-ice presence is at 100%. Here, the transport data shows a much larger range (below 50%) than the overlying data (below 75%). Salinity is around 33.8 psu and nitrate and silicate concentrations are quite high (around 27.5 and 56–57 mmol/m3, respectively).
* In the Imin cluster, the average sea-surface temperature for the overlying water is around -1.6°C, and sea-ice presence is almost at 100%. In the transport data, SST is around -1.7°C, and sea ice presence is 100%. There are more striking differences between overlying and transport data in salinity, nitrate, and silicate concentrations. For salinity, the overlying data shows an average of around 32.3 psu with a quite large range, whereas the transported data shows a much more constrained value around 33.9 psu. Nitrate and silicate concentrations in the overlying data show values around 25 and 58 mmol/m3, respectively. The transported data show values around 28 and 63 mmol/m3, respectively.
* In the high-Brig cluster, overlying and transport data depict similar values. Sea-surface temperature for both datasets is around 10°C, sea ice is not present (only as outliers), and salinity is around 34.1 psu. Nitrate and silicate concentrations are relatively high (around 16 mmol/m3). Compared to other clusters, the range is relatively large.
* In the Nlab cluster, the average sea-surface temperature for both overlying and transported datasets is around 11–12°C and sea ice is not present, except in two samples. Salinity is around 34.4 psu. Nitrate concentrations are around 15 mmol/m3 and silicate concentrations are around 12 mmol/m3.
* In the low-Brig cluster, the average sea-surface temperature for both datasets is around 15°C. Sea ice is not present, and salinity is around 34.5 psu. Nitrate and silicate concentrations are rather low (just above 10 mmol/m3and below 5 mmol/m3, respectively).
* In the low-Ocen cluster, overlying and transport data depict similar values. The average sea-surface temperature for both data is around 18°C, sea ice is not present, and salinity is around 35.4 psu. Nitrate and silicate concentrations are very low (< 5 mmol/m3).
* For the Iacu cluster, overlying and transport data are again almost identical. Sea-surface temperature for both datasets is around 22.5°C, and sea ice is not present. Salinity is around 35.5 psu and nitrate and silicate concentrations are very low (<2 mmol/ mmol/m3). The range in this cluster is very small.
* In the high-Ocen cluster, the average sea-surface temperature for both data is around 22°C. Sea ice is not present, and salinity is around 35.5 psu. Nitrate and silicate concentrations are very low (<2 mmol/m3). The range in this cluster is relatively narrow for all parameters.
* In the Spin cluster, the average sea-surface temperature for both data is around 29°C. Sea ice is not present, and salinity is just below 34 psu. Nitrate and silicate concentrations are exceptionally low (below 0.1 and 1 mmol/m3, respectively). The range in this cluster is very narrow for all parameters.

Figure 8: Comparison of environmental parameters for the 9-cluster solution in the SH\_655 dataset. For each cluster (Fig. 5), we compare the environmental parameters SST (red), sea ice presence (blue), salinity (purple), nitrate (yellow) and silicate (green) of the directly overlying surface waters (left-side bar, darker fill) to those resulting from the particle transport (right-side bar, lighter fill). Black line, box and whiskers as in Fig. 6.

# 5. Discussion

## 5.1 Dynamics at ice-proximal sites: dinocysts as sea-ice proxy

The better representation of Antarctic ice-proximal sites in our new dataset allows us to further examine Antarctic dinocyst assemblages and associated environmental dynamics, which will help constrain paleoreconstructions. The dominance of *Islandinium minutum* in Weddell Sea locations is very striking (Figs. 3, 4). In the wsi\_100 dataset, nine samples with high *I. minutum* percentages form their own cluster (Fig. 4). The cluster persists when the entire “sh\_655” dataset is considered (Fig. 5). This suggests that regionally unique environmental conditions strongly favor the presence of *I. minutum*. The source region for these samples can be placed in the Weddell Sea, suggesting that *I minutum* may be transported from the Weddell Sea towards the tip of the Antarctic Peninsula along with the icebergs (“Iceberg Alley”; Fig. 7). The low salinities of the surface waters overlying Imin cluster samples further suggests a major influence of icebergs and associated melting as the values are the lowest of all clusters (Fig. 6; also, in the sh\_655 dataset, Figs. 8 & 9). It seems that *I. minutum* dominates the assemblage in low and/or seasonally variable salinity and can thus thrive under these conditions, outcompeting other species. Other conditions such as SST, sea-ice presence, and nutrient concentrations, although slightly lower than in the other clusters, should still be high enough to promote blooms of primary productivity. As icebergs and mixing deliver enough micronutrients, there is no (local) iron-limitation expected. Hence, we suggest that a high abundance/dominance of *I. minutum* represents strongly changing and/or relatively low salinity due to the influence of melt water in conditions otherwise typical for ice-proximal sites. Low salinity signals might also appear very locally restricted/punctuated in regions of high sea-ice presence, thus explaining the sparse occurrences of *I. minutum* in other clusters (Fig. 4).

Apart from this, there are no clear regional differences in the dinocyst abundance (Fig. 4) or clustering of the wsi\_100 dataset (Fig. 6). Merging of both *Selenopemphix antarctica* varieties in the wsi\_100 dataset limits a better delineation of the clusters (Fig. 4). Two clusters are dominated by (varying contributions of) *S. antarctica* (Clusters 1 & 5). Taking our newly counted samples into account in which we separate *S. antarctica* into two varieties (Fig. 3), Cluster 5 mostly consists of *Selenopemphix* sp. 1. Also, *S. antarctica* dominates an assemblage alone, whereas *Selenopemphix s*p. 1 occurs with *Nucicla umbiliphora* (Figs. 3 & 4). Yet, no clear geographic nor environmental component can explain this difference (Figs. 3, 4 & 6). If at all, one could argue that *Selenopemphix* sp. 1 – *N. umbiliphora* occur closer to the coast in the Ross Sea (Fig. 4). This might indicate that *S. antarctica* is more closely associated to polynya conditions. Yet, there is no polynya vs. closer-to-coast gradient visible in the environmental parameters (Fig. 6). Hence, it remains difficult to attribute environmental conditions to the separate clustering. A better distinction of *S. antarctica* varieties might help to further constrain the differences, although the distinction of *S. antarctica* into subspecies itself remains uncertain.

Despite this, a continuous sea-ice presence in these two clusters argues for a high sea-ice affinity of *S. antarctica* as well as *N. umbiliphora*. This suggests that the species live either directly at the sea-ice edge or in polynyas dependent on sea ice-driven conditions. Here, a short growth season in which micro- and macronutrients become available in an essentially unlimited amount (Swalethorp et al., 2019) fuels phytoplankton growth allowing these heterotrophic dinoflagellate species to thrive.

Clusters 3 & 4 include locations furthest away from the Antarctic continent and display a dominance of *Brigantedinium* spp. and *Impagidinium pallidum*, but with lower numbers of *S. antarctica* (and *N. umbiliphora*) (Fig. 4). For Cluster 3, larger ranges for SST and sea-ice presence (Fig. 6) can be translated to seasonal (winter) sea-ice presence and (relatively longer) ice-free summers. We interpret this as a region where the sea-ice season is no longer the sole oceanographic factor influencing productivity. The geographic range and high nutrient concentrations (nitrate and by inference silicate) indicate that these locations are influenced by strong vertical mixing delivering (micro- and) macro-nutrients to the surface ocean.

This opens the question as to when low numbers of *S. antarctica* (or *N. umbiliphora*) should be considered as a signal of low sea-ice presence or as a transport signal (Nooteboom et al., 2019) in paleo-reconstructions. Again, we argue for considering the entire dinocyst assemblage. If the assemblage is dominated by species such as *Brigantedinium* spp., *Impagidinium pallidum*, or *Operculodinium centrocarpum*, with broad environmental niches and occurrences, we suggest considering *S. antarctica* and *N. umbiliphora* as indicators of low (seasonal) sea-ice presence. In contrast, if *S. antarctica* and *N. umbiliphora* exist in assemblages dominated by *N. labyrinthus, I. aculeatum or Spiniferites* spp., we argue for a transport signal, since in our clusters, *S. antarctica* and *N. umbiliphora* are completely absent when sea-ice is absent (Figs. 5, 9 and 10). Note that this transport includes that of deep-ocean and bottom currents which are strong in the Southern Ocean subsurface (see Nooteboom et al., 2019). Consequently, the (low) occurrences of *Brigantedinium* spp. in *S. antarctica*-dominated clusters may indicate the high tolerance of that species to sea ice and/or quick growth during (short) ice-free conditions.

## 5.2 SH\_655 dataset: representation of Southern Hemisphere frontal zones

Compared to previous studies (e.g., Prebble et al., 2013), our study has additional surface sediment samples on both extreme ends: the warm low-latitude end as well as the cold, sea ice-influenced, polar end. As a result, our new sediment sample set better represents the full suite of Southern Hemisphere surface oceanographic conditions. From the pole towards the equator, the clusters depict gradual warming, and decreases in nitrate and silicate concentrations (Fig. 9), as seen in the modern oceanographic conditions (Fig. 1). The sea ice-influenced zone is represented by samples from the summer and winter sea-ice edge, from regions affected by polynyas (e.g., Ross Sea and Weddell Sea) and icebergs (Antarctic Peninsula). On the warm end, the Spin cluster is a representation of warm, low-salinity but also surprisingly low-nutrient surface waters, possibly impacted by high precipitation rates (Fig. 9).

The depicted gradients in the Southern Hemisphere oceanographic conditions can be assigned to key zones in the Southern Hemisphere surface ocean, with specific oceanographic parameters (Figs. 1, 9), clearly represented by the dinocyst assemblage clusters (Fig. 5). It must be noted here that, although these clusters are characterized by one or two dominant taxa, assigning ecological affinity from assemblages should consider the full context of the dinocyst assemblage(s).

* Antarctic Zone: Sant- and Imin clusters. The Antarctic Zone is clearly characterized by an abundance of P-cysts, with dominance of *Selenopemphix antarctica* and/or *Islandinium minutum*. These are the only two clusters with significant sea-ice presence and SST below 0°C. Together with high nutrient conditions, these two clusters clearly depict the AZ conditions of upwelling, vertical mixing, and sea-ice influence. Potentially, the high nutrient concentrations can fuel phytoplankton growth that the dominant dinocyst species can feed on then. However, we cannot confirm if *S. antarctica* and/or *I. minutum* thrive under/at the edge of sea-ice conditions and could thus be used as a sea-ice proxy. The cluster also includes samples further away from the Antarctic continent (Fig. 5b), where the influence of sea ice might be limited, but cold SST and high nutrient conditions prevail. Thus, a quantitative estimate for past sea ice seems difficult. On the other hand, sea-ice dominated regions like the Ross Sea, Prydz Bay, and the Sabrina Coast contain samples with high abundances of *S. antarctica*, suggesting that this species might be at least best adapted to sea-ice conditions. The largest difference between these two clusters can be found in their salinity range (Fig. 9). Regions within the sea-ice zone dominated by *I. minutum* have substantially lower salinity than those dominated by *S. antarctica* and as such, dominance of *I. minutum* might be taken as a low-salinity indicator in cold, sea ice-prone environments (see section 5.1).
* Subantarctic zone: Nlab cluster. The Subantarctic Zone is very well depicted by the Nlab cluster, dominated by *N. labyrinthus*. Besides that species, *Brigantedinium* spp., and other P- or G-cysts (*Impagidinium* species) might be present in small numbers. The environmental parameters of this cluster are characteristic of the SAZ (Fig. 9). Although the environmental parameter profile and occurrence of many dinocyst species might indicate a transitional character of this zone, the SST range compared to other distinctly zonal clusters is quite narrow. Similarly, the STF can be very clearly identified as the northern boundary for this cluster with high numbers of *N. labyrinthus* occurrences (Fig. 5b), best seen around Australia/Tasmania and New Zealand.
* North of the Subtropical Front: Iacu- and Ocen cluster. The clusters that describe best the conditions just north of the STF are the Iacu cluster with *I. aculeatum* and the Ocen clusters with *O. centrocarpum* being the dominant species. The trends of increased SST and salinity due to higher evaporation continue, and nutrient availability drops to almost zero, characteristic for sub-tropical conditions that are less influenced by Southern Ocean/ACC-conditions. The environmental difference between these two clusters is difficult to discern. It seems that samples of the Ocen cluster have slightly higher nitrate (and silicate) concentrations and lower salinity (Fig. 9). For the high-Ocen cluster, this might be caused by samples of the cluster being located closer to land (Fig. 5b). This might argue for *I. aculeatum* needing slightly more open-ocean conditions. Considering the much narrower environmental parameter ranges in the Iacu cluster, this might indicate that *I. aculeatum* can dominate the assemblage in a quite specific range but is outcompeted by the generalist *O. centrocarpum* if conditions are not optimal. Nevertheless, the drastic assemblage changes from the *N. labyrinthus*-dominance south of the STF to the *I. aculeatum*- or O. centrocarpum dominance north of the STF might be used to track STF positional shifts in the past. While many clusters (due to the doubled sample size, see Table 1) differ significantly from clusters identified by Prebble et al., (2013), this Nlab-vs-Iacu-distinction at the STF was also observed there.
* Warm low latitudes: Spin cluster. The cluster dominated by *Spiniferites* spp. appears in very low latitudes and/or locally very warm areas, such as the Australian west coast (Fig. 5), confirmed by almost 30°C SST. Striking is also the lower salinity of this cluster, compared to the others.

Figure 9: Comparison of environmental surface conditions in different clusters for the 9-cluster solution of the sh\_655 dataset. The median, 25% – 75% quantiles and 95% confidence interval is indicated by the black line, boxes and whiskers, respectively, for environmental parameters SST (°C), sea ice presence (%), salinity (psu), nitrate and silicate (mmol/m3) for each cluster color-coded as in Fig. 5. The environmental conditions of the directly overlying water are used here, as particle transport does not cause large differences in the environmental ranges of the clusters (see Fig. 8).

The 9 clusters are mostly clearly restricted to specific zones and regions in the Southern Hemisphere. However, there seems to be a gap remaining in the 0–8°C SST range, that is partly covered by either the polar clusters and/or the high-Brig. This SST range also includes the transitional zone between 0 and 100% sea-ice presence. Better geographical representation of this transitional zone in surface sediment sample sets might improve constraint of this transitional zone. On the other hand, it might be that the nature of this part of the Southern Ocean, with large scale turbulent mixing causes a mixed dinocyst assemblage. The large ranges of the high-Brig in SST, but also in nutrient concentration (Fig. 9) might best represent this dynamic environment, with *Brigantedinium* spp. as more generalist species dominating the dinocyst assemblage (Fig. 5). This is also the case for the low-Brig, that does not show any clear oceanographic boundaries. Potentially, conditions that (temporarily) disadvantage the dominance of the expected dinocyst might allow for *Brigantedinium* spp. to take over and thrive. Yet, the difference between the low- and high-Brig in terms of SST, but also nutrient conditions, might argue for a high *Brigantedinium-*tolerance to cold and very dynamic Polar Frontal Zone conditions (Figs. 4, 9).

## 5.3 The impact of lateral transport

The size of the source regions represented in the sample stations, as visualized in Fig. 7, suggests that many surface sediment samples in the Southern Ocean are impacted by transport (see also Nooteboom et al., 2019). This is especially true for samples located within the ACC. For these locations, their source regions are consistently placed to the west, implying that the strong ACC current transports particles clockwise, eastward around Antarctica. South of the direct influence of the ACC, the source region is displaced farther towards the east due to the counter-clockwise Antarctic Coastal Current. Other features that are recognizable are the Agulhas Leakage, transporting particles along the east coast of South Africa into the Atlantic Ocean, and the so-called ‘Iceberg Alley’. This area is known for a high number of icebergs drifting along the west coast and eventually out of the Weddell Sea due to the Weddell Gyre. Locations that are not directly influenced by the ACC and these smaller features do not experience such a large transport. Thus, ice-proximal and low-latitude (north of ~40°S) sites experience relatively little transport. This can mostly be attributed to the absence of strong currents and coastal proximity.

Overall, zonal transport is much stronger than meridional. This does not exclude latitudinal transport (Nooteboom et al., 2019), but does limit its influence. Environmental conditions are not hugely different on a zonal scale as they are meridional, which suggests that cysts preserved in sediments can be generally linked to overlaying surface water conditions. The comparison of environmental parameters between transported particles versus those from the directly overlying water for the different clusters confirms this assumption (Fig. 8). Transport does not affect the environmental affinities of the clusters significantly. The only exception is the Imin, that clearly shows the influence of transport along the “Iceberg Alley”. The source region of the particles is predominantly the Weddell Sea, hence the cluster represents narrower environmental conditions (Fig. 8).

## 5.4 Dinocyst assemblages as proxy for Southern Ocean frontal systems

For the regions in the Southern Ocean that were already well-represented in previous surface sediment compilations (e.g., Prebble et al., 2013), our additional samples do not require changes in earlier interpretations. Distinct dinocyst assemblages exist on either side of the front systems in the Southern Ocean (Fig. 10). For the Subtropical Front, the difference is mostly observed between the Nlab in the SAZ and Ocen-or Iacu north of the STF. These clusters are promising as proxies for depicting the paleolocation of the STF relative to the position of sediment records: high abundance of *Nematosphaeropsis labyrinthus* means the STF was north of the study area, whereas high abundance of *Operculodinium centrocarpum* or *Impagidinium aculeatum* indicates that the STF was south of the study area. The few samples available between the SAZ and the PF generates an unclear signal, although it seems like the assemblages transition from abundant Nlab to high-Brig. The latter is associated with the high-nutrient conditions in the Antarctic Divergence.

Figure 10: Schematic representation of the generalized biogeographic distribution of dinocysts in Southern Ocean surface sediments. Pies represent average assemblage composition of the nine clusters described in this paper. Position of these pies represent their typical latitudinal band of occurrence. Also plotted are the frontal systems (blue lines, STF = Subtropical Front, SAF = Subantarctic Front, PF = Polar Front). The Subantarctic Zone (SAZ) is the water mass between the STF and PF. Map was produced using ggplot2 in R (Wickham, 2016).

# 6. Conclusion

This study fills a gap in the knowledge of dinocyst assemblage environmental preferences, specifically from the Antarctic ice-proximal locations. With a more solid framework to qualitatively interpret past oceanographic conditions (sea ice, nutrients, sea-surface temperature, salinity) from Southern Ocean dinocyst assemblages (paleo)reconstructions can now be tightly constrained. The 5 clusters in the ice-proximal assemblages demonstrate regional heterogeneity in sea-ice ecosystems. However, *Selenopemphix* *antarctica* can be used as sea-ice indicator and assemblages dominated by a combination of *Selenopemphix* sp.1 and *N. umbiliphora* may also provide information on the length of sea-ice season. *Selenopemphix* *antarctica* and *Selenopemphix* sp.1 need to be counted separately before further environmental interpretations and conclusions can be made.

We interpreted nine clusters in the entire Southern Hemisphere dinocyst database (n=665), which can be broadly related to various oceanographic zones in the Southern Ocean and have clear differences in their oceanographic affinities. Despite strong currents and deep basins, lateral transport of sinking particles has little effect on the relationship between surface oceanographic conditions and sedimentary assemblages. This means that for the clusters, oceanographic conditions of the directly overlying water are a good approximation of what sedimentary plankton assemblages relate to, while lateral transport can influence sedimentary plankton assemblages at individual sites or when the results of this study are used in paleoreconstructions. From the results of the entire Southern Ocean dataset and our identified 9 clusters, we identify frontal-zone-specific dinocyst assemblages, separated by Southern Ocean fronts PF, SAF, STF. Our results provide a solid basis for further use of dinocyst assemblages as indicator for past oceanographic conditions in the Southern Ocean, including frontal system locations through time.

# 7. Data availability

Microscope slides are stored in the collection of the Marine Palynology and Paleoceanography group under storage code 000.000.017.533. Dinocyst data (includes the counts of *Polarella glacialis*, sample weights and *Lycopodium* counts to calculate absolute concentrations), particle trace model output and various cluster analyses, as well as an R markdown providing the coding for the plots in this paper, are available at Github (Bijl, 2022; <https://github.com/bijlpeter83/SH655/>) and deposited at Zenodo (DOI:10.5281/zenodo.6786422).

# 8. Author contributions

PKB designed the research with FS. SH, RW, SN and EM provided new sediment samples. LT and SH analysed the samples. PN contributed Lagrangian particle trace simulations and interpretation of the model data with IS. All authors collaborated in analysing the results and provided input for the writing of the paper.

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# References

Adkins, J. F.: The role of deep ocean circulation in setting glacial climates, Paleoceanography, 28, 539–561, doi.org/10.1002/palo.20046, 2013.

Armand, L.K., O’Brien, P.E. and On-board Scientific Party. Interactions of the Totten Glacier with the Southern Ocean through multiple glacial cycles (IN2017-V01): Post-survey report, Research School of Earth Sciences, Australian National University: Canberra, 2018 http://dx.doi.org/10.4225/13/5acea64c48693

Behrens, B., Miyairi, Y., Sproson, A. D., Yamane, M., and Yokoyama, Y.: Meltwater discharge during the Holocene from the Wilkes subglacial basin revealed by beryllium isotope analysis of marine sediments, Journal of Quaternary Science, 34, 603-608, <https://doi.org/10.1002/jqs.3148,> 2019.

Bijl, P.K.: SH655 dinocyst surface sediment database, <https://github.com/bijlpeter83/SH655>; last access 01-07-2022; DOI:10.5281/zenodo.6786422.

Bijl, P. K., Houben, A. J., Hartman, J. D., Pross, J., Salabarnada, A., Escutia, C., & Sangiorgi, F. (2018a). Paleoceanography and ice sheet variability offshore Wilkes Land, Antarctica-Part 2: Insights from Oligocene-Miocene dinoflagellate cyst assemblages. *Climate of the Past, 14*(7), 1015–1033.

Bijl, P. K., Houben, A. J. P., Bruls, A., Pross, J., and Sangiorgi, F.: Stratigraphic calibration of Oligocene–Miocene organic-walled dinoflagellate cysts from offshore Wilkes Land, East Antarctica, and a zonation proposal, J Micropalaeontol, 37, 105–138, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.5194/jm-37-105-2018, 2018b.

Bromwich, D. H., Carrasco, J. F., and Stearns, C. R.: Satellite Observations of Katabatic-Wind Propagation for Great Distances across the Ross Ice Shelf, Mon Weather Rev, 120, 1940–1949, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1175/1520-0493(1992)120<1940:sookwp>2.0.co;2, 1992.

Crosta, X., Etourneau, J., Orme, L. C., Dalaiden, Q., Campagne, P., Swingedouw, D., Goosse, H., Massé, G., Miettinen, A., McKay, R. M., Dunbar, R. B., Escutia, C., and Ikehara, M.: Multi-decadal trends in Antarctic sea-ice extent driven by ENSO–SAM over the last 2,000 years, Nat Geosci, 14, 156–160, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1038/s41561-021-00697-1, 2021.

Delandmeter, P and E van Sebille (2019), The Parcels v2.0 Lagrangian framework: new field interpolation schemes, *Geoscientific Model Development*, *12*, 3571–3584

De Schepper, S., Fischer, E.I., Groeneveld, J., Head, M.J., Matthiessen, J.: Deciphering the palaeoecology of Late Pliocene and Early Pleistocene dinoflagellate cysts, Palaeogeography, Palaeoclimatology, Palaeoecology, 309 (1–2), pp. 17–32, DOI: 10.1016/j.palaeo.2011.04.020, 2011.

de Vernal A., Henry M., Matthiessen J., Mudie P.J., Rochon A., Boessenkool K.P., Eynaud F., Grøsfjeld K., Guiot J., Hamel D., Harland R., Head M.J., Kunz-Pirrung M., Levac E., Loucheur V., Peyron O., Pospelova V., Radi T., Turon J.-L., Voronina E.: Dinoflagellate cyst assemblages as tracers of sea-surface conditions in the Northern North Atlantic, Arctic and sub-Arctic seas: The new “n = 67” data base and its application for quantitative palaeoceanographic reconstruction, Journal of Quaternary Science, 16 (7), pp. 681 – 698, 2001.

de Vernal, A. de, Eynaud, F., Henry, M., Hillaire-Marcel, C., Londeix, L., Mangin, S., Matthiessen, J., Marret, F., Radi, T., Rochon, A., Solignac, S., and Turon, J.-L.: Reconstruction of sea-surface conditions at middle to high latitudes of the Northern Hemisphere during the Last Glacial Maximum (LGM) based on dinoflagellate cyst assemblages, Quaternary Sci Rev, 24, 897–924, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/j.quascirev.2004.06.014, 2005.

de Vernal, A., de, Radi, T., Zaragosi, S., Van Nieuwenhove, N., Rochon, A., Allan, E., De Schepper, S., Eynaud, F., Head, M.J., Limoges, A., Londeix, L., Marret, F., Matthiessen, J., Penaud, A., Pospelova, V., Price, A., Richerol, T: Distribution of common modern dinoflagellate cyst taxa in surface sediments of the Northern Hemisphere in relation to environmental parameters: The new n=1968 database, Marine Micropaleontology, 159, art. no. 101796. DOI: 10.1016/j.marmicro.2019.101796, 2020.

Esper, O. and Zonneveld, K. A. F.: The potential of organic-walled dinoflagellate cysts for the reconstruction of past sea-surface conditions in the Southern Ocean, Mar Micropaleontol, 65, 185–212, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/j.marmicro.2007.07.002, 2007.

Evangelinos, D., Escutia, C., van de Flierdt, T., Valero, L., Flores, J. A., Harwood, D. M., Hoem, F. S., Bijl, P., Etourneau, J., Kreissig, K., Nilsson-Kerr, K., Holder, L., López-Quirós, A., and Salabarnada, A.: Absence of a strong, deep-reaching Antarctic Circumpolar Current zonal flow across the Tasmanian gateway during the Oligocene to early Miocene, Global and Planetary Change, 208, 10.1016/j.gloplacha.2021.103718, 2022.

Hahsler, M., Piekenbrock, M., and Doran, D.: dbscan: Fast Density-Based Clustering with R., J Stat Softw, 1, 1–30, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.18637/jss.v091.i01, 2019.

Harland, R. and Pudsey, C. J.: Dinoflagellate cysts from sediment traps deployed in the Bellingshausen, Weddell and Scotia seas, Antarctica, Mar Micropaleontol, 37, 77–99, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/s0377-8398(99)00016-x, 1999.

Hartman, J. D., Bijl, P. K., and Sangiorgi, F.: A review of the ecological affinities of marine organic microfossils from a Holocene record offshore of Adélie Land (East Antarctica), J Micropalaeontol, 37, 445–497, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.5194/jm-37-445-2018, 2018.

Hoem, F.S., Valero, L., Evangelinos, D., Escutia, C., Duncan, B., McKay, R.M., Brinkhuis, H., Sangiorgi, F., Bijl, P.K.: Temperate Oligocene surface ocean conditions offshore of Cape Adare, Ross Sea, Antarctica, Climate of the Past, 17 (4), pp. 1423–1442, DOI: 10.5194/cp-17-1423-2021, 2021a.

Hoem, F.S., Sauermilch, I., Hou, S., Brinkhuis, H., Sangiorgi, F., Bijl, P.K.: Late Eocene-early Miocene evolution of the southern Australian subtropical front: a marine palynological approach, Journal of Micropalaeontology, 40 (2), pp. 175–193, DOI: 10.5194/jm-40-175-2021, 2021b.

Houben, A. J. P., Bijl, P. K., Pross, J., Bohaty, S. M., Passchier, S., Stickley, C. E., Röhl, U., Sugisaki, S., Tauxe, L., Flierdt, T. van de, Olney, M., Sangiorgi, F., Sluijs, A., Escutia, C., Brinkhuis, H., Scientists, E. 318, Dotti, C. E., Klaus, A., Fehr, A., Williams, T., Bendle, J. A. P., Carr, S. A., Dunbar, R. B., Flores, J.-A., Gonzàlez, J. J., Hayden, T. G., Iwai, M., Jimenez-Espejo, F. J., Katsuki, K., Kong, G. S., McKay, R. M., Nakai, M., Pekar, S. F., Riesselman, C., Sakai, T., Salzmann, U., Shrivastava, P. K., Tuo, S., Welsh, K., and Yamane, M.: Reorganization of Southern Ocean Plankton Ecosystem at the Onset of Antarctic Glaciation, Science, 340, 341–344, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1126/science.1223646, 2013.

Lange, M and E van Sebille (2017), Parcels v0.9: prototyping a Lagrangian Ocean Analysis framework for the petascale age, *Geoscientific Model Development*, *10*, 4175–4186

Locarnini, A., R., Mishonov, A. V., Baranova, O. K., Boyer, T. P., Zweng, M. M., Garcia, H. E., Reagan, J. R., Seidov, D., Weathers, K., Paver, C. R., and Smolyar, I.: WORLD OCEAN ATLAS 2018 Volume 1: Temperature, 2018.

Madec, G.: NEMO ocean engine, 2016.

Marret, F., Vernal, A. de, Benderra, F., and Harland, R.: Late Quaternary sea‐surface conditions at DSDP Hole 594 in the southwest Pacific Ocean based on dinoflagellate cyst assemblages, J Quaternary Sci, 16, 739–751, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1002/jqs.648, 2001.

Marret, F., Zonneveld, K.A.F.: Atlas of modern organic-walled dinoflagellate cyst distribution. Review of Palaeobotany and Palynology 125(1–2), 1–200, DOI: 10.1016/S0034-6667(02)00229-4, 2003.

Marret, F., Bradley, L., Vernal, A. de, Hardy, W., Kim, S.-Y., Mudie, P., Penaud, A., Pospelova, V., Price, A. M., Radi, T., and Rochon, A.: From bi-polar to regional distribution of modern dinoflagellate cysts, an overview of their biogeography, Mar Micropaleontol, 101753, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/j.marmicro.2019.101753, 2020.

Marschalek J.W., Zurli L., Talarico F., van de Flierdt T., Vermeesch P., Carter A., Beny F., Bout-Roumazeilles V., Sangiorgi F., Hemming S.R., Pérez L.F., Colleoni F., Prebble J.G., van Peer T.E., Perotti M., Shevenell A.E., Browne I., Kulhanek D.K., Levy R., Harwood D., Sullivan N.B., Meyers S.R., Griffith E.M., Hillenbrand C.-D., Gasson E., Siegert M.J., Keisling B., Licht K.J., Kuhn G., Dodd J.P., Boshuis C., De Santis L., McKay R.M., Ash J., Browne I.M., Cortese G., Dodd J.P., Esper O.M., Gales J.A., Harwood D.M., Ishino S., Kim S., Kim S., Laberg J.S., Leckie R.M., Müller J., Patterson M.O., Romans B.W., Romero O.E., Seki O., Singh S.M., Cordeiro de Sousa I.M., Sugisaki S.T., Xiao W., Xiong Z.: A large West Antarctic Ice Sheet explains early Neogene sea-level amplitude, Nature 600, 7889, pp 450–455, 2021.

Marshall, J. and Speer, K.: Closure of the meridional overturning circulation through Southern Ocean upwelling, Nat Geosci, 5, 171, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1038/ngeo1391, 2012.

Mitchell, B. G., Brody, E. A., Holm‐Hansen, O., McClain, C., and Bishop, J.: Light limitation of phytoplankton biomass and macronutrient utilization in the Southern Ocean, Limnol Oceanogr, 36, 1662–1677, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.4319/lo.1991.36.8.1662, 1991.

Mudie, P. J., Marret, F., Mertens, K. N., Shumilovskikh, L., and Leroy, S. A. G.: Atlas of modern dinoflagellate cyst distributions in the Black Sea Corridor: from Aegean to Aral Seas, including Marmara, Black, Azov and Caspian Seas, Mar Micropaleontol, 134, 1–152, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/j.marmicro.2017.05.004, 2017.

Nooteboom, P. D., Bijl, P. K., Sebille, E., Heydt, A. S., and Dijkstra, H. A.: Transport Bias by Ocean Currents in Sedimentary Microplankton Assemblages: Implications for Paleoceanographic Reconstructions, Paleoceanogr Paleoclimatology, 34, 1178–1194, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1029/2019pa003606, 2019.

Nooteboom, P. D., Delandmeter, P., Sebille, E. van, Bijl, P. K., Dijkstra, H. A., and Heydt, A. S. von der: Resolution dependency of sinking Lagrangian particles in ocean general circulation models, Plos One, 15, e0238650, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1371/journal.pone.0238650, 2020.

Nooteboom, P.D., Bijl, P.K., Kehl, C., Van Sebille, E., Ziegler, M., Von Der Heydt, A.S., Dijkstra, H.A.: Sedimentary microplankton distributions are shaped by oceanographically connected areas, Earth System Dynamics, 13 (1), pp. 357–371, DOI: 10.5194/esd-13-357-2022, 2022.

Orsi, A. H., Whitworth, T., and Nowlin, W. D.: On the meridional extent and fronts of the Antarctic Circumpolar Current, Deep Sea Res Part Oceanogr Res Pap, 42, 641–673, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/0967-0637(95)00021-w, 1995.

Park, Y. H., Park, T., Kim, T. W., Lee, S. H., Hong, C. S., Lee, J. H., Rio, M. H., Pujol, M. I., Ballarotta, M., Durand, I., and Provost, C.: Observations of the Antarctic Circumpolar Current Over the Udintsev Fracture Zone, the Narrowest Choke Point in the Southern Ocean, Journal of Geophysical Research: Oceans, 124, 4511-4528, <https://doi.org/10.1029/2019JC015024,> 2019.

Prebble, J.G., Crouch, E.M., Cortese, G., Carter, L., Neil, H., Bostock, H.: Southwest Pacific sea surface conditions during Marine Isotope Stage 11–- Results from dinoflagellate cysts, Palaeogeography, Palaeoclimatology, Palaeoecology, 446, pp. 19–31, DOI: 10.1016/j.palaeo.2016.01.007, 2016.

Prebble, J. G., Crouch, E. M., Carter, L., Cortese, G., Bostock, H., and Neil, H.: An expanded modern dinoflagellate cyst dataset for the Southwest Pacific and Southern Hemisphere with environmental associations, Mar Micropaleontol, 101, 33–48, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/j.marmicro.2013.04.004, 2013.

Qin, X., Sebille, E. van, and Gupta, A. S.: Quantification of errors induced by temporal resolution on Lagrangian particles in an eddy-resolving model, Ocean Model, 76, 20–30, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1016/j.ocemod.2014.02.002, 2014.

Rhodes, R. H., Bertler, N. A. N., Baker, J. A., Sneed, S. B., Oerter, H., and Arrigo, K. R.: Sea ice variability and primary productivity in the Ross Sea, Antarctica, from methylsulphonate snow record, Geophys Res Lett, 36, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.1029/2009gl037311, 2009.

Sangiorgi, F., Bijl, P. K., Passchier, S., Salzmann, U., Schouten, S., McKay, R., Bohaty, S. M.: Southern Ocean warming and Wilkes Land ice sheet retreat during the mid-Miocene, *Nature communications, 9*(1), 317, 2018.

Solodoch, A., Stewart, A. L., Hogg, A. M., Morrison, A. K., Kiss, A. E., Thompson, A. F., Purkey, S. G., and Cimoli, L.: How Does Antarctic Bottom Water Cross the Southern Ocean?, Geophysical Research Letters, 49, e2021GL097211, <https://doi.org/10.1029/2021GL097211,> 2022.

Spreen, G., Melsheimer, C., Malte Gerken, M. https://seaice.uni-bremen.de/databrowser/ (last access: 17-12-2022). Institute of Environmental Physics, University of Bremen, Germany, 2015 – 2021

Swalethorp, R., Dinasquet, J., Logares, R., Bertilsson, S., Kjellerup, S., Krabberød, A.K., Moksnes, P.-O., Nielsen, T.G., Riemann, L.: Microzooplankton distribution in the Amundsen Sea Polynya (Antarctica) during an extensive Phaeocystis antarctica bloom, Progress in Oceanography, Volume 170, Pages 1–10, <https://doi.org/10.1016/j.pocean.2018.10.008>, 2019.

Talley, L.: Closure of the Global Overturning Circulation Through the Indian, Pacific, and Southern Oceans: Schematics and Transports, Oceanography, 26, 80–97, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.5670/oceanog.2013.07, 2013.

Wickham H.: ggplot2: Elegant Graphics for Data Analysis, Springer-Verlag New York, ISBN 978-3-319-24277-4, <https://ggplot2.tidyverse.org>, 2016.

Williams, G. L., Fensome, R. A., and MacRae, R. A.: The Lentin and Williams Index of Fossil Dinoflagellates, [https://do](file:///Users/nooteboom/Library/Containers/com.apple.mail/Data/Library/Mail%20Downloads/A8E53289-42C6-47A5-AA2C-7C7D33155482/%20https:/do)i.org/10.4095/103330, 2017.

Wilson, D. J., Bertram, R. A., Needham, E. F., van de Flierdt, T., Welsh, K. J., McKay, R. M., Mazumder, A., Riesselman, C. R., Jimenez-Espejo, F. J., and Escutia, C.: Ice loss from the East Antarctic Ice Sheet during late Pleistocene interglacials, Nature, 561, 383-386, 10.1038/s41586-018-0501-8, 2018.

Yool, A., Popova, E. E., and Anderson, T. R.: MEDUSA-2.0: an intermediate complexity biogeochemical model of the marine carbon cycle for climate change and ocean acidification studies, Geosci Model Dev, 6, 1767–1811, https://doi.org/10.5194/gmd-6-1767-2013, 2013.

Zonneveld, K. A. F., Marret, F., Versteegh, G. J. M., Bogus, K., Bonnet, S., Bouimetarhan, I., Crouch, E., Vernal, A. de, Elshanawany, R., Edwards, L., Esper, O., Forke, S., Grøsfjeld, K., Henry, M., Holzwarth, U., Kielt, J.-F., Kim, S.-Y., Ladouceur, S., Ledu, D., Chen, L., Limoges, A., Londeix, L., Lu, S.-H., Mahmoud, M. S., Marino, G., Matsouka, K., Matthiessen, J., Mildenhal, D. C., Mudie, P., Neil, H. L., Pospelova, V., Qi, Y., Radi, T., Richerol, T., Rochon, A., Sangiorgi, F., Solignac, S., Turon, J.-L., Verleye, T., Wang, Y., Wang, Z., and Young, M.: Atlas of modern dinoflagellate cyst distribution based on 2405 data points, Rev Palaeobot Palyno, 191, 1–197, https://doi.org/10.1016/j.revpalbo.2012.08.003, 2013.