DR. HELEN COLLEY (Orcid ID : 0000-0003-0053-7468)

Article type : Concise Communications

Expression and enzyme activity of Cytochrome P450 enzymes CYP3A4 and CYP3A5 in human skin and tissue engineered skin equivalents

Sarah A. Smith[1], Helen E. Colley[1*], Parveen Sharma[2], Klaudia M. Slowik[1], Rowena Sison-Young[2], Andrew Sneddon[3], Steven D. Webb[3], Craig Murdoch[1]

[1]School of Clinical Dentistry, University of Sheffield, Claremont Crescent, Sheffield, S10 2TA, UK.

[2]MRC Centre for Drug Safety Science, Department of Molecular and Clinical Pharmacology, Sherrington Building, Ashton Street, University of Liverpool, L69 3GE, UK.

[3]Department of Applied Mathematics, Liverpool John Moores University, James Parsons Building, Byrom Street, Liverpool, L3 3AF, UK.

*Corresponding author: Dr Helen Colley, School of Clinical Dentistry, University of Sheffield, Claremont Crescent, Sheffield, S10 2TA, UK. E-mail. h.colley@sheffield.ac.uk; Tel: +44 114 2259352

Abstract

CYP3A4 and CYP4A5 share specificity for a wide range of xenobiotics with the CYP3 subfamily collectively involved in the biotransformation of approximately 30% of all drugs. CYP3A4/5 mRNA transcripts have been reported in the skin yet knowledge of their protein expression and function is lacking. In this study, we observed gene and protein expression of CYP3A4/5 in both human skin and tissue-engineered skin equivalents (TESE), and enzyme activity was detected using the model substrate benzyl-*O*-methyl-cyanocoumarin. Mass spectrometric analysis of TESE lysates following testosterone application revealed a time-dependent increase in metabolite production, confirming the functional expression of these enzymes in skin.

1 Background

Cytochrome P450 (CYP450) enzymes are a superfamily of hemoproteins that metabolize a multitude of endogenous and xenobiotic molecules and are essential to maintain homeostasis.[1, 2] As the most highly expressed xenobiotic enzymes in the liver, they are responsible for the majority of phase I reactions, metabolizing between 70-80% of all drugs.[3, 4] The CYP3 subfamily of the CYP450 enzymes comprising of CYP3A4, CYP3A5, CYP3A7 and CYP3A43 are, collectively involved in the metabolism of over a third of these drugs.[4] CYP3A4, the most abundantly expressed isoform in the liver,[5] has a large active site that enables its interaction with a wide range of structurally diverse compounds and permits metabolism of such molecules and drugs as hormones (testosterone, estrogen), anti-cancer drugs (paclitaxel, tamoxifen), and anti-fungal agents (ketoconazole) amongst others.[4, 6] CYP3A5 shares 85% amino acid sequence homology with CYP3A4 and displays

similar substrate specificity.[7] Gene expression of CYP3A4 and CYP3A5 has been detected in skin and tissue engineered skin equivalents (TESE) by genomic analysis in some studies[8, 9, S1, S2] but not others[S3] and, to date, protein expression has not been detected.[S4, S5] Given the use of skin as a drug delivery route, it is extremely important to determine functional expression of CYP3A4/5. Moreover, the increasing use of TESE as an alternative to animal testing for drug irritation and sensitivity assays has led to their proposed use in drug-induced toxicity assays. However, data on functional xenobiotic enzyme activity in these models is lacking and urgently required.

2 QUESTIONS ADDRESSED

The relative gene and protein expression of CYP3A4 and CYP3A5 in human skin in comparison to liver remains unclear. It is also unknown if the expression is localized to the dermis or epidermis and whether the enzymes are functionally active and are able to metabolise clinically relevant molecules.

3 EXPERIMENTAL DESIGN

See e-supporting information.

4 RESULTS

Gene expression levels of CYP3A4 and CYP3A5 were in much greater abundance in the liver than skin with expression being several hundred-fold greater for CYP3A4 and 9-fold for CYP3A5 respectively (Figure 1A-B). In human liver, gene expression of CYP3A4 was significantly greater ($p<0.001$) than for CYP3A5, whereas, in skin, gene expression of CYP3A5 was significantly greater ($p<0.001$) than that of CYP3A4 (Figure 1A-B). Immunoblot analysis revealed that both enzymes were also expressed at markedly lower levels in skin compared

to liver (Figure. 1C-D), confirming the qPCR data. An immuno-positive band corresponding to CYP3A5 was not detected in skin at exposure times that were saturating for liver samples, however, when probed in isolation, CYP3A5 was detected when the exposure time was extended to increase assay sensitivity (Figure 1E). mRNA transcripts for CYP3A4 and CYP3A5 were detected in TESE. Gene expression levels for CYP3A4 were very low and several hundred-fold lower than for CYP3A5, and similar to native skin, TESE expressed significantly more mRNA transcripts for CYP3A5 than CYP3A4 ($p<0.001$) (Figure 1F), and significantly more CYP3A4 ($p<0.05$) and CYP3A5 ($p<0.01$) was expressed in the epidermis compared to the dermis (Figure 1G-H). Immunohistochemical analysis showed that CYP3A4 was equally distributed throughout the epidermis although absent in the cornified layers of both skin and TESE (Figure S1A-B). CYP3A4 staining was also observed in the majority of stromal fibroblasts and was prominent in the endothelium lining the blood vessels within the connective tissue of native skin (Figure S1A). In contrast, staining was not observed in the fibroblasts populating the collagen hydrogel in the TESE (Figure S1B), supporting the qPCR data that showed very low mRNA expression in the TSE dermis. In native skin, expression of CYP3A5 was largely restricted to the stratum basale (Figure S2C) whereas expression in TESE models was extremely weak throughout the entire epithelium (Figure S1D). The discordant in immunostaining and qPCR data is likely due to low antibody affinity for CYP3A5. Both native skin and TESE revealed no staining in the dermis, suggesting that dermal fibroblasts do not express CYP3A5.

Liver extracts exhibited a maximal reaction velocity (V_{max}) of 6694 µmol/min/mg whilst native human skin extracts exhibited a lower V_{max} of 3215 µmol/min/mg. The V_{max} of TESE was approximately half that of native skin (1788 µmol/min/mg). Assuming that the enzyme

catalytic rate (K_{cat}) for CYP3A for this substrate is the same in liver, skin and TESE, we can use these V_{max} estimates to calculate the relative tissue/extract enzyme expressions: namely, skin:liver=0.48:1; TESE:liver=0.267:1; TESE:skin=0.556:1 (Figure 2A-C).

Testosterone was applied to the stratum corneum or to the culture medium of TESE for 8 and 24 hours to mimic topical or systemic delivery. TESE models were separated into epidermal and dermal components and extracts of these along with the conditioned medium were analysed by mass spectrometry for presence of the CYP3A-generated metabolite, 6β-OH-testosterone. When testosterone was applied topically, 6β-OH-testosterone was detected in the epidermis, dermis and in the culture medium at both 8 and 24 hours. At 8 hours 6β-OH-testosterone levels were similar in the epidermis, dermis and medium whereas after 24 hours the levels of 6β-OH-testosterone found in the epidermis were markedly lower, levels in the dermis were similar, whereas levels in the medium had increased five-fold compared to levels at 8 h ($p<0.001$)(Figure 2D). A markedly different pattern of metabolite production was observed when testosterone was added to mimic systemic delivery. In these models, the presence of 6β-OH-testosterone was not detected in the epidermal or dermal compartments at 8 or 24 hours, whereas after 8 hours, 6β-OH-testosterone was detected in the culture medium and these levels were significantly increased after 24 hours ($p<0.001$) (Figure 2E).

5 CONCLUSIONS

In this study we show both gene and protein expression of predominantly CYP3A5 but also CYP3A4 in native skin and TESE. These data have important clinical implications for dermal drug delivery of existing or new compounds, in particular those with structural components that are likely to be metabolised by CYP3A4/5. They also show that, although expressed at

lower levels than skin, TESE are able to replicate the kinetics of skin metabolism of topically and systemically delivered drugs.

6 FUNDING

This study was funded by a National Centre for the Replacement, Refinement & Reduction of Animals in Research (NC3R) UK CRACK-IT Challenge 20 award.

7 CONFLICTS OF INTEREST

The authors have declared no conflicting interests.

8 AUTHOR CONTRIBUTIONS

SAS, HEC, KMS, RS-Y and PS performed the experiments and analysed the data. SDW and AS analysed the enzyme kinetic data. CM, SAS and HEC designed the research study and wrote the manuscript. CM and SDW supervised the research.

Keywords

Xenobiotic metabolism, Toxicity, Steroids, Three dimensional tissue models, Epithelium

9 REFERENCES

1. Guengerich F P. Common and uncommon cytochrome P450 reactions related to metabolism and chemical toxicity. *Chem Res Toxicol* 2001; 14: 611-650.
2. Palrasu M, Nagini S. Cytochrome P450 structure, function and clinical significance: A review. *Curr Drug Targets* 2017.
3. Guengerich F P. Cytochrome p450 and chemical toxicology. *Chem Res Toxicol* 2008; 21: 70-83.
4. Zanger U M, Schwab M. Cytochrome P450 enzymes in drug metabolism: regulation of gene expression, enzyme activities, and impact of genetic variation. *Pharmacol Ther* 2013; 138: 103-141.
5. Ohtsuki S, Schaefer O, Kawakami H, et al. Simultaneous absolute protein quantification of transporters, cytochromes P450, and UDP-glucuronosyltransferases as a novel approach for the characterization of individual human liver: comparison with mRNA levels and activities. *Drug Metab Dispos* 2012; 40: 83-92.
6. Flockhart D A, Rae J M. Cytochrome P450 3A pharmacogenetics: the road that needs traveled. *Pharmacogenomics J* 2003; 3: 3-5.
7. Williams J A, Ring B J, Cantrell V E, et al. Comparative metabolic capabilities of CYP3A4, CYP3A5, and CYP3A7. *Drug Metab Dispos* 2002; 30: 883-891.
8. Luu-The V, Duche D, Ferraris C, et al. Expression profiles of phases 1 and 2 metabolizing enzymes in human skin and the reconstructed skin models Episkin and full thickness model from Episkin. *J Steroid Biochem Mol Biol* 2009; 116: 178-186.
9. Wiegand C, Hewitt N J, Merk H F, et al. Dermal xenobiotic metabolism: a comparison between native human skin, four in vitro skin test systems and a liver system. *Skin Pharmacol Physiol* 2014; 27: 263-275.

10 LEGENDS FOR ILLUSTRATIONS

Figure 1. Comparison of gene and protein expression levels between human liver, skin and TESE for CYP3A4 and CYP3A5. cDNA from human liver and skin was subjected to qPCR analysis for CYP3A4 (blue) and CYP3A5 (red) and gene expression relative to GAPDH determined for (A) liver and (B) skin. Protein expression in liver and skin was resolved by immunoblotting for (C) CYP3A4 and (D) CYP3A5. (E) CYP3A5 expression in skin following prolonged exposure during infrared detection of immuno-positive bands; β-actin was used as a loading control and immunoblots are representative of 3 independent experiments. (F) Relative gene expression of CYP3A4 (blue) and CYP3A5 (red) in whole EpiDerm-FT400™ TESE

by qPCR analysis. RNA from the epidermis and dermis of EpiDerm-FT400™ TESE was isolated, reverse transcribed and subjected to qPCR analysis for (G) CYP3A4 and (H) CYP3A5. Data are relative expression to GAPDH. Data are mean ± SD for n=3 independent experiments and 3 technical repeats per experiment. * P<0.05, ** P<0.01, ***p<0.001 by ANOVA.

Figure 2. Functional activity of CYP3A in liver, skin and TESE and testosterone metabolism by TESE. Protein extracts were generated from human liver, skin and EpiDerm-FT400™ TESE and subjected to kinetic enzyme analysis using increasing concentrations of the model substrate benzyl-*O*-methyl-cyanocoumarin (BOMCC). A Nelder-Mead algorithm was used to fit the enzyme activity data and all data sets were fit with the assumption that K_m (the binding affinity for the substrate at half the maximum velocity) was the same in each assay (K_m = 10 µM). Maximum velocity (V_{max}) was calculated as specific metabolic activity of CYP3A in µmol/min/mg. Data are from three independent experiments performed in triplicate. Goodness of fit R^2 values are: (A) 0.8855, (B) 0.9779 and (C) 0.9962**.** Testosterone was added either to (C) the surface of the epidermis to mimic topical drug deliver or (D) added to the medium that bathes the dermis to mimic systemic drug delivery. The production of the specific testosterone metabolite 6β-OH-testosterone was measured by mass spectrometry in epidermal and dermal extracts as well as the tissue culture medium after 8 and 24 h. Data are from 2 independent experiments and 3 technical repeats per experiment.

Supplementary Figure 1. Spatial localisation of metabolising enzyme expression in native skin and TESE. Formalin-fixed, paraffin-embedded sections of native human skin and EpiDerm-FT400™ TESE were analysed by immunohistochemistry to determine the spatial

expression of CYP3A4 (A & B) and CYP3A5 (C & D). IgG was used as a control (E & F). Scale bar =100 µm. Images are representative of 3 independent experiments from 3 different tissues.