University of Liverpool

Integrated Approach to the Control of Lymphatic Filariasis, Schistosomiasis, and Soil-Transmitted Helminthiasis in Liberia, West Africa

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DECLARATION

I declare that this research is my original work and it has not been presented for the award of a degree at any other university. I wish to acknowledge support staff at the Ministry of Health, Neglected Tropical Diseases Unit and laboratory technicians at the Ministry of Health who participated in the collection of my data during the fieldwork reported here. At my selected schools, principals and schoolteachers were present and they helped me organize and supervise the school children during the surveys.

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Louise Cleopatra Mapleh Kpoto Signed:

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ABSTRACT

In Liberia, the most common neglected tropical diseases (NTDs) are: lymphatic filariasis (LF); schistosomiasis (SCH); soil-transmitted helminthiasis (STH); and onchocerciasis (Oncho). My research sought to explore an integrated approach for the control of these NTDs to assist the Ministry of Health (MoH) in establishing a better co-ordinated, economical and cost-effective intervention. As a result of my research, the MoH, in collaboration with the Centre for Neglected Tropical Diseases (CNTD) at the Liverpool School of Tropical Medicine, progressively unified the disease-specific vertical programmes into a single umbrella, entitled: 'The Integrated NTDs Programme'. Previously, Oncho control was the only tropical disease programme established within the MoH and acted as a stand-alone. Steps towards integration were first tailored upon the existing vertical delivery mechanisms for Oncho i.e., Community-Directed Treatment with Ivermectin (CDTI) through existing drug distribution networks and school-based programmes.

The aim of my study was to develop and implement an integrated strategy for LF, SCH and STH first using the CDTI Oncho networks to obtain data on the prevalence and risk factors for these diseases simultaneously. During my study period, certain activities within the MoH were suspended due to the emergency response to the Ebola epidemic. This eventually led my thesis to be structured into two parts, pre- and postepidemic. **Pre-epidemic** – LF: The first national baseline disease mapping by immunochromatographic test (ICT) cards and baseline microfilaria pre-Mass Drug Administration (MDA) in Liberia. This provided epidemiological data for the entire country, which was used by the MoH to identify implementation units for MDA. The overall ICT prevalence was 24.0% with the highest percentage prevalence observed in the coastal region. The study also revealed that LF was endemic in 13 out of the 15 counties in Liberia. The baseline microfilaria result reported, in the study, a 6.0% prevalence rate. A total of 1,498 men were examined for the clinical manifestation of the disease, of which, 12.0% hydrocele and 6.0% lymphoedema cases were observed. A population-based knowledge and compliance study on LF was carried out in Bong County. Analysis of the results showed that more than 60.0% of the participants were aware of the disease, but only 43.0% of the participants admitted to taking both Albendazole and Ivermectin, demonstrating low treatment coverage, even though 50.0% of the participants knew the mode of LF transmission. Post-epidemic – STH and SCH: Parastiological surveillance studies were undertaken in Bong County as a representative epidemiological indicator for the national control programme to provide up-to-date information on SCH and STH. An epidemiological update on urogenital and intestinal SCH was undertaken amongst schoolchildren in Bong County, northern Liberia alongside observations on STH. A cross-sectional study examined 1,003 school-aged children from 10 schools representing eight health districts in the country where MDA campaigns were ongoing. In total, 12.0% of the children were infected with Schistosoma mansoni and 11.0% infected with Schistosoma haematobium, while general prevalence of STH was much lower with hookworm having the highest prevalence of 3.0%. For the first time, a knowledge attitude and practices survey for SCH, LF and STH was assessed amongst school-aged children in Bong County, northern Liberia, which highlighted the need for better health education and improved sanitation. For example, analysis of data demonstrated that from the 1,003 participants 92.0% had not heard of LF, 86.0% had not heard of STH, and 90.0% had not heard of SCH. Only 9.0% of participants had access to pipe water. To investigate the morbidity associated with SCH, a cross-sectional assessment using portable ultrasound was undertaken, which was the first ever field-based investigation in Liberia. Of the 272 school-aged children examined, morbidity was low (<1.0%) demonstrating little clinical moribidity in this school-based setting.My research was activated and first guided by recommendations from the World Health Organization (Global Plan to Combat NTDs, 2008–2015) to examine ways in which interventions against NTDs could be better co-ordinated and streamlined. The concept of integrated NTD control is attractive in resource poor settings such as Liberia as it can rationalize costs associated with logistics, staffing, and also better delivery of medicines. My research suggests that, the integrated approach to controlling NTDs is possible, cost-effective, and less time consuming. The recommendations made are that, the integrated NTDs programme at the MoH should collaborate with other health service programmes at the MoH, the Ministry of Education, the Liberia water and sewer company, the National Drug Service, and other relevant stakeholders in order to achieve its goal and produce more maintainable impact. The conclusions are that, an integrated disease programme should strengthen and not threaten or compromise the current efforts to efficiently co-ordinate, reduce, eliminate, and ultimately eradicate specific diseases within the NTDs.

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I feel privileged to have had the opportunity to carry out this study as a demonstration of knowledge gained during the period studying for my PhD. It would be impossible not to remember those who in one way or another, directly or indirectly, played a role in the realisation of this research project. Let me therefore thank them all equally. First, I am indebted to the all-powerful GOD for all the blessings He showered me and for being with me throughout my studies. Second, to my supervisors Professor Russell Stothard and Dr Benjamin Koudou – many thanks for your exemplary monitoring and support – this project would not have been a success without your help. My very special thanks and appreciation also goes to the entire Liverpool School of Tropical Medicine, Centre for Neglected Tropical Diseases team. Third, I take this opportunity to express my deep gratitude to my family, and friends who are a constant source of motivation and for their never-ending support and encouragement. Fourth, to the Neglected Tropical Diseases unit of the Ministry of Health headed by Mr Karsor Kollie – I am forever grateful. Finally, to my hard-working research assistants Mrs Lasee Kolee, Mr Stephen M. Gbanyan, Jr. and Mr Mark Zayzay – many thanks.

DEDICATION

This research project is dedicated to my husband Professor Robert M. Kpoto for the unwavering support he rendered me during the course of my study and to all the health care workers who died during the Ebola outbreak in Liberia. To my children Terelouti B. Krangar, Lewis Mapleh, and Rene L. M. Kpoto, I am passing the baton to you all.

THESIS LAYOUT

Chapter 1: Background.

- Chapter 2: Literature review.
- **Chapter 3**: Critical analysis of historical data on the epidemiology and distribution of *W. bancrofti* in vector and human populations in Liberia.
- **Chapter 4**: Baseline mapping for LF by ICT cards and baseline microfilaria pre-MDA in Liberia.
- **Chapter 5**: Assessment of knowledge and compliance towards MDA intervention for the control of LF in Bong County, Liberia: A cross-sectional study.
- **Chapter 6**: An epidemiological survey on urogenital and intestinal SCH amongst school-age children in Bong County, Liberia, with observations on STH.
- Chapter 7: Knowledge, attitudes and practices towards SCH, LF and STH amongst school-age children in Bong County, Liberia.
- **Chapter 8**: Assessment of SCH-related morbidity amongst school-age children in Bong County, Liberia by the use of ultrasound: A cross-sectional study.

Chapter 9: General discussions, recommendations and conclusions

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LIST OF ACRONYMS

| ADLA | Acute dermato-lymphangio-adenitis |
|-------|--|
| AOR | Adjusted odds ratio |
| APOC | African Programme for Onchocerciasis Control |
| BPHS | Basic Package of Health Services |
| CDC | Centers for Disease Control and Prevention |
| CDD | Community drug distributor |
| CDTI | Community-Directed Treatment with Ivermectin |
| CFA | Circulating filarial antigen |
| CI | Confidence interval |
| CNTD | Centre for Neglected Tropical Diseases |
| COR | Crude odds ratio |
| DALY | Disability-adjusted life year |
| DEC | Diethylcarbamazine |
| DNA | Deoxyribonucleic acid |
| ELISA | Enzyme-Linked Immuno-Sorbent Assay |
| EPHS | Essential Package of Health Services |
| EVD | Ebola virus disease |
| FGS | Female Genital Schistosomiasis |
| GDP | Gross domestic product |
| GPELF | Global Programme to Eliminate Lymphatic Filariasis |
| GPS | Global Positioning System |
| HIV | Human immunodeficiency virus |
| ICT | Immunochromatographic test |
| IDSR | Integrated Diseases Surveillance and Response System |
| IHR | International Health Regulation |
| IQR | Interquartile range |
| ITN | Insecticide-treated net |
| IU | Implementation unit |
| KAP | Knowldege, Attitude and Practice |
| LF | Lymphatic filariasis |
| LIBR | Liberian Institute for Biomedical Research |
| LSTM | Liverpool School of Tropical Medicine |
| MDA | Mass Drug Administration |
| MoH | Ministry of Health |
| NCBI | National Center for Biotechnology Information |
| NTD | Neglected tropical disease |
| Oncho | Onchocerciasis |
| PCR | Polymerase chain reaction |
| SCH | Schistosomiasis |
| SD | Standard deviation |
| STD | Sexually transmitted disease |
| STH | Soil-transmitted helminthiasis |
| TAS | Transmission Assessment Survey |
| UNDP | United Nations Development Programme |
| | |

| UNICEF | United Nations Children's Fund |
|--------|---|
| UNMEER | United Nations Mission for Ebola Emergency Response |
| USIN | Unique study identification number |
| WHO | World Health Organization |
| | |

CHAPTER 1: BACKGROUND

1.1 Introduction

Neglected tropical diseases (NTDs) are a varied group of parasitic, fungal, viral and bacterial envenomicing diseases that prevail in tropical and subtropical conditions. NTDs are found in approximately 149 countries globally and affect an estimated 1.4 billion of the world's poorest people. It is referred to as the 'accent afflictions'. NTDs are often associated with significant physical debilitation that results in lower economic productivity and social ostracism (Hotez, Ottesen, Fenwick, & Molyneux, 2006; Hotez, Fenwick, Savioli, & Molyneux, 2009). In recent years, a number of NTDs have been targeted for control or elimination, with some as early as 2020 (WHO, 2012a).

With the help of international partners, some endemic countries are now demonstrating strong ownership and leadership, in variable financial, political and environmental circumstances, to ensure their NTD programmes are successful in meeting 2020 targets, which is inspired by the World Health Organization (WHO) 2011 roadmap on NTDs (WHO, 2011a). Some countries are achieving elimination goals, more people are being reached, and the drug donation programme for NTDs, which is the largest public health drug donation programme in the world, continues to grow (Molyneux, 2004).

In most countries that had started Mass Drug Administration (MDA), efforts to prevent and treat NTDs was through disease-specific programmes that were funded through myriad sources at international level. Strategies for the different NTD programmes were not co-ordinated well to best harmonise activities and resources, and the international donor funding similarly lacked a macro strategy.

To address these shortcomings, the global NTDs community in May 2013, at the sixty-sixth World Health Assembly adopted Resolution WHA66.12 on NTDs to call on Member States to intensify and integrate measures and plan investment to improve the health and social well-being of affected populations (WHO, 2013a). This approach would be unlike the traditional vertical approach, which targeted specific diseases.

An integrated approach for the NTDs takes into consideration that all of these diseases in a particular country can be incorporated into a single programme, this could potentially make implementation easier and could facilitate an increase in the efficiencies of the delivery of drugs in a co-ordinated fashion and would lead to better costeffectiveness in an already resource-stretched country. It is very important to integrate these diseases because they demonstrate geographical overlap. In addition, drugs used as preventive chemotherapy also treat other diseases, different than those they were targeted against (Molyneux, Hotez, & Fenwick, 2005). It cannot also be over-emphasised that integration reduces cost for the country (Conteh, Engels, & Molyneux, 2010).



Figure 1.1: Overlapping of four of the most common NTDs in Liberia

Table 1.1: Four of the most common NTDs co-endemicity in Liberia, by counties

Source: MoHSW (2017a).

| | LF | Oncho | STH | SCH |
|------------------|----|-------|-----|-----|
| | | | | |
| Bomi | _ | + | + | _ |
| Bong | + | + | + | + |
| Grand Bassa | + | + | + | _ |
| Grand Cape Mount | + | + | + | _ |
| Grand Gedeh | + | + | + | _ |
| Grand Kru | + | + | + | _ |
| Lofa | + | + | + | + |
| Margibi | + | + | + | + |
| Maryland | + | + | + | _ |
| Montserrado | + | + | + | _ |
| Nimba | + | + | + | + |
| Rivercess | + | + | + | _ |
| Sinoe | + | + | + | _ |
| River Gee | + | + | + | _ |
| Gbarbolu | _ | + | + | _ |

NTDs Counties (+) = present in county (-) = absent in county

Source: MoHSW (2017a).

The present integrated strategy promoted by WHO to control NTDs including schistosomiasis (SCH), onchocerciasis (Oncho), lymphatic filariasis (LF) and soiltransmitted helminthiasis (STH) that are amenable to MDA was adopted by the Ministry of Health (MoH) Liberia in 2011, at the inception of the integrated NTD programme at the MoH. However, baseline data from mapping of NTDs in Liberia indicated that there is a high prevalence and overlap of the four most common NTDs in the country: Oncho,

LF, SCH and STH see Figure 1 and Table 1.1

According to the WHO tenth meeting of the Strategic and Technical Advisory Group for NTDs held in 2017, there are 20 NTDs in the world (WHO, 2017a):

- 1. Buruli ulcer
- 2. Chagas disease (American trypansomiasis)
- 3. Cutaneous leishmaniasis and cisceral leishmaniasis
- 4. Dengue/Chikungumya
- 5. Echinococcus

- 6. Foodborne trematode infections
- 7. Guinea worm (Dracunculiasis)
- 8. Human African trypanosomiasis
- 9. Leprosy (Hansen disease)

10. *LF* (*elephantiasis*)

11. Mycetoma, chromoblastomycosis and other deep mycoses

12. Oncho (river blindness)

13. Rabies

14. Scabies and other ectoparasites

- 15. SCH (Bilharzia)
- 16. Snakebite envenoming

17. STH

- 18. Taeniasis and cysticercosis
- 19. Trachoma (preventable blindness)
- 20. Yaws (Endemic treponematoses)

While some of the NTDs can be controllable by MDA, others require individual treatment. The NTDs that can be controlled by MDA is as follows: STH; SCH; LF (elephantiasis); Oncho (river blindness); and trachoma (preventable blindness). Guinea worm can be controlled by clean drinking water. NTDs that can be treated on an individual basis are as follows: leprosy; Buruli ulcer; Chagas disease; human African trypanosomiasis; Cutaneous Leishmaniasis; Visceral Leishmaniasis; and Dengue. The zoonotic parasitic diseases are Neurocysticercosis, and *Echinococcus*. Animals that serve as a reservoir for NTDs are: Brucellosis and rabies (Keenan, et al., 2013). The greatest burden of the disease can be found in sub-Saharan Africa (see Table 1.2)

| Disease | Prevalent cases (in millions) in 2013 | Percent change since 1990 |
|---|---------------------------------------|---------------------------------|
| Ascariasis | 804.4 | -25.5% |
| Trichuriasis | 477.4 | -11.6% |
| Hookworm | 471.8 | -5.1% |
| Schistosomiasis | 290.6 | 30.9% |
| Foodborne trematodiases | 80.2 | 51.1% |
| Dengue*+ | 58.4 | 610.9% |
| Lymphatic filariasis | 43.9 | -32.1% |
| Onchocerciasis | 17 | -31.2% |
| Chagas disease | 9.4 | 22.4% |
| Cutaneous/mucocutaneous leishamaniasis | 3.9 | 174.2% |
| Trachoma ⁺ | 2.4 | -39.2% |
| Cysticercosis ⁺ | 1 | -26.3% |
| Cystic echinococcosis ⁺ | 0.8 | -15.4% |
| Leprosy | 0.7 | 61.3% |
| Visceral leishmaniasis | 0.7 | 35.1% |
| Rabies*+ | 0.02 | -40.4% |
| African trypanossomiasis | 0.02 | -71.1% |
| Other NTDs | 59.7 | -5.0% |
| Total Cases | 2322 | NA |
| Additional NTDs | Prevalent(inmillions) incases2013 | Percentage change since 1990 |
| Trichomoniasis | 67.1 | 45.6% |
| Scabies | 66.1 | 24.8% |
| Typhoid fever* | 11 | -19.9% |
| Paratyphiod fever* | 6.4 | -27.9% |

Table 1.2: Prevalent cases of NTDs in 2013 and percent change from 1990 to 2013according to Global Burden of Disease Study (GBD) 2013

| Venomous animal contact* | 5.5 | -2.7% | |
|---------------------------|-------|--------|--|
| Cholera* | 2.3 | 6.1% | |
| Cryptosporidiosis* | 1.4 | -19.4% | |
| Amoebiasis* | 0.4 | 17.0% | |
| Total cases of additional | | | |
| neglected diseases | 160.2 | NA | |

* Incident cases in 2013 rather than prevalent cases.

+ Symptomatic cases only.

NOTE: For information on percent change calculations, see GBD 2013 capstone paper on incidence, prevalence, and years lived with disability (YLDs) [3]. All data presented in this table (except for rabies, cholera, cryptosporidiosis, and ameoebiasis) are also available from the instutite for Health Metrics and Evaluation (IHME) website and were previously published [3]. **Abbreviations:** NA, non-applicable. https://doi.org/10.1371/journal.pntd.0005424.t001

At the time when this thesis started in 2010, there was no NTD programme in Liberia, the only vertical NTD programme that was being implemented in the country was for Oncho, where there was community-directed efforts for control of Oncho using Ivermectin. This PhD thesis served as an eye opener to the MoH on the NTDs to initiate and fast track an integrated NTD programme. Table 1.3 gives the disease burden and disability-adjusted life years (DALYs) in sub-Saharan Africa resulting from NTDs.

| | | Percent change for | YLDs (in | YLLS (in |
|-------------------------|-------------------|--------------------------|--------------|--------------|
| | DALYs (in | DALYs | millions) in | millions) in |
| NTD | millions) in 2013 | 2005-2013 | 2013 | 2013 |
| Visceral leishmaniasis | 4.24 | 8.7% | 0.008 | 4.23 |
| Foodborne trematodiases | 3.63 | 14.6% | 3.63 | 0 |
| Schistosomiasis | 3.06 | -13.9% | 2.86 | 0.2 |
| Hookworm | 2.18 | -0.5% | 2.18 | 0 |
| Lymphatic filariasis | 2.02 | -14.3% | 2.02 | 0 |
| Ascariasis | 1.27 | -29.0% | 0.93 | 0.34 |

Table 1.3: Disease burden (DALYs) in sub-Saharan Africa resulting from the NTDs (global burden of disease study)

| Rabies | 1.24 | -14.6% | 0.0001 | 1.24 |
|--|-----------------------------------|---|----------------------------------|----------------------------------|
| Onchocerciasis | 1.18 | -19.4% | 1.18 | 0 |
| Dengue | 1.14 | 17.0% | 0.56 | 0.58 |
| Trichuriasis | 0.58 | -12.3% | 0.58 | 0 |
| African trypanossomiasis | 0.39 | -54.3% | 0.005 | 0.38 |
| Chagas disease | 0.34 | 4.6% | 0.1 | 0.24 |
| Cysticercosis | 0.34 | -16.4% | 0.31 | 0.03 |
| Cystic echinococcosis | 0.18 | -14.1% | 0.08 | 0.1 |
| Trachoma | 0.17 | -18.1% | 0.17 | 0 |
| Cutaneous and mucocutaneous leishamaniasis | 0.04 | 35.9% | 0.04 | 0 |
| Leprosy | 0.04 | 8.6% | 0.04 | 0 |
| Other NTDs | 3.13 | -11.8% | 2.26 | 0.87 |
| Total NTDs | 25.17 | NA | 16.95 | 8.21 |
| Additional neglected diseases | DALYs millions) in (in 2013 | Percent change DALYs for 2005- 2013 | YLDs (in millions) in 2013 | YLLs (in millions) in 2013 |
| Typhoid fever | 11.13 | -13.7% | 0.16 | 10.97 |
| Cholera | 5.17 | 20.1 | 0.04 | 5.13 |
| Parathphoid fever | 3.82 | -8.0% | 0.04 | 3.78 |
| Cryptosporidiosis | 3.46 | -29.6 | 0.19 | 3.27 |
| Venomous animal contact | 3 | -3.4% | 0.15 | 2.85 |
| Scabies | 1.71 | 4.8% | 1.71 | 0 |
| Amoebiasis | 0.38 | -23.8% | 0.04 | 0.34 |
| Trichomoniasis | 0.11 | 8.2% | 0.11 | 0 |
| Total deaths from additional neglected diseases | 28.78 | NA | 2.44 | 26.34 |

NOTE: For information on percent change calculations, see Global Burden on Disease (GBD 2013) capstone paper DALY [2]. The estimates presented in this tables are all are also available from the Instutite for Health Metrics and Evaluation (IHME) website and were previously published in [2-4]. Information on DALYs and YLDs for cholera, Cryptosporidiosis, and Amoebiasis is not available from IHME website or capestone papers. **Abbreviations:** NA, non-applicable. <u>https://doi.org/10.1371/journal.pntd.0005424.t003</u>

1.2 Objectives

To optimise the current strategies to an integrated approach for the control of NTDs in a post-conflict country and develop a platform for monitoring and evaluation of these diseases in a cost-effective manner.

1.2.1 Specific objectives

- To assess the geographical distribution and prevalence of LF in Liberia.
- To develop an integrated map of LF, SCH and STH.
- To determine the morbidity due to SCH in school-aged children in Bong County by the use of ultrasound.

1.3 General methods

The methodology used for the review of historical data on the distribution and epidemiology of *Wuchereria bancrofti* was based on a systematic review of the literature in four electronic databases and 17 articles were selected to be included in the study. For the study on the baseline mapping of LF the methodology used was that as described in the WHO guidelines for preparing and implementing a national plan to eliminate LF (WHO, 2000a) and WHO operational guidelines for rapid mapping of Bancroftian filariasis in Africa (WHO, 2000b).

For the epidemiology, update started with studies on urogenital and intestinal SCH with observations on STH with a cross-sectional study of 10 schools in Bong County, northern Liberia, were investigated using the Kato-Katz method. A semistructured questionnaire was used to access the knowledge, attitude and practice (KAP) towards STH, SCH and LF amongst school-aged children in Bong County, northern Liberia. For the ultrasound study a Mindary diagnostic system ultrasound Model MS

MR-17003578 with a convex transducer was used to access morbidity due to SCH. The Niamey protocol was used to score lesions within a graded system.

1.4 Justification of the study

At the genesis of this study, Liberia did not have an established NTD programme and had not started MDA for the control of LF and SCH. There was an unco-ordinated distribution of the drug Mebendazole to schoolchildren in an attempt to decrease the burden of worm infestation in the country. The epidemiology and burden of these diseases were not adequately known. Therefore, an integrated approach provided a unique opportunity to properly co-ordinate the NTD activities in the country. The MoH has now developed an integreated approach to NTDs as suggested by this study.

This research project was carried out in collaboration with the national NTD programme at the Ministry of Health and Social Welfare. It looked at the impact of MDA after two rounds of treatment, which could determine when to stop MDA, and the impact of MDA on morbidity control.

1.5 Study area

Liberia is a post-conflict country with 14 years of civil war, which ended in 2003. The deadly Ebola virus disease (EVD) hit the country in 2013 where hundreds lost their lives. Liberia is located at latitude 60:30 North of the equator and longitude 90:30 West of Greenwich meridian, with a landscape of 111,370 square kilometres. It is bordered by the Atlantic Ocean to the South, Côte d'Ivoire to the East, Sierra Leone to the North-West, and Guinea to the North-East (see Figure 2). Administratively, it is divided into 15 counties with a total population of 3,476,608 in 2008 (see Table 1.4).

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| Торіс | Status |
|--|--|
| Geographic size | 111,369 square kilometres |
| Annual rainfall | 4,000 mm (one of the highest in the world) |
| Natural resources | Iron ore, rubber, timber, diamonds, and gold |
| Founded | July 26, 1847 |
| Executive | President: Ellen Johnson-Sirleaf (2006) |
| Legislature | Bicameral (Senate and House of Representatives) |
| Per capita gross domestic product (GDP) | US\$247 (2010 estimate) |
| GDP growth rate | 1.8% (2001–2010 estimates), 5.9% (2010 estimate) |
| Population living on less than US\$1 per day | 76.2% |
| Population | 3,476,608 (32% in Monrovia; 2008 Census) |
| Population growth rate | 2.1% (2008 Census) |
| Life expectancy | 59.1 years (UNDP, 2010) |
| Under-5 mortality rate | 114/1000 live births (DHS, 2007) |
| Maternal mortality rate | 994/100,000 live birth (DHS, 2007) |
| Access to improved drinking water | 75.0% (93.0% urban, 58.0% rural) (LMIS, 2009) |
| Access to adequate sanitation | 44.0% (63.0% urban, 27.0% rural) (LMIS, 2009) |
| Human immunodeficiency virus (HIV) seroprevalence | 1.5% (1.8% female, 1.2% male) (DHS, 2007) |
| Supervised childbirth | 46.0% (DHS, 2007) |
| Institutional deliveries | 37.0% (DHS, 2007) |
| Vaccination coverage (full) | 51.0% (2010) |
| Net enrolment primary school | 74.0% male, 58.0% female (2000–2006 average) |
| Net enrolment secondary school | 37.0% male, 27.0% female (2000–2006 average) |

Table 1.4: Liberia at a glance, April 2011

Source: Adapted from MoHSW (2011a).

Figure 1.2 Political Map of Liberia



Source: MoHSW (2011a).

CHAPTER 2: LITERATURE REVIEW

2.1 LF

2.1.1 A brief history of LF

LF also known as elephantiasis is one of the ancient diseases and furthermost disfiguring of all the NTDs. There are 73 countries across the globe that are endemic for the disease and an estimated 1.39 billion population live in areas where filariasis is endemic and therefore are at risk of infection and transmission (WHO, 2012a). In the context of control some countries have reached elimination for LF while others have started preventive chemotherapy.

In 1993, LF was recognised as a potentially eradicable disease by the International Task Force for Disease Eradication (Ichimori & Graves, 2017). In 1997, the World Health Assembly recognised LF as a public health problem and targeted it for elimination by 2020; it then approved Resolution WHA50.29 calling on Member States to begin programmes for the elimination of the disease. In 2000, WHO launched the Global Programme to Eliminate Lymphatic Filariasis (GPELF), with the aim to stop the spread of LF infection (interrupting transmission) and alleviate the suffering of affected persons (controlling morbidity) (WHO, 2010a).

The antiquity of LF dates as far back as 600 BC when some Indian and Persian physicians described clear signs and symptoms of LF. In the Temple of Hatshepsut in Egypt (1501–1480) there are drawings on the tomb, which depicts the presence of LF as early as 1500 BC; also at the Tokyo National Museum in Japan, both males and females with LF dating as far back as AD 1100–1200 was found in the 'Disease Picture Scroll' (Otsuji, 2011) (see also Figure 2.1).

Figure 2.1: In Japan, picture of a Japenese woman (d) with elephantiasis-like of the lower limb



Source: History and control of LF (Otsuji, 2011).

In the year 1863, a French surgeon Jean Nicolas Demarquay (1814–1875) was the first to discover microfilaria in the fluid collected from a Cuban with hydrocele. This was followed by the discovery of adult worms in 1876 by Joseph Bancrofti, which is now known as *Wuchereria bancrofti*. Patrick Manson in 1877 was the first to describe the lifecycle of LF. Manson was able to determine that the vector of the disease was mosquitoes. George Carmichael-Low was the first to identify LF transmission
mechanism, where he noted that individuals were infected from the bite of infected mosquitoes (Despommier, Gwadz, & Hotez, 1995).

2.1.2 The global burden of LF

LF is caused by infection with the parasitic filarial nematodes (roundworms) its species are as follows: *W. bancrofti; Brugia malayi;* and *Brugia timori*. The filarial infection is transmitted by mosquitoes and humans are definitive hosts. Once an individual gets infected the infection inhabits the lymphatics and subcutaneous tissues, which eventually leads to damage of the lymphatic vessels, resulting in clinical disease manifested as hydrocele, lymphoedema and elephantiasis. LF is the second leading cause of disability globally with an estimate of about 120 million people infected with the nematodes and approximately 40 million people are suffering from its complications. It is a major cause of disfigurement and disability in endemic areas, leading to significant economic and psychosocial impact (WHO, 2017b).

According to the GPELF 2011 report, nearly 40 million people are suffering from the stigma and incapacitating clinical expressions of the disease, of which 15 million have lymphoedema and 25 million men have urogenital swelling, which is mainly scrotal hydrocele (WHO, 2019).

W. bancrofti is found in sub-Saharan Africa, South East Asia, the Indian subcontinent, parts of the Pacific Islands, parts of Latin America, and the Caribbean.

Brugia malayi occurs mainly in India, Malaysia, the Philippines, Indonesia, and various Pacific Islands where it may co-exist with *W. bancrofti. Brugia timori* occurs on the Timor island of Indonesia. Approximately two-thirds of individuals infected with LF are in Asia (see Figure 2.2).

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countries, in 2014

Source: GPELF programme, progress report (2014).

The epidemiology of LF has changed due to the MDA initiated by GPELF to cut the disease transmission. This has led to the reclassification of some countries (e.g., Trinidad and Tobago and Costa Rica) as non-endemic, while others such as Cambodia, China, the Cook Islands, Egypt, Maldives, the Marshall Islands, Niue, Palau, Republic of Korea, Thailand, Togo, Tonga Vanuatu, Vietnam, and Wallis and Futuna have eliminated transmission completely (GPELF progress report 2016; WHO, 2019).

The disease was observed to occur intermittently in children. However, in recent years, sensitive diagnostic tests (e.g., ultrasound examination, antigen detection) have now revealed that LF can be acquired in early childhood. About one-third of children infected with LF before the age of 5 remain subclinical for years or present with nonspecific presentations of swollen lymph nodes or glands, but after puberty the clinical manifestation of the disease is seen (Witt & Ottesen, 2001).

A study of 159 Kenyan pregnant women with active *W. bancrofti* infection, their neonates cord blood was tested at birth and was shown not to have the filarialspecific T cell

responses, had a 13-fold increased risk of developing childhood LF infection compared to the uninfected controls (Ondigo et. al., 2018).

In endemic areas, it has been observed that the prevalence of the microfilaremia increases with age and by the third or fourth decade of life most people in endemic areas have been exposed. The adult worm does not replicate within the human host, therefore, if an individual leaves an endemic area, the adult worm burden cannot increase, since exposure to infective larvae has ceased (Witt & Ottesen, 2001).

2.1.3 The vector

In 1877, Sir Patrick Manson proved that mosquitoes were the vectors for filariasis, which also indicated an attach point for control. In some sub-Saharan African countries where malaria is endemic, the use of mosquito nets to control LF has been encouraged by some NTD programmes at the MoH in collaboration with the National Malaria Control Programme. Mosquito vectors for *W. bancrofti* include *Aedes, Culex, Coquillettidia, Mansonia* and *Anopheles*. Vectors for *Brugia* are *Aedes* and *Mansonia*. In Africa, the *Anopheles gambiae* complex is the dominant vector for LF. There are reports of urban and rural transmissions of the disease, which is mostly due to travelling, but the transmission is low, and this could be attributed to the use of bednets, mosquito repellents and MDA. Humans are the only host for Bancroftian filariasis in comparison to Brugian filariasis, which infects humans and domestic and wild animals (Mwakitalu, Malecela, Pedersen, Mosha, & Simonsen, 2013).

2.1.4 In sub-Saharan Africa

According to the ranking of NTDs in sub-Saharan Africa by prevalence it is reported that approximately 40.0% of the world's 120 million cases of LF occur in sub-Saharan

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Africa, with approximately 46 million to 51 million LF cases. A review of the prevalence and distribution of NTDs reported that and an estimated 6.0% to 9.0% of the sub-Saharan African population is infected with LF with an estimated 382 million to 394 million people are at risk of infection in sub-Saharan Africa, of which this figure includes 176 million children (Hotez, et al., 2006). In sub-Saharan Africa, 39 Member

States are endemic of LF. The highest number of people at risk of LF are from Nigeria, Democratic Republic of Congo, Tanzania, Ethiopia and Kenya (WHO, 2008).

NTDs are associated with poverty as it occurs in countries with low economic power and where there is poor sanitation, poor hygiene and a poor health system. NTDs are common amongst the rural communities and in some slum communities in Africa (Hotez, et al., 2007) (see Table 2.1).

Table 2.1: Poverty in sub-Saharan Africa compared to the world

| Percentage of population living on less than US\$1.25 per day | 51.0% |
|--|----------------|
| Total population living on less than US\$1.25 per day | 390.60 million |
| | 28.0% |
| Percentage of world's population living on less than US\$1.25 per da | у |
| Total population living on less than US\$2 per day | 556.7million |
| Percentage of world's population living on less than US\$2 per day | 22.0% |

Source: Hotez, et al., (2007).

2.1.5 In West Africa

In West Africa, the vectors of malaria, the female *A. gambiae* mosquitoes, are the main vectors of LF. Current practices in the management of LF in West Africa have been influenced by the push for integrated control of NTDs amenable to MDA and vector

control is now being encouraged for the prevention, control, elimination and eradication of NTDs (de Souza, et al., 2012).

2.1.6 In Liberia

Historical review conducted for LF in Liberia indicates that the disease dates as far back as the 1900s. Data on LF in Liberia shows the infection is caused by *W. bancrofti* and the parasite is transmitted by *A. gambiae and Anopheles funestus mosquitoes* (see Chapter 3).

Figure 2.3 Prevalence and distribution of LF in Liberia Source:

MoHSW (2011a).



A nationwide LF mapping exercise was conducted in 2010 by the Liberian MoH and partners with technical and financial assistance from WHO, and the Centre for Neglected Tropical Diseases/Liverpool School of Tropical Medicine (CNTD/LSTM). From the results, 13 of the 15 counties were endemic for LF. Clinical evidence of LF has been seen in many parts of rural Liberia, especially along the coastal regions (see Figure 2.3).

2.1.7 The parasite and its life cycle

The Centers for Disease Control and Prevention (CDC) (see Figure 6) provide a vivid description of the life cycle of *W. bancrofti*, the explanation is as follows: The life cycle of filariasis commences after a blood meal when the filarial larvae is injected into human skin by a mosquito. The larvae then travel through the biting site and enter the lymphatic vessels where they reside. After 9 months, the larvae develop into adult worms. The female and male adult worms mate and produce microfilariae. The microfilaria then goes into the lymph and blood vessels of the infected person, who if gets bitten by the mosquito, the microfilaria then develops into third stage larvae and can infect another person if they also get bitten by the infected mosquito.

In some parts of the world, the microfilaria can be visible in the bloodstream during the evening hours, mostly between 22:00 p.m. and 02:00 a.m., where this is called 'nocturnal periodicity'. In the South Pacific, the microfilaria can be found any time in the bloodstream, and this is called 'subperiodic', but they are found in high levels during midday. It takes about 12 months from the bite of a mosquito to the detection of microfilariae in the bloodstream of an infected person. The adult worm lives for about 5 years, but in some few cases can live up to 12 to 15 years (CDC, 2019a).

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Source: Reproduced from CDC (2019a).

2.1.8 Clinical presentation of LF

The symptoms and signs of LF when contracted in childhood does not manifest itself until adolesence or adulthood. However, there are reports that state that children in endemic regions have suffered lymphoedema of the limbs and acute dermatolymphangio-adenitis (ADLA) signs and symptoms. Approximately 70.0% of people infected with LF may have asymptomatic microfilaremia, while some may progress to the clinical manifestations of the disease and develop lymphatic dysfunction causing lymphoedema, hydrocele, scrotal and elephantiasis. It has also been reported that people who have newly arrived to disease endemic areas may develop afebrile episodes of lymphadenitis and lymphangitis. The common acute clinical presentation of LF is ADLA, which manifests as nocturnal coughing, wheezing, fever, chills, pain in the affected area, vomiting and eosinophilia. Lymphoedema of the lower limb is the most frequent chronic clinical presentation of LF and is graded as:

Grade I: Pitting oedema, reversible on elevation of the affected limb.

Grade II: Pitting or non-pitting oedema, which does not reverse on elevation of the affected limb and there are no skin changes.

Grade III: Non-pitting oedema that is not reversible, with thickening of the skin.

Grade IV: Non-pitting oedema that is not reversible, with thickening of the skin along with nodular or warty excrescences – the stage of elephantiasis.

The other lymphoedema of the upper limb, breast and male genitalia, are less commom

as compared to the lower limb (Kumaraswami, 2000).

2.1.9 Diagnosis of LF

To make a diagnosis of LF, a complete history taken from patients is necessary. A high suspicion of LF should be suspected in anyone with an exposure history who presents with pathognomonic signs and symptoms or unexplained eosinophilia and who have travelled to or live in endemic areas. Definitive diagnosis can be made by various methods, which range from detection of circulating filarial antigen (for *W. bancrofti* infection only), demonstration of microfilariae or filarial deoxyribonucleic acid (DNA) in the blood, to detection of the adult worms located in the lymphatics (Pani, et al., 1995).

2.1.10 Treatment and prevention of LF

The treatment of LF varies from country to country depending on the co-endmicity of some NTDs. Diethylcarbamazine (DEC) is the best treatment for people with LF, but the drawback is that the drug is contra-indicated in patients with Oncho due to the possibility of the Mazzotti reaction and as a result this should be used in caution with patients with Loaiasis (see also Table 2.2).

| Disease | Drugs and dosages | Threshold for implementation | Frequency intervention | of |
|---------|-------------------|------------------------------|---------------------------|----|
|---------|-------------------|------------------------------|---------------------------|----|

Table 2.2: Current treatment guidelines for LF, STH and SCH

| LF | Albendazole 400 mg for | Prevalence of >1.0% | Annual: | |
|--------------------------------|--|---|---|--|
| | children aged >2 years plus DEC 6mg/Kg in countries where Oncho is not coendemic Or Ivermectin 150µ/Kg in countries where Oncho is endemic | | Treatment must be combined with limb care of patients with elephantiasis or hydrocele surgery | |
| STH (i.e., | Albendazole 400 mg for | Prevalence >50.0%: treat school-aged children, and | Annual or twice | |
| <i>Ascaris</i> , hookworms, | children above 2 years, or Mebendazole 500 mg | adults at high risk, twice yearly | yearly (depending on prevalence): | |
| and Trichuris trichiura) | | Prevalence >20.0% to <50.0%: treat school-agedchildren once per year.Prevalence <20.0%: individualised treatment. | Water and sanitation strategies must be implemented | |
| | | Preschoolchildren and women of child-bearing age should also be treated (as part of maternal and child health programmes) | | |
| SCH | Praziquantel 40 mg/Kg | Prevalence >50.0%: treat all school-aged children. | Annual treatment: | |
| | (using dose pole) for children over 4 years (or 94 cms) | Adults at high risk may also be treated | Treatment holidays can be given if prevalence drops. | |
| | | Prevalence >10.0% to <50.0%: treat children once every 2 years | Water and sanitation strategies | |
| | | Prevalence <10.0%: individualised treatment | must be implemented | |

Source: WHO (2006a).

Ivermectin is effective in reducing the microfilaremia, but this has no effect on the adult worm. Albendazole has no direct effect on microfilariae, but this leads to a slow decline in microfilaremia due to its macrofilaricidal activity against the adult worms. However, side effects due to rapid killing of microfilariae have not been seen when Albendazole is used with Ivermectin during MDA (Partono, Maizels, & Purnomo, 1989). MDA has been the concentration of control of the disease. Vector control with insecticide-treated nets (ITNs) is also useful for *W. bancrofti* elimination, in communities where *Anopheles* mosquitoes transmit the parasite. Unfortunately, there is no vaccine available for LF.

2.2 SCH

2.2.1 A brief history of SCH

SCH is one of the ancient parasitic infections affecting humans. Symptoms of the disease such as haematuria were first recognised as far back as 1900 BC by Egyptian physicians. They called the disease 'menstruating males of Egypt'. In the year 1851, a German Physician, Theodor Maximilian Bilharz, (1825–1862) was the first to discover the disease by finding long white helminths in the portal vein. The discovery was made while he worked at the Egyptian department of hygiene, he then described long white helminthes in the portal vein of men who died of haematuria in series of autopsies he carried out. He noted that these changes were consistent across the cadavers on which he performed the autopsies. He also noted helminthes eggs in the stools of the cadavers. The disease is also called 'Bilharziasis' named after Bilharz for his contributions to the disease Brazilian Parasitologist, Manuel Augusto. Pirajá da Silva (1873–1961) also made a contribution to the discovery of the life cycle of intestinal SCH ,(Tan & Ahana, 2007).

2.2.2 The global burden of SCH

SCH is a disease caused by infection with parasitic blood flukes. Although it is controlled in some countries, it still remains a major public health problem. There are an estimated 207 million cases of SCH worldwide of which 93.0% occur in sub-Saharan Africa (Steinmann, Keiser, Bos, Tanner, & Utzinger, 2006). The parasites that cause SCH live in certain types of freshwater snail in subtropical and tropical regions. People living around swamp lands, dams and reservoirs are particularly at risk for the disease.

The disease occurs when the skin comes into contact with contaminated water and is penetrated by the schistosome parasite (Brooker, 2007). This is graphically shown in Figure 2.5



Figure 2.5: Life cycle of SCH

Source: Reproduced from CDC (2019b).

Increasing population and movement have contributed to increased spread and transfer of SCH to new areas. Most endemic countries are amongst the developing countries in the world with poor health systems. Constraints to disease control in these developing countries vary from lack of political commitment to limited or poor infrastructure for public health interventions (Chitsulo, Engels, Montresor, & Savioli, 2000).





Source: http://gamapserver.who.int/maplibrary/files/maps/global_schistosomiasis_2009

2.2.3 Epidemiology

S. mekongi

It has been reported that there are approximately 207 million people infected with SCH, amongst which 120 million are symptomatic, 20 million have severe disease, about 600 million people are at risk of infection and around 200,000 deaths may occur every year (Ultroska, et al., 1989). This is indeed a public health emergency and should claim the attention of every national government in the world.Figure 2.6 shows the distribution of SCH in 2009.

| | Species | Geographical distribution |
|--------------------------------|---|--|
| | Schistosoma manson | <i>i</i> Africa, the Middle East, the Caribbean, Brazil, Venezuela, and Suriname |
| Intestinal SCH | Schistosoma japonici | China, Indonesia, the Philippines |
| | Schistosoma mekong | <i>i</i> Several districts of Cambodia and the Lao People's Democratic Republic |
| | Schistosoma guineer related S. intercalatu | asis Rainforest areas of Central Africa and m |
| Urogenital SCH | Schistosoma haer | <i>matobium</i> Africa, the Middle East, Corsica (France) |
| Source: Nicolls, e | t al., (2008). | |
| <u>Table 2.4: H</u> | uman schistosome spe | cies and snail |
| Human schis | tosome S | Snail (mollusc) species |
| S. mansoni | E | Biomphalaria |
| S. haematobi S. intercalati | ium I Im | Bulinus |
| S. japonicum | e c | Dncomelania |

| Table 2.3: | Geographical | distribution | of SC | CH |
|-------------------|----------------|--------------|--------|----|
| | o togi upiniou | | 01 0 1 | |

Tricula

Every human schistosome species has a special snail as its intermediate host. In addition, the environment plays a significant role in the locality of the fresh water snails (Clements, Moyeed, & Brooker, 2006). Table 8 outlines the human schistosome species and the genus or snail associated with transmission.

2.2.4 In Africa

SCH is widely distributed in Africa. Of the global 207 million estimated cases of SCH, 93.0% are from sub-Saharan Africa. The following sub-Saharan African countries account for the highest prevalence of SCH and are listed in order of highest prevalence, respectively: Nigeria; Tanzania; Ghana; and the Democratic Republic of Congo (Adenowoa, Oyinloyea, Ogunyinkaa, & Kappo, 2015). Due to MDA using Praziquantel and other successful control projects in a number of countries, the distribution of SCH has altered over the past years. However, despite all the interventions, the number of people infected or at risk of infection remains the same (Steinmann, et al., 2006).

2.2.5 In Liberia

Studies on the epidemiology of SCH were carried out at the Liberian Institute for Biomedical Research (LIBR) from the 1970s to 1989. Many water bodies (streams and ponds) in Lofa, Bong, and Nimba Counties were considered to be the sources of infection. These unpublished reports were lost during the Liberian civil crisis. However, a study was conducted by Dennis et al. in 1983, on the prevalence and intensity of schistosomial infections in Bong County and the bionomics of the snail intermediate hosts, which found some fresh water bodies to be harbouring the snails (*Biomphalaria* and *Bulinus*) species. The study also revealed a higher overall prevalence of *S. mansoni* (25.0%) as compared to *S. haematobium* (23.0%). This result correlated with the 2011 integrated mapping survey for STH and SCH using the Kato-Katz method. The results

indicated that SCH was prevalent in Bong County at 63.0% for *S. mansoni* and 56.0% *for S. haematobium*. Figures 9 and 10 show the prevalence and distribution in Liberia for *S. mansoni* and *S. haematobium*.



Figure 2.7: S. mansoni prevalence and distribution in Liberia

Source: MoHSW (2011a).

Figure 2.8: S. haematobium prevalence and distribution in Liberia



Source: MoHSW (2011a).

Figure 2.9: SCH endemicity in Liberia



Source: MoHSW (2011a).

2.2.6 Pathogenesis of SCH

Only few people who become infected with the disease develop symptoms. The natural course depends on the following three factors: i) age of initial exposure; ii) intensity of exposure; and iii) immunity and genetic susceptibility (Gryseels, Polman, Clerinx, & Kestens, 2006). It has been reported in 1998 by Butterworth that, the intensity of the infection increases during the first two decades of life and decreases in adulthood.

2.2.7 Clinical manifestation of SCH

The clinical disease is due to the infected person's response to the migrating eggs, which could sometimes lead to entrapment, inflammation and subsequent fibrosis (Gryseels, 1996). Many times, when the cercariae enter the host, it is associated with fever,

diarrhoea or respiratory symptoms. For the intestinal SCH the disease may progress and lead to hepatomegaly or splenomegaly. Heavy infection may lead to periportal fibrosis. Infection with the urogenital SCH, which is *S. haematobium*, the disease may remain in the urinary tract causing dysuria or haematuria. In late stages of the disease, bladder calcifications may be observed, which could lead to an increased risk of bladder cancer (Pollack, 1981).

2.2.8 Diagnosis of SCH

There are many screening tools available for the diagnosis of SCH such as serology, urinalysis and microscopy of urine and/or stools. Some studies have reported that antibodies detection is more sensitive than egg detection for the demonstration of the infection (Stelma, Talla, Verle, Niang, & Gryseels, 1994; Clerinx, Bottieau, Wichmann, Tannich, & Van Esbroeck, 2011).

2.2.9 Treatment and prevention of SCH

There are three important purposes on the treatment of SCH.

- 1. Preventing complications from the diease.
- 2. Reduction of egg production.
- 3. Reversing the chronic or acute disease.

The drug used for the treatment of SCH is Praziquantel, as well as good sanitation and safe drinking water. SCH control strategies include health education, MDA, and improvement of water and sanitation.

2.3 STH

2.3.1 A brief history and global burden of STH

In 1943, Dr Norman R. Stoll (1952 to 1973) an American parasitologist conducted a survey amongst some elementary schoolchildren in Princeton, USA, where he noted that the schoolchildren harboured worm infections. In 1946, de Silva and colleagues

also reported that there was a high burden of STH amongst North Americans (de Silva et al. 2003).

STH is the most prevalent NTD in the world. There are four main nematode species of human STH infections, also known as geohelminths: *Ascaris lumbricoides, Trichuris trichiura* (whipworm), and the hookworms *Ancylostoma duodenale* and *Necator americanus*. These infections occur mostly in developing countries especially in tropical and subtropical regions. Their prevalence in these regions can be attributed to: limited clean drinking water; poor health care; and poor sanitation systems (Haque, 2007). WHO in its progress report 2001–2010 and its strategic plan 2011–2020 reported that children aged 1 to 4 years in STH endemic areas are also at high risk for STH infection. The diseases related to infections of STH can be chronic or asymptomatic and include: malnutrition; cognitive development; growth disorders (short stature); and malabsorption of nutrients. This could lead to poor performance in school. According to the WHO estimates, approximately 836 million children around the world are in need of medication for STH. In 2016, WHO in its report on the global health observatory data reported that at least 75.0% of school-aged children had received medication for

STH (Shi, et al., 2015).

2.3.2 Epidemiology and transmission of STH

Global estimates of the disease report that around 2 billion people are infected with STH where the highest numbers are in Asia followed by sub-Saharan Africa. According to the CDC, globally 807 million to 1,121 million of the STH infections are estimated to be due to *Ascaris*, 604 million to 795 million are due to whipworm, while 576 million to 740 million are due to hookworms. Transmission of STH is associated with poor sanitation practices, infected persons whose faecal contaminates soil, food or water and

who have been barefooted (de Silva, et al., 2003). Figures 2.10, 2.11 and 2.12 illustrate the lifecycle of STH (hookworm, *T. trichiura* and *Ascaris*).

Figure 2.10: The hookworm life cycle



Source: Reproduced from CDC (2019c).

Figure 2.11: The *T. trichiura* life cycle



Source: Reproduced from CDC (2017). Figure 2.12: *A. lumbricoides* life cycle



Source: Reproduced from CDC (2019d).

2.3.3 In Liberia

STH namely, *A. lumbricoides*, *T. trichiura* and hookworms are widely distributed in Liberia; and are prevalent in all 15 counties. A nationwide study conducted in 2011 on 3,144 schoolchildren in 59 schools, showed that the highest prevalence at 50.0% to 100.0% was found in the south-eastern counties (Maryland, Grand Kru, Sinoe and Rivercess). Maryland, Grand Kru, Rivercess and Sinoe Counties in the central part of Liberia showed moderate prevalence at 20.0% to 50.0%. The lowest prevalence at 0.1% to 20.0% was found in the northern counties including: Lofa; Bong; Gbarpolu; Bomi; Montserrado; and Nimba.





Source: MoHSW (2011a).

Results showed that prevalence of *Ascaris* was 20.0%, hookworm was 9.0% and *T. trichiura* was 3.0%, (MoHSW, 2011a). Table 9 shows the results of the STH prevalence survey conducted in 2010 using the Kato-Katz method.

| County | Location/site (GPS co-ordinates) | <u>N</u> | <u>A.</u> lumbricoides | <u>Hookwor</u> <u>m</u> | <u>T. trichiura</u> |
|------------------|--|----------|---------------------------|----------------------------|---------------------|
| Romi | Klay (6.75566"N-10.85056"W) | 100 | 17(17%) | 2(20%) | 3(3%) |
| Doim | Senjeh (6.86498"N -10.8194"W) | 100 | 9(9%) | 5(5%) | 2(2%) |
| Sub-total | | 200 | 26(13%) | 7(4%) | 5(3%) |
| Bong | Gbarnga (7.11379"N -9.46214"W) | 100 | 21(21%) | 1(1%) | 0(0%) |
| Dolig | Suakoko (6.95005"N -9.45022"W) | 99 | 4(4%) | 4(4%) | 1(1%) |
| Sub-total | | 199 | 25(13%) | 5(3%) | 1(1%) |
| | Garwula (7.06343"N -10.87632"W) | 100 | 32(32%) | 6(6%) | 1(1%) |
| Grand Cape Mount | Tewor (7.06275"N -10.04662"W) | 100 | 17(17%) | 3(3%) | 0(0%) |
| Sub-total | | 200 | 49 | 9 | 1(1%) |
| Grand Bassa | Buchanan (6.00743"N -9.9009"W) | 100 | 13(13%) | 11(11%) | 2(2%) |
| | District #3 (5.88508"N) | 100 | 16(16%) | 6(6%) | 1(1%) |
| Sub-total | | 200 | 29 | 17(9%) | 3(2%) |
| Crand Cadab | Tchien (6.96887"N -11.31224"W) | 100 | 16(16%) | 0(0%) | 1(1%) |
| Grand Gedeh | Gbarzon (6.71755"N -11.06412"W) | 100 | 22(22%) | 5(5%) | 6(6%) |
| Sub-total | | 200 | 38(19%) | 5(3%) | 7(7%) |
| Grand Kru | Barclayville (6.0652"N - 8.13602"W) | 100 | 53(53%) | 5(5%) | 3(3%) |
| | Buah (5.19103"N -7.96096"W) | 100 | 50(50%) | 10(10%) | 3(3%) |
| Sub-total | | 200 | 103(52%) | 15(8%) | 6(3%) |
| Gharnolu | Bopolu (4.94518"N -8.16542"W) | 99 | 9(9%) | 6(6%) | 2(2%) |
| Obarpola | Gbama (4.67583"N -8.22685"W) | 100 | 6(6%) | 13(13%) | 3(3%) |
| Sub-total | | 199 | 15(8%) | 19(10%) | 5(3%) |
| Marthursen | Careysburg(8.41415"N - 9.72119"W) | 197 | 13(7%) | 10(5%) | 1(1%) |
| Montserrado | Left Bank (8.38408"N- 10.20299"W) | 199 | 6(3%) | 7(4%) | 5(3%) |
| Sub-total | | 396 | 19(5%) | 17(4%) | 6(2%) |
| Moryland | Happer (6.67074"N -10.22611"W) | 100 | 56(56%) | 8(8%) | 33(33%) |
| Maryland | Pleebo (6.71456"N -10.21816"W) | 100 | 53(53%) | 17(17%) | 11(11%) |

Table 2.5: Prevalence of STH, by county in Liberia

| Sub-total | | 200 | 109(55%) | 25(13%) | 44(22%) |
|-------------|---------------------------------------|-------------------|----------|---------|---------|
| Mongihi | Gibi (4.3765"N -7.71116"W) | 100 | 16(16%) | 6(6%) | 1(1%) |
| wargibi | Kakata (4.38449"N -7.69465"W) | 100 | 11(11%) | 10(10%) | 0(0%) |
| Sub-total | | 200 | 27(14%) | 16(8%) | 1(1%) |
| Nimba | Sanniquellie(6.43304"N- 1.49024"W) | 100 | 4(4%) | 7(7%) | 0(0%) |
| - (IIII) | Saclepea (6.34755"N -7.72861"W) | 100 | 4(4%) | 6(6%) | 0(0%) |
| Sub-total | | 200 | 8(4%) | 13(7%) | 0(0%) |
| Lofa | Foya (7.28003"N -8.82301"W) | 100 | 9(9%) | 10(10%) | 3(3%) |
| Luia | Voinjama (7.10562"N -8.90699"W) | 100 | 10(10%) | 1(1%) | 1(1%) |
| Sub-total | | 200 | 19(10%) | 11(6%) | 4(2%) |
| Since | Greenville (5.52205"N -9.61881"W) | 100 | 22(22%) | 44(44%) | 5(5%) |
| Since | LowerKanyan(5.68675"N -9.50088"W) | 100 | 20(20%) | 25(25%) | 4(4%) |
| Sub-total | | 200 | 42(21%) | 69(35%) | 9(5%) |
| | Central C (5.2204"N -8.00809"W) | 100 | 40(40%) | 25(25%) | 8(8%) |
| Rivercess | Timbo (5.19608"N -7.38255"W) | 50 | 26(52%) | 25(50%) | 5(10%) |
| Sub-total | | 150 | 66(44%) | 50(33%) | 13(9%) |
| River Cee | Gbeapo (5.01103"N -9.03833"W) | 100 | 34(34%) | 2(2%) | 2(2%) |
| | Potupo (4.9893"N -8.96154"W) | 100 | 20(20%) | 0(0%) | 0(0%) |
| Sub-total | | $\frac{200}{3.1}$ | 54(27%) | 2(1%) | 2(1%) |
| Grand Total | | 44 | 629(20%) | 280(9%) | 107(3%) |

Source MoHSW (2017a). GPS = global positioning system.

2.3.4 Clinical presentation, diagnosis and treatment of STH

Infected persons initially may at times present with respiratory manifestations. They may also present with vomiting or coughing during the migration of the adult worm (Proffitt & Walton, 1962). Complications of STH vary from iron deficiency anaemia, stunted growth, malnutrition, intestinal obstruction, learning impairment to pancreatitis and hepatobillary involvement.

The diagnosis of STH can be established by good history taking and physical examination. Laboratory diagnosis on stool examinations to detect worm eggs is commonly practised in developed countries. The Kato-Katz method is the most widely used method for the diagnosis of STH. However, there are still no reliable serologic tests available to diagnose STH (Lamberton & Jourdan, 2015). Anthelminthic treatment of STH infection is prescribed as follows: Albendazole (400 mg once) or Mebendazole 100 mg twice daily for 3 consecutive days. (See table 2.2)

2.3.5 Prevention of STH

Preventive measures of STH entails good hygiene, access to safe drinking water, proper washing and cooking of food, frequent handwashing and protection of the feet for those in endemic areas. Deworming programmes in schools administering anthelminthic drugs (Albendazole 400 mg for children above 2 years, or Mebendazole 500 mg, see Table 6) and preventive chemotherapy targeting at-risk populations are measures to maintain low individual worm burdens (Albonico, Montresor, Crompton, & Savioli, 2006).

2.4 Prevalence and distribution of onchocerciasis in Liberia

The African program for Onchocerciasis, conducted a rapid epidemiological mapping of Onchocerciasis (REMO) for Liberia in 1999 and reported that the disease is prevalent in all 15 counties. With an estimated 1,113,213 of Liberia's 4 million population at risk. Onchocerciasis in Liberia was first diagnosed by the Harvard expedition to Africa in 1926 – 1927 and was found to be more prevalent in the rainforest region of the country as compared to the coastal regions. *Simulium yahense* has been identified as the *Simulium* species that transmits the disease in Liberia. Since the REMO mapping. The

Ministry of Health and Social Welfare in collaboration with its partners has conducted Mass Drug Administration for onchocerciasis in the country and has achived a therapeutic coverage of 83%. (Ministry of Helath NTD master Plan 2016-2020.)



Figure 2.14 REMO mapping of Liberia 1999

Source: https://www.who.int/apoc/cdti/remo/en/

CHAPTER 3: A CRITICAL ANALYSIS OF HISTORICAL DATA ON THE EPIDEMIOLOGY AND DISTRIBUTION OF *W. BANCROFTI* IN VECTOR AND HUMAN POPULATIONS IN LIBERIA

3.1 Abstract

3.1.1 Background

The filarial infection is transmitted by several species of mosquitoes within the genera *Culex, Aedes* and *Anopheles*. In West Africa, LF is transmitted by Anopheles mosquitoes. Humans are the definitive hosts. The adult worms inhabit the lymphatics and subcutaneous tissues causing damage to them resulting in the clinical manifestation of the disease as hydrocele, lymphoedema and elephantiasis.

3.1.2 Objectives

To examine the epidemiology of LF in humans and the distribution of *W. bancrofti* mosquito populations in Liberia using historical data. This chapter does not contain new data collected by the researcher. The data is based on historical review.

3.1.3 Methods

A systematic review of the literature in four electronic databases (PubMed, University of Liverpool Library, the National Center for Biotechnology Information [NCBI] bookshelf, and WHO Library – Thomas Allen) published before the Liberian civil war, which occurred in 1990, was undertaken. The search terms used were 'vector', 'lymphatic filarisis' and 'Liberia'. In total, 1,689 titles and abstracts were identified, where 17 articles were selected to be included in this paper using thematic analysis. To augment the literature on epidemiology of LF and medical entomology, fieldwork was undertaken involving collection of vectors by wild-caught and laboratory-reared mosquitoes and serology was performed by peripheral blood smear.

3.1.4 Results

A total of 17 articles met the inclusion criteria and their results are as follows:

- *Entomology:* Based on the dissections of the mosquitoes for the presence of the infective larvae of *W. bancrofti* incrimination of the main vector for the distribution of LF in Liberia was *A. gambiae* followed by *A. melas.* From the data review, wild-caught mosquitoes produced the greatest number of live mosquitoes e.g., in a study by Gelfand (1955a) the most effective method for mosquito collection was the wild-catch method.
- *Epidemiology:* In humans, LF was reported in all four regions of Liberia with varying endemicity. The microfilariae prevalence is highest in the south-eastern (1.2% to 37.3%) and northern regions (0.5% to 26.2%), and the lowest prevalence was in the rainforest regions.
- Entomology and Serology data: High prevalence of infections of W. bancrofti in both mosquitoes and human serological data were noted in the coastal region. In a study conducted by Brinkmann (1972), in the Marshall Territory (coastal region), out of the 871 persons tested using night blood smear, a prevalence rate of 12.7% was recorded, while out of 306 mosquitoes dissected the infection prevalence was 27.1%.

3.1.5 Conclusions

Historical documentation on entomological and human population distribution of *W*. *bancrofti*, has clearly established that the vectors and human distribution of filarial in Liberia is prevalent and is the highest in the coastal region of the county. However, the infection may also be high in other regions of Liberia where no study has been undertaken. Therefore, there is a need for investigation in these regions.

3.2 Introduction

LF is one of the oldest and most debilitating of all the NTDs. It is caused by infection from one of the three species of the parasitic filarial nematodes (roundworms): *W. bancrofti, B. malayi* or *B. timori.* The filarial infection is transmitted by either of the following vectors the mosquito species: *Culex, Aedes* and *Anopheles.* Humans are the definitive host. LF affects populations that are the poorest in the world, mostly those living in communities that have poor water supply and sanitation (Perera, Whitehead, Molyneux, Weerasooriya, & Gunatilleke, 2007).

According to WHO (2016a), an estimated 67.88 million people are infected with the disease, approximately 36 million people are disfigured, and 947 million across the world remain at risk of infection with LF. The life cycle of the microfilarial begins with introduction of third stage filarial larvae onto human skin by a mosquito during a blood meal; subsequently, larvae migrate through the bite wound into the lymphatic vessels. Once a person gets infected, the infection inhabits the lymphatic vessels and subcutaneous tissues and eventually leads to their damage, resulting in clinical diseases manifested as hydrocele, lymphoedema and elephantiasis (CDC, 2014).

In Liberia, according to a study conducted by Young (1953) there is a descending infection percentage prevalence moving from the coast to the rainforest exhibited by the three Bioclimatic zones. The coastal zone of the country has a higher percentage prevalence of LF followed by the savanna zone. The rainforest zone has the lowest infection rate. The common mosquito vectors of LF in Liberia are: *A. gambiae* and *A. melas*. These vectors are in abundance in many localities of the country. Other less common mosquito vectors are: *Anopheles funestus; Aedes aegypti; Cynoscion nebulosus;* and *Mansonia africana* (Briscoe, 1948).

Clinical manifestations of advanced forms of LF can be common and include elephantiasis in the lower extremities and scrotum, particularly amongst the adults in the Kru and Grebo Tribes in the counties of the south-eastern coastal region, than amongst those in the savanna and rainforest zones (Zielke & Chlebowsky, 1979).

A 14 year civil conflict in Liberia tore apart the social fabric costing at least 200,000 lives, of which Liberia is gradually recovering. The entire health system in Liberia was severely affected during the civil conflict, which led to a total collapse of a control on the interventions for all diseases including NTDs.

In the context of controlling for LF, there is currently a need to update information in Liberia and also to reassess the involvement of mosquito transmission in Liberia. To this end, I sought to review the historical literature of the epidemiology of LF in humans and the distribution of *W. bancrofti* mosquito populations and also, where appropriate, to conduct fieldwork to collect contemporary information.

3.3 Methods

A systematic review of the published literature before the 1990 Liberian civil war was conducted using four electronic databases: (i) PubMed; (ii) University of Liverpool Library; (iii) NCBI bookshelf; and (iv) WHO Library – Thomas Allen. A total of 1,689 titles and abstracts were identified, 83 of these articles were found appropriate for review, 51 articles were selected and 17 articles were included for thematic analysis.

3.3.1 List of articles selected for thematic analysis. This was done by closely examining common themes and topics that was related to epidemiology and distribution of *W. bancrofti* in vector and human population in Liberia.

| | | | Type of |
|----|---|-----------------------------------|------------------------------------|
| No | Name of study | Researcher | Study |
| 1 | Studies on Bancroftian filariasis in Liberia, West Africa | Zielke & Chlebowsky, 1979 | Human MF Sampling |
| 2 | Epidemiological investigations of Bancroftioan filariasis in the coastal zone of Liberia | Brinkmann, 1977 | Human MF Sampling |
| 3 | Distribution and prevalence of <i>W. bancrofti</i> in various parts of Liberia | Kuhlow & Zielke, 1976 | Human MF Sampling |
| 4 | Microfilariae and trypansomes in a blood survey of Liberia | Young, 1953 | Human MF Sampling |
| 5 | Filariasis bancrofti studies in Liberia | Poindexter, 1950 | Human MF Sampling |
| 6 | Filariasis in Liberia | Burch & Greenville, 1955 | Human MF Sampling |
| 7 | The Anopheline mosquitoes of Liberia, West Africa | Gelfand, 1954 | Vector Sampling |
| 8 | Man-biting mosquitoes in coastal Liberia | Richard Fox, 1958 | Vector Sampling |
| 9 | Dynamics and intensity of <i>W. bancrofti</i> transmission in the savannah forest regions of Liberia | Kuhlow & Zielke, 1978 | Vector Sampling |
| 10 | Studies on the vectors of W. bancrofti in Liberia | Gelfand, 1955a | Vector Sampling |
| 11 | Malaria studies on the firestone rubber plantation in Liberia | Barber, Rice, & Brown, 1932 | Observation with plasmoquine |
| 12 | A. gambiae in relation to malaria and filariasis in coastal Liberia | Richard Fox, 1957 | Vector Sampling |
| 13 | Havard African expedition in Liberia in 1926 | Liberianhistory.org, nd | Human MF Sampling |
| 14 | Mosquitoes of Liberia: a general survey | W. Peters, 1956 | Vector Sampling |
| 15 | Distribution of Bancroftian filariasis in Africa | Hawking, 1957 | Vector Sampling |
| 16 | Microfilariae and Trypansomes found in a blood survey of Liberia | Young, 1953 | Vector Sampling |
| 17 | Field notes on mosquitoes collected in Liberia, West Africa | Briscoe, 1950 | Vector Sampling |

 Table 3.1 The 17 articles selected for the historical review





3.3.2 Mosquito collection methods for the entomology survey

Entomology control plays a major role in eliminating LF; therefore, it is important to understand the behaviour of the vectors to develop tools for vector control. Therefore, the examination of the vector, their species, distribution, abundance and infectivity need to be understood before any initiation of the MDA.

The human landing catch is one of the most accurate ways of mosquito collection in Africa, this is because it mimics the real situation of the mosquitoes wanting to bite the individual; however, there are ethical concerns about the collectors being bitten by the mosquitoes, the labour intensiveness, and possible collection bias. There are alternative mosquito collection methods to overcome these ethical concerns such as the Biogents sentinel traps (Govella, Chaki, Mpangile, & Killeen, 2011). In this historical review, most of the methods for mosquito collection bridge the ethical concerns.

For the entomology survey in this historical review, mosquitoes were gathered by two methods: (i) wild-caught mosquitoes; and (ii) laboratory-reared mosquitoes. The collection of wild-caught mosquitoes involved:

- 1. Supervised two-man team bait collectors (human landing catch).
- 2. Trap made of strong framework, covered with screen outside household.
- 3. Hand collecting on human bait in opened areas.
- 4. Hand collecting of the mosquitoes resting in the local houses each morning.

To augment field observations experimental infections of mosquitoes were also undertaken. Laboratory-reared mosquitoes of different species were fed on infected participants.

The mosquitoes were than dissected to check for infection with W. bancrofti.

3.3.3 Laboratory analysis

All mosquitoes were collected outside the laboratory and were taken to the laboratory. Samples were identified by species (*A. gambiae* and *A. melas*) if possible by the identification of their eggs from the females.

At the time of dissection, each mosquito was teased apart separately, the abdomen was discarded and the head and thorax were dissected in saline using a binocular microscope at 30X and then using a compound microscope at 75X to examine the mosquitoes.

3.3.4 Epidemiological survey (microfilaria)

The 17 articles reviewed did not mention ethical clearance from the MoH, but only stated that permission for the surveys were obtained from the town chief or villages before the teams embarked on their research activities.

The investigators during the period under review identified microfilariae in humans in peripheral blood using microscopy to detect the filarial parasite. Blood preparations varied from thin to thick blood smears stained by the Giemsa method or the Knott method using both the day smear method and the night smear method.

Thick blood smear was made from a finger prick blood sample. The sample was air-dried and stained with Giemsa, washed and dried for 30 minutes, then examined under a microscope. This allows the distinction of the *W. bancrofti* from other filarial species (McMahon, Marshall, Vaughan, & Kolstrup, 1979).

For the Knott's concentration technique: approximately 1 mL of blood is placed in 10 mL of 1.0%–2.0% of formalin or citrate–saponin solution after which the mixture is centrifuged at 500 g for a minute. The sediment is examined under the microscope for *W. bancrofti* (Melrose, Turner, Pisters, & Turner, 2000).

The blood smears in most of the surveys were at times taken from the entire population or subset of the villages and towns. The sampled population age varied across the various studies ranging from babies to adults (from 9 months to >50 years old). Table 10 gives the distribution and prevalence of infection of *W. bancrofti* in human populations in Liberia using both the day and night smears. The blood was examined under microscope after staining the slides with Haematoxylin using the thick smear technique.

Table 3.2: Dissection of A. melas and A. gambiae in Marshall Territory, LiberiaA. melasA. gambiaeA. melas/gambiae_+

| | No. | No. | % | No. | No. | % | No. | No. | % |
|------------|-----------|----------|-----|-----------|----------|------|-----------|----------|------|
| | Dissected | Positive | | Dissected | Positive | | Dissected | Positive | |
| Malaria* | 427 | 6 | 1.4 | 369 | 21 | 5.7 | 1,142 | 27 | 2.4 |
| Filariasis | 306 | 83 | 27. | 262 | 51 | 19.5 | 592 | 132 | 22.3 |
| Total | | 11 | 1 | | 4 | 1.5 | | 11 | 1.9 |
| | | | 3.6 | | | | | | |

Source: Gelfand (1955a, 1955b); * Salivary gland infections only.

3.4 Results

3.4.1 Entomology/mosquitoes survey

A total of 17 articles were reviewed and the data analysed. Six articles were selected based on entomological distribution of LF, while 11 articles were selected based on human distribution of LF. Eight of the articles conducted surveys using night blood smears and the remaining nine articles conducted surveys using day blood smears.

Participants were distributed according to regions and counties. Gender distribution could not be ascertained in some of the studies making it difficult to conduct a total aggregate of all the participants by gender.

Seasonal variations of the vectors were noted where the highest catches of mosquitoes were during the rainy season from April to October, and the lowest catches of mosquitoes were in the dry season from December to March.

The results of the systematic review also showed that the coastal region has the highest percentage of prevalence for *A. melas* and *A. gambiae* mosquitoes, this was demonstrated by Gelfand (1955a, 1955b) in the Marshall Territory (now known as Margibi County) in the coastal region of Liberia for both malaria and filariasis. The survey also showed that *A. melas* (27.0%) was a more effective vector in transmission than *A. gambiae* (20.0%) (see Table 3.3).



Figure 3.2: Seasonal prevalence of A. gambiae

Source: Briscoe (1948).

| Table 3.3: Summary results of various surveys showing distributi | on of | W . |
|--|-------|------------|
| <i>bancrofti</i> in mosquitoes in Liberia | | |

| Year | Region | Location | Proportion (%) | Mosquitoes species | Observer |
|---------------|------------------|-----------------------|-------------------|--------------------------------|---------------------|
| 1947 | South Central | Robertsfield | 1.6 | Anopheles gambiae | Diller (1947) |
| 1950 | South Central | Robertsfield | 0.8 | Anopheles gambiae | Briscoe (1952) |
| 1952– 1955 | South Central | Marshall Territory | 19.5 | Anopheles gambiae | Gelfand (1955a) |
| | | | 27.1 | Anopheles melas | |
| | | | 22.3 | Anopheles melas/ gambiae | |
| | | | 10.0 | Anopheles hancocki | |
| | | | 0.0 | Anopheles hancocki | |
| | | | 0.6 | Culex thallasius | |
| | | | 0.0 | Culex nebulosus | |
| | | | 0.0 | Culex fatigan | 8 |

| | | | 0.0 | Culex | | |
|-------|---------|-----------|------|----------------|-----------|----------|
| | | | | nebulosus | | |
| | | | 0.0 | Culex decens | | |
| | | | | 4.4 Manson | ia | |
| | | | | uniformis | | |
| | | | 4.9 | Mansonia | | |
| | | | | africanus | | |
| | | | 0.0 | Ades africanus | 5 | |
| | | | | Theo | | |
| | | | 3.4 | Ades aegypti | | |
| | | | | 0.0 Ades | | |
| | | | | nigricephalus | | |
| | | | 0.0 | Ades | | |
| | | | | punctothoracis | I. | |
| 1957_ | South | Marshall | 3.6 | Anopheles | Fox | (1957. |
| 1959 | Central | Territory | 210 | aambiaa | 1058 |) |
| 1930 | Central | Terntory | | gambiae | 1750 |) |
| 1976 | North | Bolil | | 48.9 A | Anopheles | |
| | Central | | | gambiae | | |
| | | | 47.9 | Anopheles | | |
| | | | | funestus | | |
| | | | 2.86 | Anopheles nili | | |
| | | | | Ghandu 8 | 37 | |
| | | | | Anophel | es | |
| | | | | gambiae | ••• | |
| | | | 3.1 | Anopheles | | |
| | | | | funestus | | |
| | | | 95.7 | Anopheles nili | | |
| | | | | 1 | | |
| | | | | Grahntown 9 | 02.9 | |
| | | | | Anophel | es | |
| | | | | gambiae | | |
| | | | | Anopheles | | |
| | | | 1.2 | funestus | | |
| | | | | Anopheles | | |
| | | | 74.4 | gambie | | |
| | | Kaikatown | | | _ | |
| | | | | Anopheles | Kuhlov | v & |
| | | | 23.3 | funestus | Zielke | e (1978) |



Figure 3.3: Proportion of wild catch and infected mosquitoes

Source: Gelfand (1955a).

Figure 3.4: Proportion of *W. bancrofti* in mosquitoes, by Bioclimatic zones in five Liberian villages



Source: Gelfand (1955a).

In 1948, Briscoe conducted a study, which revealed that there are various species of mosquitoes found in the coastal and savanna zones of Liberia. *A. melas* and *A. gambiae* are the most abundant mosquitoes documented. Fairly abundant were also *A. funestus, Aedes aegypti, Culex nebulosus, C. mucheti* and *Mansonia Africana*.



Figure 3.5 The prevalence of microfilariae in four counties in the coastal region in Liberia

Source: Kuhlow & Zielke (1976).

3.4.2 Human survey of W. bancrofti in human populations in Liberia

In all the studies where gender was included, men in Liberia had higher prevalence of microfilaria 9.5%–24.0% as compared to their female counterparts 7.4%–19.0% (Zielke & Chlebowsky, 1979). A study carried out by Brinkmann (1972) also noted that men were twice as likely to have the infection compared to women. This increase in percentage was attributed to working in outdoor environments and that men tend to sleep late at night making them more prone to getting bitten by mosquitoes. The highest prevalence of parasites was observed amongst people between the ages of 40 and 49 years.
| Region | Total examined | Prevalence of | Prevalence of | Author, | Technique used | |
|----------------|-------------------|---------------|---------------|------------|---------------------|--|
| | | microfilariae | microfilariae | Year | | |
| | | in females | in males | | | |
| | | (%) | (%) | | | |
| South-Eastern, | 1,136 | 7.4% | 9.5% | Diller | Thick blood film | |
| Coastal | | | | (1947) | biood min | |
| South-Eastern, | 180 | 19.0% | 24% | Gratama | Knott | |
| Coastal | | | | (1966) | method | |
| Coastal | 871 | 4.2% | 8.4% | Brinkmann | Thick | |
| | | | | (1972) | blood film | |
| North Central | 1 1,968 | 14% | 19.0% | Zielke & | z Night | |
| /non-coastal | | | | Chlebowsky | smear | |
| | | | | (1979) | | |

 Table 3.4: Comparison of percentage prevalence of microfilariae in Liberia, by gender

In the human population, the day blood smear revealed that there was active transmission of microfilariae in all four regions of Liberia. The highest percentage of prevalence for microfilariae was seen in the south-eastern region, coastal zone in Harper, Maryland County with a prevalence of 8.0% (Young, 1953), and the lowest percentage of prevalence of 0.3% was seen in the north-western region in the Gola Rainforest, Bomi County. These results correlate well with those by Kuhlow and Zielke (1976) who surveyed four counties in Liberia and found that Maryland County had the highest percentage prevalence of microfilaria (37.3%) compared to the other three counties: Bassa County (37.3%); Sinoe County (14.5%); and Rivercess County (14.0%).

The historical data correlates with the result of a nationwide mapping survey, which was conducted in Liberia by the MoH in collaboration with its partners, to determine the prevalence and epidemiology of LF in the country, the overall immunochromatographic test (ICT) prevalence was 24.0% with the highest percentage prevalence observed in the coastal regions as observed in the historical studies. In 2012, in order to conduct MDA after a period of 14 years of civil war, a total of 3,132 persons were examined between 22:00 p.m. and 02:00 a.m. This result also correlated with that of the historical data that reported the highest prevalence in the coastal regions.



Figure 3.6: Regional distribution of human population of *W. bancrofti* in Liberia per year of study

Source: Diller (1947); Briscoe (1952); Gelfand (1955a); Gratama (1966); Brinkmann (1972).

| Yea r | Regions | Counties | | Prevalence microfilariae | rate (%) | of | Author, Year |
|----------|---------------|------------------------|-------|-----------------------------|-------------|----|----------------------|
| | | | | | | | Strong & Shattuck |
| 1930 | North Central | Bong | | 0.02 | | | (1930) |
| | South Central | Montserrado Roberts | Field | 0.3 | | | |

Table 3.5: Natural infection of *W. bancrofti* in humans in Liberia, by using day smear

| | | /Margibi | 0.8 | |
|--------|-----------------|--------------------|-----|------------|
| | South-Eastern | | | |
| | | Grand Bassa | 5.0 | |
| | | Sinoe – Greenville | | Poindexter |
| 1950 | | & Maryland – Webo | | (1950) |
| | | Tchien/Grand | 5.0 | |
| | | Gedeh | 3.0 | |
| | North- | Gola | | |
| | Western | Forest/Gabrpolu | 0.3 | |
| | North Central | Nimba | 0.3 | |
| | North Central | Nimba | 1.0 | |
| | | Bong | 1.5 | |
| | | Lofa | 0.3 | Young |
| 1953 S | outh-Eastern Ma | ryland | 8.0 | |
| | | Grand Gedeh | 0.0 | (1953) |
| | South Central | Montserrado | 2.0 | |
| | | Margibi Margibi | 3.5 | |
| | | | | |

Table 3.6: Natural infection of W. bancrofti in humans in Liberia, by using night smear

| Year | Regions | County | Prevalence | of Author, Year |
|------|--------------------|-------------|-------------------|-------------------|
| | | | microfilariae (%) | |
| 1930 | North Central | Bong | 1.9% | Strong & Shattuck |
| | | | | (1930) |
| 1947 | South Central | Margibi | 8.8% | Diller (1947) |
| 1950 | South Central | Margibi | 8.2% | Briscoe (1952) |
| | South-Eastern | Grand Gedeh | 2.6% | Young (1953) |
| | North Central | Nimba | 1.3% | |
| 1953 | | Lofa | 1.2% | |
| | Southern-Eastern B | Maryland | 16.7% | Hawking (1957) |
| | North-Western | Grand Cap | be 21.0% | Gratama (1966) |
| 1966 | | Mount | | Poindexter (1950) |

| | | Gbarpolu | 1.0% | |
|------|-----------------|------------------------|--------------|-----------------|
| | | | | |
| | South-Eastern B | Maryland | 8.3% | |
| | | | | |
| | South Central | Montserrado Margibi | 5.9% 6.6% | |
| | | Grand Bassa | 20.0% | |
| | South-Eastern A | Grand Gedeh | 14.0% | |
| | | Sinoe | 15.0% | |
| | North Central | Nimba | 0.8% | |
| | South-Eastern A | Rivercess | 14.5% | Kuhlow & Zielke |
| 1976 | South-Eastern A | Sinoe | 33.1% | (1976) |
| 1770 | South-Eastern B | Maryland | 37.3% | |
| | South Central | Bassa | 13.0% | |
| | | | | |

Table 3.7: Continued and concluded natural infection of *W. bancrofti* in humans in Liberia, by using night smear

| | | | Prevalence % of microfilarae | | | |
|------|-----------------|----------|------------------------------|------------|----------|--|
| | | | Wreboken | 3.9% (153) | | |
| | South-Eastern B | Grand | Jiboken | 0.0% (29) | Hawking, | |
| | | Gedeh | Sureke | 3.0% (200) | 1977 | |
| | | | Gbalake | 2.5% (120) | | |
| 1977 | | | Dwekehn | 1.2 (161) | | |
| 1777 | | | Laryi | 0.0% (58) | | |
| | | | Chebioh | 0.0% (80) | Hawking | |
| | | | Tumbanville | 1.8% (112) | 1977 | |
| | South-Eastern A | Sinoe | Worteh | 13.3% (75) | | |
| | | Maryland | | | | |

| | | | Tuoh | 1.2% (84) | | |
|------|---------------|-------|------------|-------------|------------|---|
| | | | Blewen | | Zielke & | & |
| 1979 | South Central | Bassa | Kun | | Chlebowsky | 7 |
| | | | Balle Wreh | 5.6% (143) | 1979 | |
| | | | | | | |
| | | | | 15.2% (46) | | |
| | | | | 17.0% (47) | | |
| | | | Quizah | 4.8% (104) | | |
| | | | | Men 19.0%; | - | |
| | North Central | Lofa | Upper Lofa | women 10.4% | | |
| | | | | | | |

3.5 Discussion

3.5.1 Vector population

Sir Patrick Manson (1878) was the first who showed evidence that mosquitoes were the primary vectors of *W. bancrofti*. The principal mosquito species that convey the lymphatic filariae of humans are found in five genera: *Anopheles (W. bancrofti, B. malayi* and *B. timori)*; *Aedes (W. bancrofti* and *B. malayi*); *Culex (W. bancrofti)*; *Mansonia (W. bancrofti* and *B. malayi*); and *Ochlerotatus* (Chernin, 1983; Bockarie & Molyneux, 2009).

In sub-Saharan Africa, entomological investigations reported that *A. gambiae* complex are the principal vectors of *W. bancrofti* (Kiszewski, et al., 2004). In rural areas of East Africa bancroftia filariasis is mainly transmitted by *A. gambiae* and *A. funestus* and *Culex pipiens fatigans* is the main vector in urban areas. It has also been documented that effective vector control for malaria has been linked to the control of LF (Bockarie & Molyneux, 2009). In the study, national data from 17 countries on ITN ownership was analysed and the results reported there were some regions with >70.0% ITN ownership and suggested if this trend continued and is monitored, this may impact vector control of vector borne diseases (Kelly-Hope, et al., 2006).

This paper gives up-to-date information on the distribution of LF in the vector and human populations in Liberia. The main vectors for the transmission of *W*. *bancrofti* judging by dissections of wild-caught mosquitoes across the various surveys in Liberia are *A. melas* and *A. gambaie* (Gelfand, 1955a). It has also been reported in Mali that *A. gambiae* and *A. funestus* species are the predominant vectors of LF (Coulibaly, et al., 2006). In Gambia, the vectors responsible for the transmission of LF are *A. arabiensis*, *A. gambaie* and *A. funestus* (Knight, 1980); while in Freetown the LF vectors were identified as *A. costalis* and *A. funestus* (Gbakima & Sahr, 1996). In Ghana *A. gambiae* and *A. funestus* have been reported as LF vectors (Gyapong, et al., 2002).

The principal determining factors of the distribution and abundance of the mosquitoes are altitude, rainfall and humidity (Lardeux & Cheffort, 2001). Liberia, which is mostly flat land, has high humidity and heavy rainfall during the rainy season, which has the right environmental factors for mosquitoes to survive. Seasonal variations of the vectors were noted in this study. *A. gambiae* was seen essentially during the rainy season, while *A. melas* was found mainly during the early dry season in Liberia. *A. melas* was also found to be the predominant species in human dwellings (Peters, 1956).

The highest catch of the mosquitoes was noted during the rainy season from April to October, and the lowest catch was in the dry season from December to May (Briscoe, 1948). It has been observed that climate determines the spatial, seasonal distribution and inter-annual variability of many infectious diseases. Marshall Territory, now known as Margibi County, had the highest percentage prevalence of mosquitoes *A. gambiae* (19.5%) and A. *melas* (27.1%) (Gelfand, 1955a, 1955b). Ribbands (1944a, 1944b) reported that *A. gambaie* was observed to breed in seawater. Entomological studies along the coast of Ghana has also shown endemic foci of LF along the coast of the country (Dunyo, et al., 1996).

The study also observed that the breeding for *A. gambiae* and *A. melas* were mainly in small shaded streams, ditches and rocky pools that were in connection with running water and water containing dead vegetation, wells, swamps, grassy waterholes, slow flowing water temporary pools, borrow-pits, ditches, rock pools, small grounded pools found near houses, old cars and sandy small pools with muddy water fully exposed to sunlight (Peters, 1956). In Ghana, Gyapong (2000) observed that rainfall, humidity, temperature and surface area contributed to disease transmission.

The methodology for vector collection varied across the different studies in Liberia. The mosquitoes were caught using three different methods: (i) some of the investigators set up nightly collection stations where mosquitoes were trapped; (ii) human baited traps; and (iii) early morning catching of mosquitoes from the huts of the local population. In Ghana, an entomological study was carried out in the Upper East region, to investigate the transmission dynamics and intensity of LF. Mosquitoes were collected by indoor spraying of houses in cluster communities. The dissection and processing of these mosquitoes were also approached in various ways. Wild mosquitoes caught were dissected for the presence of filarial larvae that resemble those of *W. bancrofti*, while laboratory-reared mosquitoes of various indigenous species were fed on human volunteers who demonstrated microfilaria of *W. bancrofti* in the peripheral blood film.

These mosquitoes were then dissected to verify the presence or absence of *W*. *bancrofti* larvae. Dissection methods used were not uniform but varied. In some studies, the head, thorax and abdomen of the mosquitoes were dissected separately, while in other studies the dissection of the thorax was not done and other investigators did not mention the parts of the mosquitoes dissected or infected with larvae of *W. bancrofti*. Dissection was performed over a period of time and the various species were not all

present at any one time. In the Ghana study (Appawu, Dadzie, Baffoe-Wilmot, & Wilson, 2001) each mosquitoe was dissected and put on a slide separately, with head thorax and abdomen dissected separately for each mosquitoes. The polymerase chain reaction (PCR) method can also be used to examine *A. gambiae* complex members for specific species identification; this technique provides accurate information about the transmission of *W. bancrofti* (Goodman, Orelus, Roberts, Lammie, & Streit, 2003).

3.5.2 Human population

Before the 1980s, the only parasitological method used to confirm diagnosis of infection with *W. bancrofti* was through the identification of microfilariae in peripheral blood using the thick smear method or Knott's technique (Knott, 1935). Since then, several other tests have been made to confirm the diagnosis of infection with *W. bancrofti*. Examples of such tests are the ICT used to detect the filarial antigen, which are released by the adult *W. bancrofti* worms in humans. ICT has the advantage to be used both during the day and at night (Weil, Lammie, & Weiss, 1997). The urine Enzyme-Linked Immuno-Sorbent Assay (ELISA test) has been used in countries for the purpsoe of verification post-MDA to detect the antifilarial IgG4 antibodies in human urine post-MDA (Takagi, et al., 2019).

The method used to identify microfilariae in humans by the authors of the various studies under review were by peripheral blood; microscopy was used to detect the filarial parasite. Blood preparations vary from thin to thick blood stained by using the Giema or the Knott methods. Both the day smear method and the night smear methods were observed across studies. They also observed that all mirocfilaria found were that of *W. bancrofti* (Young, 1953).

The standard method or gold standard for diagnosing active infection of LF is the identification of microfilariae in blood smears by microscopic examination using the night blood film examination (blood taken between 21:00 p.m. and 01:00 a.m.) (WHO, 2005). In recent times, PCR assay based on the amplification of a highly repeated DNA sequence found in W. bancrofti was developed to address some of the limitations of the traditional diagnostic methods (Fischer, at al., 2007). Wuchereria bancrofti antigen Wb123 is highly specific and sensitive, which aided in the development of its lateral-flow strip immunoassay. Enzyme linked immunosorbent assay (ELISA) tests carried out on patients infected and uninfected with W. bancrofti infections revealed that there was high sensitivity 93% and 97% (ELISA) with specificities of 92% and 96% (strips), respectively. Separation of the patients uninfected with W. bancrofti to those infected with other helminths or filarial infections including strongyloidiasis and onchocerciasis together with those who were free from any parasite infection, the assay specificities ranged between 91% and 100%. Additionally, the geometrical means obtained by ELISA of W. bancrofti-infected patients was established to be elevated as compared to those uninfected with W. bancrofti (P < 0.0001). This implies that W. bancrofti Wb123 protein is highly sensitive and specific to the IgG4 proving that it has the potential as a post-mass drug administration monitoring tool.

The sample population age varied across the different studies ranging from babies to adults (9 months to >50 years old). The participants also varied across the studies ranging between 105 to 10,128 persons per study. Brinkmann in 1977 conducted a survey that covered the four regions of the country. The results revealed a high infection rate of microfalariae in the coastal region. This corroborated with the other three investigators' findings that showed that infection with nocturnal bancrofti occured in the human population in various parts of the country and they were more common in the coastal zone compared to the forest zone (Poindexter, 1950; Young, 1953). However, Kuhlow and Zielke (1976) reported high prevalence amongst the Gissi Tribe in the Lofa County located in the north-western region of Liberia. In Ghana, a baseline LF study revealed high prevalence of microfalariae 1.8%–20.0% in the northern savanna and the southern coastal areas with low prevalence in the middle forest belt (Gyapong, Adjei, Gyapong, & Asamoah, 1996).

Kuhlow and Zielke (1976) conducted surveys in four counties located in the coastal region of Liberia; their results revealed that the prevalence of microfilariae was highest in Maryland (37.3%), Sinoe (33.1%), Rivercess (14.0%) and Grand Bassa (13.0%). This result compared W. bancrofti transmission in three different Bioclimatic zones namely, coastal, savanna and forest zones, and showed that only 5.0% of the 8,072 persons examined from 82 localities in various parts of these zones in the country were positive for W. bancrofti. All these analyses showed an uneven distribution of the prevalence of microfilariae in the coastal zone showing the highest prevalence rate of 9.0% in the forest zone (Kuhlow & Zielke, 1976). Previous authors (Poindexter, 1950; Young, 1953) also reported that infections with nocturnal W. bancrofti occurred in various parts of Liberia were much common along the coastal region compared to the forest and savanna regions with a high prevalence of W. bancrofti 14.2% amongst the Gissi Tribe in the upper Lofa County, a forest region. A national survey for filariasis was conducted in Ghana to determine the prevalence and distribution of W. bancrofti microfilariae, the results showed a high prevalence of the disease and microfalariae in the northern Guinea savanna and the southern coastal savanna, the middle forest belt was somewhat free (Gyanpong, et al., 1996).

Young (1953) examined blood film of 10,128 persons of all ages and found a prevalence of microfilariae of 4.3% by using the night blood smear and *W. bancrofti* was the only species identified. The heaviest foci were again found in the coastal area. The daytime examination was performed mainly on students. In this study, the prevalence of microfilariae in adults at night was 6.0%. This was 15 times higher than the daytime rate of 0.4%. Young also showed that the natural infection of *W. bancrofti* in humans in Liberia using the day smear was seen to be the highest in the south-eastern region in Harper, Maryland County, having the highest daytime prevalence of 8.0%, which indicates active transmission of *W. bancrofti* (Young, 1953). A similar finding was shown in a study conducted in the coastal area of India, West Bengal. Night blood samples of 4,016 participants revealed the microfilariae rate of 9.1% (Chandra, Chatterjee, Das, & Sarkar, 2007). The intensity of the infection could not be ascertained in all of the studies as various methods of identification of *W. bancrofti* were used ranging from the night blood film method to the Knott method.

Overall, the serological data shows that microfalariae prevalence rate is higher in the coastal regions of the country, this correlated with the entomological findings of studies that reported that LF vectors were also highest in the coastal regions.

3.6 Limitations

- 1. The total mosquitoes collected and dissected in the studies could not be verified.
- 2. Lack of standardisation of the methodologies used for blood smear collection.
- 3. The intensity of the infection could not be ascertained in all studies carried out.
- 4. Total aggregate of the gender and population of both humans and mosquitoes was not possible.
- 5. Dissection of mosquitoes to determine the location of infective larvae present in the mosquitoes was not standardised across the studies.

3.7 Study strength

The main strength of this systematic review is that it provides an up-to-date listing of the literature on the distribution of the vector and human populations of *W. bancrofti* in

Liberia. **3.8 Conclusions**

Historical documentation has clearly established the vectors and human distribution of *W. bancrofti* in Liberia. It is probably prevalent in other entomological host and human populations in the country, but these have not yet been recorded. Therefore, further studies on the vectors and human distribution of filariasis needs to be conducted in Liberia to provide recent data of the vectors and human infections caused by *W. bancrofti* species. The serological data showed that the microfilariae prevalence rate is higher in the coastal region of Liberia, which correlated with various entomological studies in the country that also reported that the LF vectors were also highest in the coastal regions.

CHAPTER 4: BASELINE MAPPING FOR LF BY ICT CARDS AND BASELINE MICROFILARIA PRE-MDA IN LIBERIA

4.1 Abstract

4.1.1 Background

A nationwide mapping was undertaken in Liberia in 2010 to determine the prevalence and geographical distribution of LF. In 2012, a baseline microfilaria survey was conducted to determine the prevalence and intensity of the disease in order to perform longitudinal monitoring of the impact of MDA during the intervention phase.

4.1.2 General objective

To map the distribution of LF in Liberia prior to implementation of a mass drug distribution.

4.1.3 Specific objectives:

- To determine the geographical distribution and prevalence of LF.
- To identify the implementation units before the start of MDA and implementation.
- To determine the population at risk for LF in Liberia.

4.1.4 Methods

In 2010, a total of 1,560 persons aged >15 years were sampled in all 15 counties of Liberia using the ICT cards to determine the presence of the filarial antigen. In 2012, a total of 3,132 persons aged >5 years had blood samples taken between 01:00 a.m. and 02:00 a.m. to determine the intensity of the microfilaria in the 11 endemic counties in Liberia.

4.1.5 Results

The overall ICT prevalence was 24.0% with the highest prevalence rate observed in the coastal region in Maryland and Grand Bassa Counties where antigenaemia exceeded 45.0%. Impact of gender and age on the distribution and levels of rates were not analysed for ICT as both parameters were not properly recorded. A total of 3,132

persons were examined between 22:00 p.m. and 02:00 a.m. in 2012 for microfilaria. The overall prevalence rate of microfilaria was 6.2%, with men having a higher prevalence compared to women (8.0% for men and 4.2% for women). A total of 1,498 men were examined for LF morbidity. The survey found a prevalence of 12.8% for hydrocele and 6.3% for lymphoedema, Maryland (7.5%) and Grand Gedeh (3.6%) had the highest prevalence rates for hydrocele. Maryland (3.0%) and Rivercess (2.3%) were observed to have the highest percentage for lymphoedema cases.

4.1.6 Conclusions

The mapping using the ICT cards showed that LF was endemic in 13 out of the 15 counties in Liberia. The survey provided a nationwide epidemiological data on *W*. *bancrofti* infections in Liberia and serves as the bedrock for the establishment of the NTD programmes in Liberia.

4.2 Introduction

In 2000, WHO established the GPELF with two main objectives: (i) to interrupt the transmission of LF; and (ii) to manage the morbidity associated with it and to prevent disability. The World Health Assembly Resolution WHA50.29 called on Member States to work towards the elimination of LF as a public health problem by 2020 (WHO, 1997).

The swelling due to LF is called lymphoedema and can be found on the breasts, legs and genitals of men. The advanced form of lymphoedema is called elephantiasis. The clinical manifestation of elephantiasis is graded as follows. Elephantiasis of the limb is graded from 0 to 3: 0 = Normal; 1 = Loss of contour or lymphoedema; 2 = Thick skin and loss of elasticity; and 3 = Evidence of elephantiasis. Hydrocoele is graded from 0 to 3: 0 = Normal; 1 = Swelling of spermatic chord; 2 = Swelling up to 10 cms in diameter; and 3 = Swelling greater than 10 cms. Scrotal elephantiasis is graded from 0 to 3: 0 = Normal; 1 = Lymphoedema; 2 = Thick skin and loss of elasticity; and 3 = Evidence of elephantiasis, (Gyapong, et al., 1994, cited in Melrose, 2004).

According to WHO (2017b), around 947 million people worldwide are at risk of LF, of which approximately 40 million are incapacitated and disfigured by the disease. Although not fatal, WHO has ranked LF as the world's leading cause of permanent and long-term disability. Cutting the transmission of *W. bancrofti* can be achieved through annual MDA to the entire population at risk for a period of 5 to 6 years with the goal of reaching 65.0%–80.0% coverage yearly and/or vector control, which could lead to the elimination of the disease (WHO, 2011b).

The recommended therapy for cutting the transmission of LF is treatment of the population at risk with Ivermectin and Albendazole in areas where Oncho is coendemic with LF, such as in Liberia, whereas DEC and Albendazole are administered in areas where Oncho is not co-endemic. The regimens for MDA is as follows: annual treatment of the entire population at risk with a single dose of Albendazole (400 mg) and Ivermectin (150–200 mg/Kg) or DEC (6 mg/Kg) for 4 to 6 years or DEC, Albendazole and Ivermectin used for a period of 1 to 2 years as recently recommended by WHO for countries or districts not endemic for Oncho. There was a trial of a triple regimen therapy for LF in Papua New Guinea, which included the use of Ivermectin 200 µg and DEC 6 mg/Kg and Albendazole 400 mg all together given once a year for a period of 3 years. The result showed that there was a greater clearance of microfilaria compared to the two doses of DEC 6 mg/Kg and Albendazole 400 mg (King, et al., 2018).

There are surveys as far back as the 1930s that determine the presence of LF from clinical cases reported in the Marshall Territory (Gelfand, 1955a). However, there

has been no national survey to determine the prevalence of the disease before 2010. The first nationwide baseline mapping for LF by using the ICT in Liberia was conducted in 2010 by the MoH in collaboration with WHO before the establishment of the NTD programme in the country. A prevalence map was developed, which indicated that LF was endemic in 13 of the 15 counties in the country (MoHSW, 2017a). In 2010, all 15 counties in Liberia were mapped. The survey revealed that 13 out of the 15 counties were endemic for LF. Those counties with levels of antigenaemia <1.0% ICT considered non-endemic for LF. The 2010 map guided the implementation of MDA by the LSTM and NTD Programme at the MoH Liberia. This paper presents an evolution of the national programme and looked at potential factors that could influence the prevalence of LF in Liberia.

Liberia has a total population of around 4.5 million when applying the growth rate of 3.2% according to the nationwide census in 2008 (Government of the Republic of Liberia, 2008). This paper will be the first comprehensive description of the geographical distribution of LF throughout Liberia.

| 1947 Microfilariae s | survey conducted in Margibi County |
|----------------------|---------------------------------------|
| using night blo | od on 297 participants, showed a mean |
| microfilaria pre | evalence of 8.8% (Diller, 1947) |

| <u>Table 4.1: Key events rela</u>ted to elimination of LF implementation in Liberia | | | | | | |
|--|--|--|--|--|--|--|
| Year | Event | | | | | |
| 1947 | Microfilariae survey conducted in Margibi County | | | | | |

0 1.

10.10 17

1 - 1

| 1949 | Young and Johnson recorded the finding of A. melas larvae in Monrovia (Young | ð |
|-------|--|---|
| | Johnson, 1949) | |
| 1952- | -1955 Greater than 11,000 A. melas and A. gambiae complex were collected in | |

Margibi County of which 733 were dissected and 27.1% were positive for filariasis (Gelfand, 1955a) 2010 National mapping of LF survey based on the use of ICT

| 2012 | First | baseline | sentinel | site | microfilaria | survey | carried | out | prior | to | MDA |
|------|--|----------|----------|---------|-----------------|-----------|----------|-------|---------|-----|-----|
| | | | ir | npler | nentation | | | | | | |
| 2012 | | | F | irst ro | ound impleme | entation | of MDA | | | | |
| 2013 | 013 Second round implementation of MDA | | | | | | | | | | |
| 2015 | 5 Third round implementation of MDA | | | | | | | | | | |
| 2016 | | | Ν | lid-te | erm sentinel si | ite surve | y implen | nenta | tion af | ter | |
| | | | tł | ree r | ounds of MD | А | | | | | |
| 2017 | | | F | ourth | round implei | mentatio | n of MD | А | | | |
| | | | | | | | | | | | |

4.3 Randomisation method

Survey design: Cross-sectional survey.

Inclusion criteria: Random community sampling of 15 years and above that lived in the district and consented to participate.

Exclusion criteria: <15 years and considered to be sick.

Study period: July to October 2010.

4.3.1 Planning for mapping at national level

The national level planning meeting was held in Monrovia at the MoH. A 1-day training was held for research assistants at the MoH. Pre- and post-test was administered during the training to help access the knowledge of the participants before and after the training exercise. At the end of the training, participants were divided into three groups of five based on their counties of assignment. Each team developed a detailed work plan to enable them to systematically complete the survey. The meeting included surveillance officers, laboratory technicians and community health volunteers from the 15 counties. Included in the training was a WHO representative. After the training, participants were knowledgeable about the LF mapping procedures and demonstrated some skills and

knowledge in carrying out the survey. Surveillance officers were responsible to work with their respective counties and health teams to carry out sensitisation in the targeted communities.

4.3.2 Ethical issues

Ethical approval for the survey was obtained from the research unit of the MoH. Participants of the study gave consent before they were enrolled. Two years after the baseline study MDA was started in all endemic counties. The investigating teams in collaboration with the county health officers in the various counties, obtained verbal as well as written consent from the town chiefs and elders as well as the study participants before the survey commenced.

4.3.3 Field activities

Pre-sensitisation was carried out before the survey, community members were asked to assemble at a central point, preferably a palaver hut or clinic. The details and objectives of the survey were explained. Individuals who consented to be part of the study were invited to a designated place within selected communities.

4.3.4 Data collection

A total of 1,560 persons aged >15 years old was sampled for daytime antigenaemia, using the ICT provided by WHO in Liberia (Weil, et al., 1997). During the survey, 60 μ L of fingerprick blood was collected from each participant and placed on the ICT cards. Results were regarded as negative if after 10 minutes the card indicated a negative result; positive results were usually seen within 5 minutes of beginning the

test.

Based on WHO guidelines, in each of the sampled communities at least 100 people aged >15 years old were sampled. The survey was stopped if one of the first 50 adults were found to be positive since this gives sufficient precision (1/50 = 0.02%) for an implementation unit (IU) to be declared positive. If 100 ICTs were negative in a county this county was considered negative for LF. Assessing the prevalence of LF using ICT cards is easy to use when trained properly because it has been observed that in large surveys, crowd control as well as handling the cards by untrained staff members may read the cards wrongly, which could contribute to misinterpretation of the results and may lead to an inappropriate decision for a district (Ottesen, 2000).

4.3.5 Microfilaria prevalence and density data collection prior to MDA implementation

Survey design: Cross-sectional survey.

Inclusion criteria: Random sampling of people 15 years and above that lived in the district and consented to participate.

Exclusion criteria: <15 years and considered to be sick.

Study period: 2–20 August, 2012.

Microfilaria prevalence and density data was conducted to define LF sentinel sites in Liberia and to collect baseline data on LF microfilaraemia. Sampling was conducted in accordance with WHO guidelines; recommending to have one sentinel site by county and one to two spot check sites or IU was selected. In Liberia, a county is considered an IU (WHO, 2010a). According to WHO, when counties are not highly populated, several IUs could share one sentinel site considering that, at a minimum, there has to be at least one sentinel site and one spot check site per 1,000,000 people (WHO, 2010a). In Liberia, the sizes of the IUs are small, therefore, some units were merged and one sentinel and spot check site represents the merged units. Sites were chosen from areas of known high and low LF transmission, low or high mosquito densities, and high or low antigenaemia prevalence.

Pre-sensitisation was carried out before the survey at each sentinel site. Individuals were asked to assemble at a central point, preferably a health centre, clinic or town hall. All those consenting were tested using ICT cards and those tested positive were further tested for microfilaraemia using night blood collection. However, some individuals who were tested positive with ICTs refused to be tested at night. To ensure the standardisation of activities and data, prior to the study commencing, all technicians were given a 2-day practical training.

According to a study by Weil et al. in 1997 on the comparison of tests for the detection of circulating filarial antigen, prevalence of microfilaraemia before the 1990s was assessed by parasitological methods, which used the microscope to determine filarial infection. The sensitivity of these methods depends on the time of the day the blood was collected, which in Africa, microfilariae circulates in the peripheral blood between midnight and 02:00 a.m. (Knott, 1935) This method is widely used in countries that are endemic for filariasis.

The data collected from the sentinel sites were used by the NTD programme to determine the impact MDA has had on LF.

4.4 Methodology

A total of 3,132 samples were collected and examined using the night blood method. The gender distribution for the participants were as follows. A total of 60 μ L blood sample was taken from fingertips of each participant between midnight and 02:00 a.m. Blood collected was smeared on the slide and allowed to air-dry at room temperature. The following day, the dried smears were dehaemoglobinised by flooding with distilled water for about 5 minutes, air-dried again, fixed with methanol for 30 to 60 seconds, and then stained with Giemsa for 10 minutes before being examined for microfilaria under a light microscope. Positive findings of microfilaria were recorded.

4.5 Results

Table 4.2 provides the summarised results of the distribution of LF in Liberia.

| | Mappi | ng | Baseline mic | rofilaria | Morbidity | | |
|-------------|-------|-----------------|--|---------------------|----------------------------|-----------------------------|--|
| | No. o | f persons | | | | | |
| | using | % prevalence | No. o persons | f % prevalence | | % | |
| positive | | antigen cards | s examined microfilaria microfilaria | of for positives | % Hydrocele positive | Lympho edema positive | |
| Total by | | | | | | | |
| counties | 1,560 | 24.1% | 3,132 | 6.2% | 12.8% | 6.3% | |
| Montserrado | 210 | 16.7% | 300 | 0.3% | 0.5% | 0.0% | |
| Bong | 110 | 2.7% | 300 | 3.0% | 0.6% | 0.0% | |
| Sinoe | 38 | 39.5% | 294 | 20.0% | 0.0% | 0.0% | |
| Rivercess | 84 | 15.5% | 300 | 5.0% | 0.0% | 2.3% | |
| Maryland | 52 | 46.1% | 299 | 21.0% | 7.5% | 3.0% | |
| Grand Bassa | 28 | 46.4% | 300 | 8.0% | 0.0% | 0.0% | |
| Lofa | 162 | 8.0% | 300 | 4.7% | 0.0% | 0.7% | |
| Grand | | | | | | | |
| Gedeh | 128 | 6.3% | 298 | 0.3% | 3.6% | 0.0% | |
| Nimba | 147 | 1.4% | 300 | 0.0% | 0.6% | 0.0% | |
| Grand Cape | | | | | | | |
| Mount | 101 | 2.0% | 290 | 1.7% | 0.0% | 0.3% | |
| River Gee | 119 | 5.0% | 151 | 2.0% | 0.0% | 0.0% | |
| Margabi | 100 | 2.0% | _ | _ | _ | _ | |

Table 4.2: Crude baseline LF prevalence with antigen detection and microfilaria test,by county and sex in Liberia

| Grand Kru | 50 | 25.4% | _ | _ | _ | _ |
|-----------|-----|-------|-------|------|-----------|---|
| Bomi | 104 | 0.0% | _ | - | _ | _ |
| Gbarpolu | 126 | 0.0% | _ | - | _ | _ |
| By sex | | | | | | |
| Male | _ | _ | 1,496 | 8.0% | All males | _ |
| Female | _ | _ | 1,636 | 4.2% | All males | _ |
| | | | | | | |

Source: MoHSW (2017a).

Figure 4.1: Google map showing ICT survey and the "baseline" survey carried out in Voinjama district (2010) and Foya district (2012) in Lofa County



4.5.1 ICT

All 15 counties in Liberia were surveyed using ICT cards. The results revealed that 13 of the 15 counties had a prevelance of ICT positive >1.0%. Figure 22 gives a graphic description of the communities. The overall ICT prevalence was 24.0% with the highest in the south-eastern coastal regions (Maryland and Grand Bassa Counties).





Source: MoHSW (2017a).

4.5.2 Microfilaraemia prevalence

A total of 3,132 blood samples were examined between 22:00 p.m. and 02:00 a.m. of which 1,496 were males (47.8%) and 1,636 were females (52.2%). The results showed an overall prevalence rate of 6.3% with men having a higher prevalence compared to women.

| Table 4.3: Crude data on the distributions of microfilaria prevalence and antigenaemia levels |
|---|
| in Liberia from 1930–2022, by counties and year Baseline results 2012 before first MDA |

| Total by counties | Historical data 1930–1990 (range) | Mapping 2010 by ICT | Baseline by ICT | % prevalence microfilaria |
|----------------------|---|---------------------------|--------------------|---------------------------------|
| Montserrado | 7.6–21.0% | 16.7% | 0.0% | 0.3% |
| Bong | 1.9–3.6% | 2.7% | 3.3% | 3.0% |

| Sinoe | 15.0-33.0% | 39.5% | 19.3% | 20.0% |
|---------------------|------------|-------|-------|-------|
| Rivercess | _ | 15.5% | 3.3% | 5.0% |
| Maryland | 15.3–37.0% | 46.1% | 21.3% | 21.0% |
| Grand Bassa | 13.0-20.0% | 46.4% | 7.3% | 8.0% |
| Lofa | 2.1–15.2% | 8.0% | 4.7% | 4.7% |
| Grand Gedeh | 33.0% | 6.3% | 33.6% | 0.3% |
| Nimba Grand Cape | 0.8–39.0% | 1.4% | 0.0% | 0.0% |
| Mount | 2.0-21.2% | 2.0% | 1.7% | 1.7% |
| River Gee | _ | 5.0% | 0.0% | 2.0% |
| Margabi | 3.8–14.0% | 2.0% | _ | _ |
| Grand Kru | _ | 25.4% | _ | _ |
| Bomi | _ | 0.0% | _ | _ |
| Gbarpolu | _ | 0.0% | _ | _ |

Source: MoHSW (2017a).

A total of 1,498 men were examined for LF morbidity the survey found an overall prevalence of 12.8% for hydrocele and 6.3% for lymphoedema. Maryland (7.5%) and Grand Gedeh (3.6%) Counties had the highest percentage prevalence for hydrocele. Maryland (3.0%) and Rivercess (2.3%) Counties had the highest percentage for lymphoedema.

Table 18 shows an uneven geographical distribution of *W. bancrofti* in Liberia. It also shows significant variations between regions and within regions. All the surveys were cross-sectional surveys.

4.5.4 Comparison of Liberia historical serological and entomological data to LF baseline data prior to MDA implementation in Liberia

The critical analysis of historical data on the distribution of *W. bancrofti* in vectors and human populations in Liberia reported that the south-eastern region in coastal areas had the highest prevalence of *W. bancrofti* vectors and microfilaria (see Chapter 3). This correlates with the nationwide survey performed in 2010 by the MoH, which also reported that indeed the south-eastern coastal region had the highest prevalence of microfilaria (MoHSW, 2017a).

In Liberia, for the purpose of determining the distribution of these diseases, various studies have been conducted. However, the first published report was by Diller in 1947, who reported that the disease was endemic in all Bioclimatic regions of the country and clinical manifestations was also found in the coastal region. Comparisons of results across the various surveys prior to MDA implementation in Liberia have demonstrated that LF is highly prevalent in the southern coastal regions of the country compared to the savanna and rainforest zones. There has also been evidence of repeated infected mosquito bites in the coastal region leading to the development of clinical filariasis in Liberia (Poindexter, 1950).

Various studies across West Africa showed the prevalence of *W. bancrofti* in the coastal regions. In Ghana, a study carried out by Gyapong et al. (2002) showed that the disease has a higher prevalence in the northern savanna areas with a microfilaria prevalence between 20.0%–40.0% and the coastal savanna areas (10.0%–20.0%). In Sierra Leone, a nationwide survey showed that LF was endemic in all 14 districts with *W. bancrofti* antigenaemia prevalence >1.0%, with a higher prevalence in the northeast regions (Bombali 52.0%; Koinadugu 46.0%; Tonkolili 37.0%; and Kono 30.0%) and lower in the south-west regions of the country (Koroma, et al., 2012). Also in the East

African country of Malawi a survey in 2002 showed that *W. bancrofti* infections were widespread in the country. The antigenaemia prevalence using the ICT cards was about 80.0% in some of the sampled villages (Ngwira, Tambala, Perez, Bowie, & Molyneux, 2007). In Kenya, similar studies were carried out by Wijers (2016) to get an insight on the prevalence of the disease in the coastal province. A cluster sample survey revealed 28.4% of the total of 5,004 males examined were found to be microfilaria positive.

4.5.5 Recent epidemiological studies in Liberia

Using the same platform used at the baseline in 2012, the NTD programme in Liberia in December 2016 carried out a survey after three rounds of interruption of MDA due to the Ebola virus. This survey confirms our baseline study carried out in 2012, which showed that microfilaria prevalence was highest in the south-eastern coastal region of the country. In order to determine the level of impact made through the MDA intervention for the control and elimination of LF, a couple of strategies have been instituted by the NTD programme in Liberia, including sentinel and spot check sites to evaluate the impact of the drugs. The sentinel sites are also used throughout the programme to determine the baseline parasitological indicators and to evaluate the changes in the indicators throughout the course of the programme.

MDA impact is ascertained when microfilaria surveys are performed using sentinel and spot checks. Since the implementation of MDA, there have been two rounds of sentinel and spot check sites in the country. The first sentinel and spot check site was conducted in 2012, while the second was conducted in 2016. According to WHO protocol, the first sentinel site survey should be conducted after two to three rounds of MDA (WHO, 2010b). The long period between first and second rounds of sentinel sites was due to the disruption of MDA activities caused by the Ebola outbreak in early 2014 to late 2015.

Sentinel sites require a minimum of 500 individuals, located in an area known to be highly endemic, where the population is stable and ideally with no previous history of Oncho and LF MDA. While spot check sites are used in association with sentinel sites and are selected based on the same criteria, they can change during each survey.

4.5.6 Key objectives of sentinel and spot check sites activities

- To determine the impact of the national LF elimination programme by detecting • changes in LF prevalence and intensity using sentinel and spot check sites.
- To assess when the level of microfilaraemia is less than 1.0% in all sentinel sites ٠ suggesting that transmission has been interrupted and the country is ready to implement a Transmission Assessment Survey (TAS).

A survey to confirm LF transmission in big cities affected by conflict-related rural-urban migration in Sierra Leone and Liberia was conducted in 2014. The results revealed no evidence of active transmission of LF in cities, where internally-displaced persons from rural areas lived for many years during more than 10 years due to conflict (de Souza, et al., 2014).

| Table 4.4: Microf Counties | ilaria prevalence : | <u>at spot check/s</u> Type of site | <u>s, by county in 201</u> 2 ticipants No. teste | | <u>201</u> 2 tested | |
|-------------------------------|---------------------|--|---|-----------------------------|------------------------|--|
| | | Microfilaria prevale | | a per site positive ence | | |
| | | | | | | |
| Grand Gedeh | Spot check | 300 | 1 | 0.3% | ,) | |
| Grand Bassa | Spot check | 300 | 1 | 0.3% |) | |

300

13

4.3%

Spot check

Sinoe

| Total | | 3,300 | 58 | 0.2% |
|------------------|----------|-------|----|------|
| Rivergee | Sentinel | 300 | 4 | 1.3% |
| Montserrado | Sentinel | 300 | 0 | _ |
| Lofa | Sentinel | 300 | 4 | 1.3% |
| Bong | Sentinel | 300 | 1 | 0.3% |
| Nimba | Sentinel | 300 | 0 | _ |
| Grand Cape Mount | Sentinel | 300 | 1 | 0.3% |

Source: NTDs unit MoH sentinel site survey

4.5.7 The role of vector control programmes on LF

The Roll Back Malaria Partnership programme established in 1998 by WHO and funded by its partners the United Nations Children's Fund (UNICEF), the United Nations Development Programme (UNDP) and the World Bank advocates for the use of integrated vector management and utilises the use of multi-programme synergies. However, global financial crisis affected the sustainability of the programme (Mutero, et al., 2015).

A study conducted by Njenga et al. (2011a) in coastal Kenya found that despite some of the villages had missed MDA during 2 years, the microfilariae were maintained in the absence of further MDA by the use of ITNs. The anitgenaemia levels declined form 34.6% in 2002 to 10.8% in 2009. These results were especially apparent when the major vector of *W. bancrofti* in these areas was Anopheles (Njenga, et al., 2011b).

| Table 4.5: Distribution of bednets per county (Roll Back Malaria) | | | | | | | |
|---|-------|-------|-------|-------|-------|--|--|
| Counties Nimba | 2006 | 2007 | 2008 | 2009 | 2010 | | |
| | 27.3% | 32.1% | 35.1% | 36.3% | 48.8% | | |
| Bong | 34.6% | 39.0% | 41.7% | 44.0% | 57.2% | | |
| Bomi | 27.6% | 30.4% | 32.5% | 35.8% | 50.1% | | |
| Rivercess | 22.9% | 26.4% | 28.6% | 30.1% | 43.0% | | |
| Margibi | 22.3% | 25.4% | 27.6% | 29.8% | 42.8% | | |
| Grand Gedeh | 27.4% | 31.6% | 34.2% | 35.0% | 46.9% | | |

| Grand Kru | 21.2% | 24.0% | 25.3% | 25.7% | 36.6% | | |
|--|-------|-------|-------|-------|--------------|--|--|
| Sinoe | 21.2% | 24.8% | 27.0% | 27.9% | 40.2% | | |
| Maryland | 23.6% | 26.3% | 27.5% | 27.9% | 38.7% | | |
| Montserrado | 21.6% | 24.3% | 26.3% | 29.0% | 42.2% | | |
| Grand Cape Mount | 32.5% | 36.2% | 38.4% | 41.3% | 55.9% | | |
| Grand Bassa | 23.0% | 26.3% | 28.5% | 30.2% | 43.1% | | |
| Lofa | 33.8% | 40.0% | 43.1% | 44.2% | 57.0% | | |
| <u>Gbarpolu</u> | 32.3% | 37.2% | 40.0% | 42.0% | <u>55.8%</u> | | |
| Source: President Malaria Initiave (2018). | | | | | | | |

In Liberia, according to the National Drug Service, there has been an upscale in bednets distribution by the MoH and its partners. The three Demographic and Health Surveys carried out in the country in 2007 (30.1%), 2009 (48.6%) and 2013 (59.6%), respectively showed an increase in household ownership of bednets in the country (see Table 20). ITNs could be a confounding variable in the decrease in both the microfilaria and antigenaemia levels in the country. The use of ITNs and improvement in environmental sanitation has proven to be effective in eliminating LF in the Solomon Islands (Webber, 1977). Preventive measures on the use of ITNs seems to be effective, but the cost-effectiveness still needs to be researched.

4.6 Discussion

Mansonella perstans

Mansonella perstans is a vector-borne human filarial nematode, which is transmitted through tiny insects known as biting midges or sucking flies. Geographically, it is widely spread across sub-Saharan Africa as well as parts of America (Central and South). The parasite can cause various morbidities and complications if not well managed. However, despite the prevalence of the parasite there is still lack of definite therapy for its complete elimination. Specifically, in Africa, there is a lack of adequate documented evidence on its transmission, symptoms, diagnosis and prevention. This thus poses a gap, which requires further investigation in order to improve the management of the filarial nematode and for recommending appropriate strategies on its treatment and control. Understanding this is vital in reducing any risks, complications, morbidity or mortality associated with Mansonella perstans. The donations of Ivermectin and Albendazole began since the establishment of GPELF and have been tested and found to be safe and effective in reducing the number of circulating microfilariae in the blood and preventing transmission of the infection (Horton, et al., 2000). Yearly MDA is carried out with at least 65.0% coverage of the total population in endemic areas (WHO, 2013b). The first campaign was in Egypt and Samoa and by the 2009 MDA had covered at least 496 million people at risk, and 37 countries were in the process of completing their fifth MDA. Since the establishment of GPELF in 2000, a total of 6.2 billion treatments have been delivered to greater than 820 million people at risk for at least once. By using the strategy of the GPELF the elimination of LF has occurred in some countries, for example, Korea, Japan, China, Thailand and the Solomon Islands. These countries followed the elimination strategy and have stopped the transmission of the infection permanently (Molyneux, 2003). Timor Leste and Senegal and have now achieved 100.0% geographical coverage bringing these countries on track to achieving elimination. Community sensitisation on the knowledge and perception of the disease must be carried out in order for successful elimination of LF (Cabral, 2017). Two pharmaceutical companies have given donations for the antifilarial drugs Albendazole by GlaxoSmithKline and Ivermectin by Merck and Co.

4.6.1 Baseline prevalence and intensity of infection for ICT

The baseline study on LF in the country has contributed immensely to the knowledge and epidemiology of LF in Liberia it has been the basis for which the NTD programme has been built. It provided the epidemiological information of the distribution, prevalence and intensity of LF, which could be used to build an elimination strategy of LF in the country. The baseline data collected included demographic information, prevalence of the disease and its intensity. The NTD programme in the country used this baseline data for the longitudinal monitoring impact of MDA that has been carried out annually.

There have been different studies on Bancroftian filariasis in Africa that have showed how highly endemic the disease is. The baseline study of the epidemiology assessment of LF was the most comprehensive survey ever documented to have been carried out in Liberia prior to starting MDA implementation. The survey showed variation in and between regions. Due to the population size of the country and implementation of health-related activities 13 out of the 15 counties of Liberia have been declared endemic for LF. The transmission was confirmed to be in all regions and Bioclimatic zones of the country with the south-eastern coastal region (Grand Bassa and Maryland Counties) having the highest prevalence rate of 46.4% and 46.1% respectively). The prevalence ranged from 2.0%–46.4% with an overall prevalence of 24.0%. This confirms the findings of a previous study by Brinkmann (1977) who worked in the coastal zone of Liberia and reported W. bancrofti prevalence ranging from 2.0%-37.0% and had a median microfilariae density between 250 mf/mL and 1,200 mf/mL. All studies in Liberia have reported that the south-eastern coastal region has high transmission of *W. bancrofti*. In Sierra Leone, a nationwide LF mapping study was carried out to obtain baseline data on the prevalence of W. bancrofti infection using ICT for circulating filarial antigen, where the overall LF prevalence was 21.0% (Koroma, et al., 2012).

The baseline results also found high prevalence in the northern savanna regions of Lofa and Bong Counties. Lofa County had ICT prevalence of 8.0% and microfilaria prevalence of 4.7%, Bong County had ICT prevalence of 2.7% and microfilaria prevalence of 3.0%, but Nimba County had negative results for both the ICT and microfilaria prevalences. This shows how disproportionately the disease is distributed in the country. Unevenness in the geographic distribution of the disease in the country has also been reported by Diller (1947), when he conducted a survey in four counties.

The Ghana Health Services in 1999 conducted a nationwide prevalence survey to determine the prevalence and distribution of LF in which they reported that the disease had a higher prevalence in the northern savanna regions of the country and the coastal region and a lower prevalence in the forest region (Gyapong, 2000). These findings correlate to our results obtained during the baseline study, which showed that the coastal and savanna regions have a higher prevalence of LF as compared to the rainforest regions. This could be attributed to the presence of the vector in these regions as compared to the rainforest. A study on the distribution of LF in West Africa reported that the disease foci occurred in the savanna, coastal and some rainforest regions. The low altitude and tropical nature of Liberia could be a contributing factor to the prevalence of the disease. Cano et al., (2014) showed that LF transmission increased with mean maximum temperature and decreased with altitude.

4.6.2 The microfilaraemia

In Liberia in 1947, a microfilaria survey was done in Margibi County and night blood showed that 297 (8.8%) of the participants were positive for the larval worms. A breakdown by age and sex indicated that the infection can be acquired during the early years of life and continues into adulthood. The prevalence of microfilaraemia and antigenaemia increased with age. The circulating filarial antigen is an indicator of adult worm loads (Weil, et al., 1997). The age-related increase in the prevalence of antigenaemia indicates that the infection is age-related. A study of a cohort of children in Haiti established that filarial infections are acquired early in life (Hamlin, 2012). The historical surveys noted that many males with scrotum enlargement, knee and ankle swelling and females with elephantiasis of the breasts were seen (Diller, 1947). The baseline microfilaraemia survey in 2012 in 11 counties showed an overall microfilaria prevalence of 6.2% with a range of 0.3%–21.0%. Maryland a coastal county had the highest prevalence of 21.0%. This is because Maryland the most populated costal county in the southeast of the country, and the communial life style of the people. They are more netted together. Males were twice as positive as females; this may be due to the fact that males sit outside with friends late at night making them exposed to mosquito bites. The prevalence of infection has been stated to be higher in males than females in many studies. It has been hypothesised that gender differences, sex hormones, immune effectors and females in reproductive age have markedly lower levels of infection. The microfilaraemia survey corroborated the results of the mapping survey in 2010.

During the survey in 2012, the presence of the clinical form of the disease was assessed and was found to be hydrocele 0.0%–8.0% and for lymphoedema was 0.0%–4.6%; this is much higher then that of Ghana were the baseline prevalence of the clinical forms were 0.4% for elephantiasis and 0.0%–35.0% for hydroceles (Gyapong, Webber, Morris, & Bennett, 1998). In Liberia, the overt forms of the disease have caused a significant social stigma. People presenting with the clinical signs tend to go further in the forest to live to avoid having contacts with others. Gelfand (1955b) stated in one of his communications that an entire community in Marshall Territory (now known as Margibi County) was deserted because some of the community members showed clinical signs of the disease.

The baseline study also gives an estimate of the population at risk for LF in Liberia. In 2010, the population of the country was 3.9 million, but with a 3.9% annual

82

change, the population has increased to 4.7 million in 2017. Of the 15 counties surveyed and mapped using ICT, 13 had >1.0% circulating filarial antigen (CFA) positive. This is around 80.0% of the country's population at risk. The total population at risk of LF in Liberia is projected to be 4 million.

4.6.3 Impact of vector control on the reduction of microfilaria

Low microfilaria prevalence of *W. bancrofti* was found in Liberia during the baseline study despite 2 years of interrupted MDA due to the Ebola crisis in the country. Studies conducted between 1930 and 1990 have revealed a decline in the endemicity of LF pior to MDA. This could be attributed to vector control through distribution of bednets by the Roll Back Malaria programme through the MoH and the indoor residual spraying (see Figure 24). However, there is a need for studies to be conducted. WHO gave a Position Statement in 2011, which endorsed integrated vector management for the control of LF and malaria (WHO, 2011c).

4.7 Conclusions

In Liberia, 13 counties were endemic for LF although at different levels in terms of burden. Mapping and sentential sites survey conducted respectively in 2012 and 2016 clearly illustrated the foci of LF. The mapping results indicate that Maryland County, which is a coastal county, despite four rounds of MDA, still had the highest LF prevalence. This could be attributed to communites being missed during the MDA due to difficult road conditions. The research gives the baseline data for the country's NTD programme to design and implement MDA and serves as the bedrock for the monitoring and evaluation of LF in Liberia.

4.8 Recommendations

- Additional research on LF to be conducted in Bomi and Gbarpolu Counties, which had less than 1.0% ICT prevalence and were considered non-endemic for LF because the protocol for mapping by WHO may have caused these two counties to have been missed.
- 2. The NTD programme should collaborate more with the malaria control programme to ensure that areas with high transmission rate of LF be given priority during distribution of ITN and residual spraying.
- 3. Those counties in the south-east are given priority during MDA in order not to have them missed, as was the case for Maryland County.

4.9 Study limitations

Some participants who had a CFA positive by the ICT were not willing to give blood or night blood smears owing to fears of undue experimentation.

CHAPTER 5: ASSESSMENT OF KNOWLEDGE AND COMPLIANCE TOWARDS MDA INTERVENTION FOR THE CONTROL OF LF IN BONG COUNTY, LIBERIA: A CROSS-SECTIONAL STUDY

5.1 Abstract

5.1.1 Background

In Liberia, there have been four MDA campaigns for LF interrupted due to the EVD. In the country, annual MDA is carried out using Ivermectin and Albendazole.

5.1.2 Objective

To access the efficiency of drug distribution by the community drug distributors (CDDs) and to assess associated compliance and knowledge of individuals in Bong County after four rounds of MDA.

5.1.3 Methods

A descriptive cross-sectional population KAP survey was conducted using structured questionnaires. The survey was conducted from January to February 2018 in Bong

County.

5.1.4 Results

More than 60.0% of the participants were aware of the disease and more than 50.0% of the participants knew of the common symptoms of the disease. Around 63.0% of the participants were compliant to the MDA programme. Those that were non-compliant were afraid of the side effects. However, only 43.3% of the participants admitted to taking both medications, while 50.0% of the participants knew the mode of transmission of the disease.
5.1.5 Conclusions

The findings from this study have shown that there was some awareness regarding LF in the communities. Majority of the participants had knowledge of the MDA programme. However, there is a great need for pre-MDA campaigns and health education to improve compliance by the community.

5.2 Introduction

The GPELF was established in 2010 by WHO with the goal of eliminating LF by 2020. The GPELF launched a two-pronged strategy for the elimination of the disease. First, by MDA to people living in endemic communities in order to stop the spread of the infection (interrupting transmission). Since the beginning of MDA there are some counties that have been declared free of LF. The second strategy of GPELF is morbidity control to reduce the suffering of those already affected by the disease (WHO, 1997). Five rounds of medications are needed to interrupt transmission. In countries like Liberia where Oncho is also endemic it is recommended that annual MDA using Ivermectin (150–200 μ g/Kg) combined with Albendazole (400 mg) in communities where LF and Oncho are more prevalent be distributed to the entire population at risk, but with the exception of pregnant women, children <90 cms in height and very sick people. According to the 2017 WHO report on LF, about 28.0% of the countries that are endemic for LF are now having post-MDA surveillance to confirm that they have achieved elimination (WHO, 2017c). A randomised controlled study conducted in 2013 by Awadzi et al. (2013) to determine if the combination of Ivermectin with Albendazole was safe, reported that the drug was effective with no serious adverse effects observed. Compliance with Ivermectin and Albendazole medication is cardinal to achieving the elimination of LF as advocated by GPELF. A study conducted by

Yirga, Deribe, Woldemichael, Wondafrash and Kassahun (2010) in south-western Ethiopia, showed four positive predictors that can influence compliance to Ivermectin: (i) high risk perception; (ii) family support; (iii) perceiving that CDDs are doing their work well; and (iv) perceiving that measuring the height is the best way to determine a person's treatment dose. Health education before the MDA helps to improve compliance to the drugs (Nuwaha, Okware, & Ndyomugyenyi, 2005). Figure 25 gives a graphic description of vector control for the control of LF.

5.3 Assessment of coverage

Coverage is defined as the percentage of individuals in a given population that received drug or a drug combination in an intervention area (Albonico, Engels, & Savioli, 2004). WHO also defined coverage as the percentage of all residents of an endemic area who swallow the drugs should at least be \geq 65.0%. If high drug coverage is not attained, untreated individuals could potentially act as reservoirs of transmission, hindering control and elimination efforts (WHO, 2011b). For the assessment of coverage, the LF coverage report 2016–2017 was obtained from the MoH NTDs unit, Liberia (.Due to the EVD outbreak in 2014, the NTD programme was suspended. However, there have been four rounds of MDA, which all reported high coverage. To verify the coverage reports, a cross-sectional survey was carried out from 23 March 2017 to 20 May 2017 by the MoH and its partners. This study was conducted in eight counties by the NTDs unit (one district each). Using population-proportionate sampling, the survey report showed that an overall coverage percentage of 84.0% had taken the medication. This is above the required 65.0% coverage recommended by WHO.

The Community-Directed Treatment with Ivermectin (CDTI) first adopted by the African Programme for Onchocerciasis Control (APOC) was found to be effective.

Community members selected by the community leaders were trained in the distribution of medication (Amazigo, 1999).

| | | | | (| Count of |
|----------------|-------------------|--|--|---|--|
| County | District | Communities (individuals) surveyed | Reported coverage* % (total population) | Verified coverage** % (total population) | communities with <65% verified coverage (total population) |
| Bong | Panta Kpaai | 27 (809) | 84.0 | 44.4 | 17 |
| Grand Bassa | Buchanan | 28 (929) | 84.0 | 29.5 | 24 |
| Grand | | | | | |
| Cape Mount | Garwular | 30 (943) | 82.0 | 71.4 | 11 |
| Grand Gedeh | Konobo | 29 (895) | 81.0 | 57.9 | 12 |
| Lofa | Voinjama | 30 (913) | 88.0 | 94.3 | . 30 |
| Marylan d | Pleebo Sodoken | 30 (895) | 82.0 | 61.8 | 13 |
| Riverces s | Jowen | 24 (823) | 83.0 | 92.5 | 0. |
| Sinoe | Kyanyan | 27 (939) | 83.0 | 84.2 | 3 |
| Total | | 225 (7,146) | | 67.0 | 110 |

Table 5.1: Coverage of LF MDA, by district

Source: MoH NTDs unit 2016–2017 coverage report. *Reported coverage = Mass drug treatment reported by drug distributors during MDA; **Verified coverage = A coverage survey done to verify mass drug treatment by a programme.

5.4 Objectives

To access the efficiency of drug distribution by the CDDs and to assess associated compliance and knowledge of individuals in Bong County post-Ebola MDA. The researcher used structured questionnaires.

5.5 Methodology

5.5.1 Study area

Geographically, Bong County is located in the North Central region of Liberia. It is one of 15 counties. According to the 2008 census, Bong County has a population of 328,919 and measures approximately 3,387 square miles. The 2008 census puts Bong County as the third largest county of Liberia (Government of the Republic of Liberia, 2008). The Bong County has 12 political districts. However, the MoH has also divided Bong County into eight health districts. The Bong County is co-endemic for LF, STH and SCH. The survey was conducted in Zota (18,943 population) and Suakoko (28,277 population) districts.

5.5.2 Design (randomisation)

This survey was a population-based survey, using the EPI 30 x 7 method (WHO, 2008). Participants were selected from 30 clusters in each district where LF is endemic. For each cluster, seven houses were randomly selected. Those interviews were with the heads of the household, spouse and anyone above the age of 15 years. A standardised pre-tested questionnaire was used to collect data on compliance and knowledge of mass drug treatment using Ivermectin and Albendazole. The participants were asked on their KAP towards LF. The respondents were the head of each household their spouses and anyone above the age of 15 years and anyone above the age of 15 years.

Figure 5.1: Interviewing a man with elephantiasis in Suakoko district, Bong County



Note: Written consent was obtained from this participant for his photo to be placed in this thesis (see Appendix D).

5.6 Results

5.6.1 Household survey

The survey was conducted in two districts, Zota and Suakoko. A total of 1,191 study participants were surveyed in 60 different communities within the Zota and Suakoko districts and 420 households were visited. There were 210 households visited in each district (see Table 22). Majority of the participants were in the age group 25–44 years (N =642) (see Table 23).

There were 617 (51.8%) females and 574 (48.2%) males that took part in the study. Liberia is predominantly a Christian nation and 86.6% of the participants were Christian. Around 58.3% of the participants were farmers and only 11.7% of the participants had a high school diploma (see Table 5).

| District | No. of surveyed | clusters | No. households surveyed | of | Surveyed population |
|----------|--------------------|----------|-------------------------------|----|------------------------|
| Suakoko | 30 | | 210 | | 632 |
| Zota | 30 | | 210 | | 559 |
| Total | 60 | | 420 | | 1,191 |

Table 5.2: Surveyed population, by the two districts

| <u> Table 5.3: Survey</u> | ed population, b | y gender and a | age |
|---------------------------|------------------|----------------|-----|
| | | | |

| | Age | | | | | | |
|--------------|-------------|-------------|------------|--------------|--|--|--|
| Sex | 15–24 years | 25–44 years | 45 years + | Total | | | |
| Female | 123 | 329 | 166 | 618 | | | |
| Male | 111 | 313 | 149 | 573 | | | |
| <u>Total</u> | <u>234</u> | <u>642</u> | <u>315</u> | <u>1,191</u> | | | |

Table 5.4: Sociodemographic characteristics

| Tuste et the socionation of the characteristics | | | | |
|---|---------------------------------|-----------|------|--|
| Sociodemographic | c characteristics | N = 1,191 | % | |
| Age | 15–24 years | 234 | 19.6 | |
| | 25–44 years | 642 | 53.9 | |
| | 45 years and above | 315 | 26.4 | |
| Gender | Male | 574 | 48.2 | |
| | Female | 617 | 51.8 | |
| Religion | Christian | 1,031 | 86.6 | |
| | Muslim | 158 | 13.3 | |
| | None | 2 | 0.2 | |
| Educational | Completed high school | 139 | 11.7 | |
| level | Elementary | 301 | 25.3 | |
| | Have never had formal education | 449 | 37.7 | |
| | Junior high | 207 | 17.4 | |
| | Secondary high | 95 | 7.9 | |
| Marital status | Cohabiting | 161 | 13.5 | |

| | Divorced | 47 | 3.9 |
|------------|---------------------------|------------|--------------|
| | Married | 587 | 49.3 |
| | Single | 314 | 26.4 |
| | Widowed | 82 | 6.9 |
| Occupation | Business person Farmer | 197 694 | 16.5 58.3 |
| | Fisherman | 44 | 3.7 |
| | House wife | 50 | 4.2 |
| | Hunter | 76 | 6.3 |
| | Others | 130 | 10.9 |
| | | | |

5.6.2 Assessment of drug compliance in relation to the two districts

Compliance (sometimes referred to as adherence) is a term used to denote a degree to which a client correctly follows advice. In this study it indicates how correctly the individuals followed instructions from drug distributors (Albonico, et al., 2004). For the community to be compliant to the MDA, the CDDs have to be accepted by the community, selected by the community, live within the community and be people of integrity (WHO, 1996). They could also be placed on payrolls to enable them to adequately carry out their duties.

Liberia is endemic for LF and a survey to determine the compliance of the medication is necessary to determine the impact of the NTD programme. Following the fourth round of drug distribution, a post-MDA survey was undertaken in Bong County from January to February 2018, this was done to assess possible factors influencing compliance/non-compliance and potential adverse effects. A total of 1,190 individuals were enrolled in this study.

In this study, 63.6% of the participants admitted to taking the medication during the MDA, 9.3% said they took Albendazole alone, and only 43.3% said they took both medications. The reasons for non-compliance varies, but majority (20.0%) said it was due to itching of skin after taking the medication, while 12.9% said it was due to weakness and dizziness (see Table 5.5). The side effects of MDA are specific to the drug that are given and individual tolerance. This includes, vomiting, neausea, rashes, abdominal pains, dizziness and headheaches.

| Table 5.5: Compliance and non-compliance | | | | |
|--|-----|-------|------|--|
| Compliance | | Ν | % | |
| Did you take the medication? | Yes | 757 | 63.6 | |
| | No | 434 | 36.4 | |
| Did you take Ivermectin alone? | Yes | 110 | 9.2 | |
| | No | 1,081 | 90.8 | |
| Did you take Albendazole alone? | Yes | 111 | 9.3 | |
| | No | 1,081 | 90.8 | |
| Took both of the medication together? | Yes | 516 | 43.3 | |
| | No | 675 | 56.7 | |
| Reasons for non-compliance | | N | % | |
| Weakness and dizziness | | 154 | 12.9 | |
| Impotency | | 1 | 0.1 | |
| Abdominal pain | | 80 | 6.7 | |
| Diarrhoea | | 42 | 3.5 | |
| Blood in faeces | | 5 | 0.4 | |
| Sweating | | 32 | 2.7 | |
| Nausea | | 13 | 1.1 | |
| Vomiting | | 100 | 8.4 | |
| Itching of skin | | 238 | 20.0 | |
| Fever and chills | | 58 | 4.9 | |

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5.6.3 KAP towards intervention of LF

A total of 1,191 participants participated in the survey. The study found that 69.3% had knowledge of the disease and 71.5% had known about MDA (see Table 5.6).

| | | Ν | % | | |
|------------------------------|------------------------------|----------|----------|----|------|
| Knowledge of LF | Yes | 825 | 69.3 | | |
| - | | 366 | | No | 30.7 |
| Total | | 1,191 | | | |
| | - | | | N | % |
| Knowledge of how the | Through drinking dirty water | 21 | 1.8 | | |
| disease is acquired | Through traditional rituals | 22 | 1.8 | | |
| | Through mosquito bites | 18 | 1.5 | | |
| | Through farming | 8 | 0.7 | | |
| | Others | 1 | 0.1 | | |
| | | N | % | | |
| | - | | | | |
| Knowledge of clinical | Enlarged legs | 623 | 52.3 | | |
| manifestation of the disease | Enlarged scrotum | 108 | 9.1 | | |
| | Enlarged breast | 75 | 6.3 | | |
| | Fever and chills | 38 | 3.2 | | |
| | Mumps | 131 | 11.0 | | |
| | | N | <u>%</u> | | |
| Knowledge of prevention | - By taking medication | 138 | 11.6 | | |
| | By drinking clean water | 38 | 3.2 | | |
| | By using mosquito nets | 11 | 0.9 | | |
| | By spraying your house | 14 | 1.2 | | |
| | Others | 4 | 0.3 | | |
| MDA programme | | <u>N</u> | <u>%</u> | | |
| Do you know about MDA? | Yes | 851 | 71.5 | | |
| | No | 293 | 24.6 | | |
| | Don't Know | 47 | 3.9 | | |
| | | <u>N</u> | <u>%</u> | | |

Table 5.6: KAP

| How many times was the | One | 436 | 36.6 |
|--------------------------|------------|------------|-------------|
| drug distributed in your | Two | 103 | 8.6 |
| community? | Three | 132 | 11.1 |
| | Four | 111 | 9.3 |
| | Don't know | <u>409</u> | <u>34.3</u> |
| | | | |

5.7 Discussion

There is overwhelming evidence that for chemotherapy-based control strategies such as that implemented for LF to be successful, adherence to treatment is cardinal for ensuring and sustaining good coverage. In this study, only 63.6% of participants admitted to taking the medication during MDA. Whereas 12.9% said that they did not take the medication because they experienced weakness or dizziness after taking the medication and 20.0% said that they experienced itching of the skin once they consumed the medication. Results from a similar study in Haiti showed that 74.0% of the respondents said they were given medication during MDA but only 71.0% swallowed the tablet (Eberhard, 1996). The results obtained from this study will be communicated to the MoH on the importance of pre-MDA educational campaigns in Liberia. If the community is aware of the risk factors associated with the disease, there will be a higher compliance rate and that in the upcoming MDA the NTD unit should have strategies, which will help to improve compliance and sustain the success of the programme.

For MDA to be successful there has to be pre-MDA sensitisation to help to alleviate the fears of community members who believe that the drugs cause adverse effects. There have been similar studies in India, which reported that reasons for noncompliance to the implementation ranges from fear of side effects to lack of perceived benefits (Babu & Mishra, 2008). In this study, 71.5% of participants had knowledge about the MDA before the distribution of the drugs, which indicates that the

awareness of the implementation of MDA was adequate. However, a similar study carried out in Lofa County in Liberia reported 89.0% of respondents knew the purpose of the MDA (Bogus, et al., 2016). A KAP survey carried out in India on the predicators of compliance of MDA reported that in one community 97.1% of the respondents knew about MDA in advance, while in another community only 69.1% of the respondents knew about MDA (Cantey, Rout, Rao, Williamson, & Fox, 2010). For successful implementation of MDA it is very important that the community is aware of the purpose of the MDA as there are many health-related activities ongoing in the various communities, which makes it difficult for them to differentiate. Health education should be carried out and the risk and benefits of the drug be properly explained to the community (Talbot, et al., 2008). Of the total 1,191 respondents, 617 (51.8%) were females and 574 (48.2%) were males. This can be attributed to the fact that in Liberia women mostly stay at home or do farming compared to men who go on hunting expeditions. The result in this study correlates to a similar study carried out in PortauPrince, Haiti, which reported that 70.0% of the respondents in their study were women and 88.0% of their respondents said information about MDA was disseminated to them before the process had started (Beau de Rochars, et al., 2004).

In our survey, the majority of participants indicated that LF is obtained from drinking dirty water and through traditional rituals and not transmitted by mosquitoes. This is in agreement with the findings of a study in West Bengal, India, which reported that only 13.9% of the respondents did not know the mode of transmission of LF and few had incorrect knowledge that direct contact, water and air are modes of transmission (Karmakar, et al., 2011). In this study 52.3% of participants knew that enlarged legs was a symptom of LF as well as enlarged scrotum, where some of the participants thought that mumps was a clinical manifestation of LF. This result

corresponds to a study in Malaysia, which reported that majority of the participants responded that the common symptom of LF was leg swelling (Nazeh, et al., 2014).

5.8 Conclusions

The findings from this study have showed that there was some awareness regarding LF in the communities. Majority of the participants had knowledge about the MDA programme. However, the study also revealed that those participants who did not take the medication did so because of the perceived side effects. This shows that the information of the disease has not been adequately conveyed to the community, making some of the people in the community to be non-compliant. Pre-MDA campaigns by the health promotion unit in collaboration with the NTD unit at the MoH will improve the rate of compliance and hence, programme success.

CHAPTER 6: AN EPIDEMIOLOGICAL SURVEY ON UROGENITAL AND INTESTINAL SCH AMONGST SCHOOL-AGE CHILDREN IN BONG COUNTY, LIBERIA, WITH OBSERVATIONS ON STH

6.1 Abstract

6.1.1 Background

Urogenital (*S. haematobium*) and intestinal (*S. mansoni*) SCH are serious public health problems in sub-Saharan Africa and school-age children are often at high risk for acquiring STH, i.e., *Ascaris*, *T. trichiura*, hookworms and STH. SCH infection is typically acquired by playing and bathing in cercarial infested water while STH is usually acquired ingesting eggs of *Ascaris* or *T. trichiura* or by walking barefoot on soil that is contaminated with hookworm larvae infections. Helminth infections are most obvious in school-age and pre-school-age children. An epidemiological followup study was performed to assess the prevalence and intensity of SCH and STH amongst school-age children in Liberia after MDA using Bong County as an exemplar.

6.1.2 Methods

A cross-sectional parasitological study was conducted from September to October 2017. A total of 1,003 school-aged children (5–15 years of age) were recruited from 10 randomly selected schools in Bong County. The school list was a representative of high and low infection prevalence in the Bong County. The listing of the schools was obtained from the Ministry of Education. Participants were registered on a prestructured and tested questionnaire and were given two labelled containers with their name and identification number (one for the stool sample and the other for the urine sample). Stool samples were collected and processed by the Kato-Katz method for STH and *S*.

mansoni. To determine the prevalence of *S. haematobium* urine specimens were collected and processed using the 10 mL urine filtration method.

6.1.3 Results

Of the total 1,003 school-aged children sampled, 504 (50.2%) were female and 499 (49.8%) were male. The results from the study showed that 123 (12.3%) schoolchildren were infected with *S. mansoni* and 112 (11.2%) schoolchildren were infected with *S. haematobium*. The results of the STH survey revealed a low prevalence for the disease with hookworm at 3.3% and *Ascaris* at 1.2%.

6.1.4 Conclusions

This study clearly showed that STH, *S. mansoni* and S. *haematobium* are endemic in Bong County and are a public health problem in Liberia. There is a need for implementation of MDA, continuous health education, and mobilisation and sensitisation activities, at both district and community levels. The low prevalence of STH in Bong County could be attributed to the annual national deworming programme in the country.

6.2 Introduction

STH, SCH, LF and Oncho are the most common NTDs in sub-Saharan Africa and are major public health problems (Molyneux, et al., 2005). SCH, or Bilharziasis, is a disease caused by parasitic trematode flatworms of the genus *Schistosoma*. There are more than 200 million people infected globally with the disease, with many more at risk of being infected, and 192 million cases of SCH occur in sub-Saharan Africa. The highest cases are found in Nigeria, Tanzania, Democratic Republic of Congo and Ghana

(Steinmann, et al., 2006). WHO has declared SCH as a public health disease, and an NTD targeted for elimination by 2020 (WHO, 2012b). However, there are some countries that still have ongoing transmission (Fenwick, 2017).

Studies on the epidemiology of SCH were carried out at the LIBR from the 1970s to 1989. Many water bodies (streams, ponds and rice paddies) in Lofa, Bong and Nimba Counties examined during the period were found to be harbouring molluscs (*Biomphalaria* and *Bulinus* species), which are the intermediate hosts of human schistosome. As a result of these findings, an SCH Surveillance Unit was established in each of the affected counties and was set up jointly by the World Bank and the Government of Liberia Development Projects. The SCH Surveillance Unit collected baseline data on the prevalence of SCH and other helminthic infections, amongst other activities, in these affected counties also known to be agriculture project areas, as it has been reported that SCH prevalence increases as a result of irrigation and agriculture projects (MoHSW, 2017a). Results from these studies showed that the prevalence of *S*. *mansoni* was significantly higher than *S. haematobium* (Mangal, Paterson, & Fenton, 2008).

People living close to water bodies, lakes, rivers, reservoirs and ponds that are infected with snails, which is the intermediate host, are at high risk of contracting the disease. These are some of the challenges, which in the future may cause the elimination of this disease to be unattainable by 2020 (Fenwick, 2006). After the civil war and to restart interest in disease control the Liberian MoH in collaboration with LSTM and APOC in 2010 conducted a study in all 15 counties consisting of a sample size of 3,144 schoolchildren who provided both stool and urine samples. The mean age of the schoolchildren examined was 10.7 years, with an age range between 7 and 14 years. The prevalence of *S. mansoni* and *S. haematobium* were 9.0% and 6.0% respectively

(MoHSW, 2017a).

There are five species of *Schistosoma* that can cause disease in humans of which there are three major species, which cause morbidity in humans namely, *S. haematobium* found in Africa and the Middle East, *S. mansoni* found in Africa and South America, and *S. japonicum* found in China and East Asia. The other two species are *S. mekongi* found in Laos and Cambodia and *Schistosoma intercalatum* found in West and Central Africa. Both *S. japonicum* and *S. mekongi* are mainly zoonotic species. In some countries, mixed infection has been observed with *S. mansoni* and *S. haematobium*. SCH may cause approximately 200 million deaths annually (Chitsulo, et al., 2000). Pre-school and school-age children are at high risk of infection, where infection is caused by bathing in freshwater, lakes, ponds and rivers that are contaminated with cercarial larvae. It has been observed amongst children living in endemic areas that the burden of the worms progressively increase and climax during adolescence (Gryseels, at al., 2006).

SCH can cause a wide range of symptoms and consequences depending on the species, the worm burden and the length of time infected. Individuals may have moderate to mild parasitic load and limited morbidity, while heavy infection with the disease occurs in fewer populations (Tukahebwa, et al., 2013). It has been reported in many studies that malaria and SCH are co-endemic in many regions and that developing hepatocellular carcinoma may be potentiated in people with co-infection (Yosry, 2006). Infection with *Schistosoma* is not evenly distributed within communities, which may be attributed to distribution of infected aquatic snails, acquired resistance, age-related and differences in exposure patterns. Each schistosome species requires a specific freshwater snail species *S. mansoni* (*Biomphalaria*), *S. haematobium* and *S. intercalatum* (*Bulinus*), *S. japonicum* (*Oncomelania*) and *S. mekongi* (*Tricula*) (Clements, et al., 2006). The current control strategy recommended by WHO for SCH

and STH is by MDA with Praziquantel and Albendazole (Praziquantel 40 mg/Kg body weight + Albendazole 400 mg), targeting mainly school-age children and adults at high risk of infection. Pre-school-age children and infants are also being considered for inclusion in the target population (WHO, 2010d).

STH, commonly known as intestinal worm infection, is a public health problem in the tropics. They are the most common infections affecting the most deprived communities in sub-Saharan Africa (Hotez, et al., 2009). STH are transmitted due to exposure to contaminated soils containing larvae e.g., hookworm, or by ingestion by eating foods that are contaminated with eggs of *T. trichiura* and *Ascaris* (WHO, 2012b). The warm climate is essential for the STH eggs to thrive. Risk factors for developing STH are poor hygiene, poor sanitation and poverty (Vandemark, Jia, & Zhou, 2010).

Of the 7 billion population of the world it is estimated that between 807–1,121 million are affected with *Ascaris*, 604–795 million are infected with *T. trichiura* and 567–740 million are infected with hookworms. School-age children in particular are mainly affected with STH. It has been reported that approximately 89 million of the 181 million school-age children in sub-Saharan Africa are affected with one or more of the STH infections. About 135,000 deaths of STH occur annually (Brooker, Clements, & Bundy, 2006). The three main STH infections are: *Ascaris lumbricoides* (roundworms); *T. trichiura* (whipworms); and hookworm (*Ancylostoma duodenale* and *Necator americanus*). Common clinical conditions of STH can be anaemia, poor nutrition, stunted growth, diminished cognitive development and decreased physical fitness (Bethony, et al., 2006; Stothard, et al., 2009a).

Morbidity control of STH is primarily achieved through MDA administering a single tablet of Albendazole 400 mg or Mebendazole 500 mg. This control strategy has been used for decades and is still the forefront intervention for STH control alongside

improvements in food, water, sanitation and hygiene. The addition of Mebendazole to vitamin A supplementation expanded the treatment to children aged 12–59 months with high coverage. The control of STH is also integrated into other programmes such as nutrition, school health and often parts of SCH control programmes or LF control programmes.

In Liberia, following the civil war and collapse of health infrastructures, there are not many studies conducted to assess STH prevalence and intensity. However anecdotal data from MoH clinics and hospitals indicate that Ascaris, T. trichiura and hookworm are the widely distributed STHs in Liberia (MoHSW, 2011a). However, in 2010 a study by the MoH and its partners reported that of the 3,144 schoolchildren from 59 schools around the county, the main STH species were Ascaris followed by hookworm, which also ranged from 0.0% to >50.0% per school. Liberia has now established a nationwide co-ordinated deworming programme due to the estimated high prevalence of the disease. UNICEF and the MoH in collaboration with the School Health Division of the Ministry of Education have targeted children aged 2 to 5 years in all 15 counties for deworming. It is believed that 65.0% of children under 5 years and pregnant women in rural areas in Liberia are anaemic due to Ascaris, T. trichiura, and hookworm (MoHSW, 2011a). Deworming also takes place when the LF drugs are distributed. To remedy this weakness, this chapter describes a survey that was conducted in Bong County to provide up-to-date information on SCH and STH infections.

6.3 Objectives

6.3.1 General objective

• To provide up-to-date information on the distribution, prevalence and intensity of SCH and STH in Bong County using an integrated surveillance approach with stool and urine sampling.

6.3.2 Specific objectives

- To determine demographic and clinical information from each of the schoolage children that was investigated on SCH and STH.
- To determine the prevalence and infection intensity of SCH (*S. haematobium* and *S. mansoni*) and STH (*Ascaris*, *T. trichiura* and hookworm) in Bong
 County.
- To create an integrated map for SCH and STH across sampled schools.

6.4 Methodology

6.4.1 Ethical considerations

The study protocol received ethical approval from the University of Liberia – Pacific Institute for Research and Evaluation Institutional Review Board at the MoH and the Ethics Committee of the LSTM. Approval was also received from the County Health Team and Ministry of Education in Liberia. The method and purpose of the study was explained to selected school authorities.

Prior to conducting demography and obtaining informed consent, the research team held repeated community meetings in all of the selected schools to communicate the purpose of the study and to answer questions at individual and community levels. The meetings highlighted: participant selection and participation; procedures and protocol; duration of the study side effects; risks and benefits of the study; confidentiality and sharing the results; the right to refuse or withdraw from the study; and who to contact in case of any event. School heads were asked to sign written consent forms on behalf of schoolchildren targeted to participate in the study. The procedure was then explained to each schoolchild, who met the inclusion criteria, and asked to consent verbally to participating in the study. Amongst the schoolchildren that met the inclusion criteria, only those who provided consent were registered and requested to provide samples. Verbal consent was documented by recording the name of each schoolchild who provided consent. Paper forms were stored for the duration of the study plus 3 years per Institutional Review Board protocol for primary data storage. Demographic and morbidity forms included the following details: the participant's name; a unique study identification number (USIN); date of the sample collected; date of birth; sex; family relationships; and place of residence.

6.4.2 Team composition and training

The research team comprised of five members: a principal investigator; one microscopist; one datarecorder; one specimen collector; and one smearer. A 1-day orientation was held in Bong County for the research team. The principal investigator was Dr Louise Kpoto, the datarecorder was a school principal/teacher, and the microscopist, specimen collector and smearer were all county laboratory technicians.

The orientation was aimed at updating the team on the: research protocol; administration of informed consents; handling and transportation of samples to The C B Dumbar Laboratory for microscopy; parasitological examinations (Kato-Katz and the urine filtration methods); completion of data forms; and the manipulation of the GPS (Garmin eTrex 20) for collecting geographical co-ordinates. The GPS was truned on to determine our locations using satellites signals. Pre-test was carried out after the team training and before the project began. The pre-test was aimed at standardising the specimen collection. The day was spent on evaluating the consistency of egg counting amongst the laboratory technicians. A simple method consists of preparing 10 slides and comparing the reading of each slide by each laboratory technician with that of the quality control officer. A discrepancy of 5.0%–10.0% for egg per slide count was considered normal, but a larger discrepancy led to the test not being considered valid, and reasons were identified and corrected. Accurate egg per slide count is particularly important for the Kato-Katz method for intensity assessment. Each school was investigated for 2 days and then the team moved on to the next school.

6.4.3 Selection of study population

The method used to select primary schools from the sampling frame was by stratified random sampling and involved:

- The sampling frame consisted of a list, obtained from the MoH, on all primary schools in the Liberian districts classified as high or medium risk for *S*. *haematobium* and *S. mansoni*.
- Ten schools were randomly selected from a sample frame, which included schools co-endemic for *S. haematobium, S. mansoni* and STH.
- The sampling frame included: the name of each school; the district in which each school was located; the identification code of each school; the location of each school (geographical co-ordinates); the total number of students in each school; and whether the school was a high- or medium-risk area for *S. haematobium* and *S. mansoni*.

6.4.4 Inclusion and exclusion criteria

The inclusion criteria involved: the willingness of the school to participate and provide informed consent; the school being a resident in the Bong County, North Central region of Liberia; schoolchildren aged 5–15 years; and the school located in an area at high or medium risk of SCH and STH.

Schools who were unable to provide informed consent or refused to participate were excluded from the study (Figure 6.1).

Figure 6.1:Criteria for exclusion to participate



6.4.5 Sample size calculation

Based on the inclusion criteria of schoolchildren aged 5–15 years, the average number meeting this criterion was 100 schoolchildren. Using the Creative Research Systems sample size calculator for a confidence level of 95.0% with a confidence interval of 5, from the population of 134 schoolchildren meeting the criteria per school, the calculated sample size was 100 schoolchildren per school and 1,000 schoolchildren for the 10 schools.

6.4.6 Data collection for SCH and STH

Upon arrival to the selected schools, the schoolchildren that met the selection criteria were registered on the child personal form by the recorder and were each given two labelled containers, which contained their name, their USIN and their class. Each schoolchild was explained, which cup was for the stool sample and, which cup was for the urine sample. When the schoolchild returned the specimen cups, the recorder for quality control, checked that the labels on the cups were correct and the specimens properly collected.

6.5 Survey methods

To determine the prevalence and intensity of *S. mansoni* and STH infection, the KatoKatz method was used. To determine the presence and intensity of urinary SCH, the 10 mL urine filtration method was used. The detection of haematuria was also carried out on the schoolchildren by using the hemastix method.

6.5.1 The Kato-Katz method

In the Kato-Katz method for diagnosing *S. mansoni*, the general principle is that people infected with intestinal schistosomes pass the eggs of the worms with their faeces. By examining a stool specimen under a microscope, it is possible to count the number and the type of eggs that are present (see Figure 28).

Figure 6.2: Using the Kato-Katz method for the diagnosis of S. mansoni



6.5.1.1 Safety precautions

- The stool should be considered potentially infectious.
- Wear gloves and laboratory coats whenever handling stool samples.
- Benches, instruments and equipment should be routinely decontaminated with disinfectants after use.
- Materials contaminated with infectious waste should be disinfected before disposal.
- Drinking or eating during laboratory procedures is prohibited.
- Appropriate disinfectant(s) should be used for disposal of contaminated materials, wooden spatulas and specimen containers, and for cleaning of workbenches.
- Used specimen containers must be disinfected before washing. 6.5.1.2 Equipment for the Kato-

Katz method

Kato-Katz equipment involved:

- Stool sample in container (polythene squares tied with grass or plastic pot).
- Microscopic glass slides.
- Cellophane sheets (hydrophilic, 30–50 µm thick).
- Malachite green (or methylene blue).

- Glycerol.
- Metal sieve (Endecott Sieve) with 200–250 µm mesh size.
- Slide boxes.
- Newspapers.
- Wooden or plastic applicators.
- Forceps.
- Kato-Katz plastic template with a hole of 6 mm on a 1.5 mm thick template (delivering 41.7 mg of faeces).

Microscopic examination equipment involved:

- Microscope.
- Hand tally counter.
- Laboratory forms.

Disinfectants and waste disposal equipment involved:

- Disinfectant wipes.
- Medicated soap.
- Methylated spirit.
- Waste container (containing disinfectant).

6.5.1.3 Preparation steps of Kato-Katz reagents

Below are the five steps involved in the Kato-Katz reagents.

Step 1: Weigh out 3 g of Malachite green powder (or methylene blue).

Step 2: Dilute it in 100 mL of distilled water (this is the 'stock solution').

Step 3: Dilute 60 mL of glycerine in 40 mL of distilled water.Step 4: Take 1 mL of Malachite green (or methylene blue) stock solution and add it to 100

mL of the 60.0% glycerol solution (this is the 'working solution').

Step 5: Cut cellophane into 25 mm x 30 mm pieces and soak them overnight in the working solution.

6.5.1.4 Before preparation

- Use clean containers for stool collection.
- Ensure the reusable sieves, templates and spatulas are well cleaned and dry.
- Clearly and correctly label the stool containers to match the participant USIN.

6.5.1.5 Kato-Katz steps

Step 1: Place two glass slides alongside each other and label both slides with the sample number and then place a plastic template on top of each.

Step 2: Place a small amount of the faecal specimen on a newspaper and press through the metal sieve. Using a spatula, scrape the sieved faecal material through the sieve so that only the debris remains on top.

Step 3: Scrape up some of the sieved faeces from the underside to fill the hole in the templates, avoiding air bubbles and levelling the faeces off to remove any excess. Step4: Carefully lift off the templates and place it in a bucket of water mixed with concentrated detergent so that they can be reused.

Step 5: Place one piece of the cellophane, which has been soaked overnight in the Malachite green (or methylene blue) working solution, over the faecal specimen. Step6: Place a clean slide over the top and press it evenly downwards to spread the faeces in a circle (this can be done by inverting the slide onto clean newspaper and pressing firmly). If done well, it should be possible to read the newspaper print through the stool smear.

Step 7: The ideal time for observing *S. mansoni* eggs is 24 hours after preparation, however, in bright sunlight the slides clear rapidly and a 24 hour delay is not necessary.

6.5.1.6 Microscopic examination for S. mansoni

Step 1: After 10 minutes place a little amount of eosin on the slide and place it under microscope using x 10 objective.

Step 2: Count ALL eggs present using a hand tally counter; start in one corner of the sample and systematically scan the whole sample in a 'zig zag' scheme.

Step 3: Record the number and the type of each egg on a recording form alongside the sample number. If no eggs are seen, record '0'.

Step 4: Each sample should be examined by two technicians, one technician reading slide A and the other technician reading slide B.

Step 5: 10.0% of slides A and B should be randomly selected and re-examined by a more experienced technician.

Discrepancy in egg count should not be greater than 10.0%. If discrepancy between readers is greater than 25.0% all slides should be re-read.

The participants were placed in three categories (light, moderate or heavy) based on the intensity of the infection.

For S. mansoni:

Light (1–100 eggs per gram).

Moderate (101–400 eggs per gram).

Heavy (>400 eggs per gram).

For S. haematobium, intensity of the egg per 10 mL of urine:

Light (1–50 eggs per 10 mL).

Moderate (51–499 eggs per 10 mL).

Heavy (>400 eggs per 10 mL). **Step 6:** Once examination of the slides is completed, including quality control, remove

the faeces and cellophane using a tissue into the waste container. Place all slides used when conducting the Kato-Katz method into the disinfectant. These slides should be cleaned and used again for the survey.

6.5.2 Hemastix method

In diagnosing *S. haematobium*, all manufactured kits come with instructions on how to use them. It is very important to follow the instructions to ensure the quality of the results.

6.5.2.1 Equipment for Hemastix test

- Case record form.
- Hemastix test strip and Hemastix pot with scale.
- Scissors.
- Gloves.
- Disinfectants and waste disposal.

6.4.2.2 Steps for reagent strips

Step 1: Collect a fresh urine specimen in a clean plastic container. Ensure that the urine is tested in the field within 2 hours of collection. If there is a delay, refrigerate the specimen if possible.

Step 2: Remove one strip from its bottle (you can cut the strip in two to save resources) and label the strips with the patient identification.

Step 3: Completely immerse the reagent areas of the strip into the urine specimen for a few seconds.

Step 4: When removing the strip, run its edge against the rim of the container to remove any excess urine.

Step 5: Put the strip horizontally on the table so that the chemicals do not mix together.

Step 6: Read the strip between 1 minute and 2 minutes after it has been dipped in the urine specimen.

Step 7: Match the colour of the strip with the colour chart on the bottle label and record the results on the monitoring form. Record '0' if the result is negative.

113

1 = trace non-haemolysed.
 2 = trace haemolysed.
 3 = +
 4 = ++
 5 = +++

6.5.3 Urine filtration Standard Operating Procedures

In diagnosing *S. haematobium*, all manufactured kits come with instructions on how to use them. It is very important to follow the instructions to ensure the quality of the results.

6.5.3.1 Safety precautions

- The urine should be considered potentially infectious.
- Wear gloves and laboratory coats whenever handling urine samples.
- Benches, instruments and equipment should be routinely decontaminated with disinfectants after use.
- Materials contaminated with infectious waste should be disinfected before disposal.
- Drinking or eating during laboratory procedures is prohibited.
- Appropriate disinfectant(s) should be used for disposal of contaminated specimen containers and for cleaning of workbenches.
- Used specimen containers must be disinfected before washing.

6.5.3.2 Equipment For

general use:

- Gloves.
- Laboratory forms.

For urine filtration:

- Urine pots (20 mL).
- Swinnex Filter Holder.
- Tweezers/Forceps.
- Syringe, plastic, 10 mL.
- Nucleopore Membrane Filter, 13 mm diameter and pore size 12 µm.
- Microscope glass slides.
- Lugol's Iodine (5.0% solution).

For microscopic examination:

- Microscope.
- Hand tally counter.

For disinfectants and waste disposal:

- Bucket (to discard urine).
- 1.0% hypochlorite solution (domestic bleach).
- Methylated spirit.
- Medicated soap.
- Rubber washing gloves.
- Disinfectant wipes.
- Waste container (containing disinfectant).

6.5.3.3 Sample collection

The number of eggs in the urine varies throughout the day, with the highest between 10:00 a.m. and 14:00 p.m. The specimen should be taken between these times and consist of a single urine sample. Since eggs are more often found at the end of a urine flow, at least 10 mL should be collected at the end of urination (the terminal urine). The easiest way to ensure a terminal urine sample is to ask individuals to 'try to fill' a large pot, e.g., 250 mL. Note that some children, particularly those who are heavily infected with SCH, may not be able to provide 10 mL of urine. Do not discard these smaller

samples, but note the volume (mL) of urine provided. Specimens should be examined as soon as possible after collection as the eggs may hatch and then become invisible, or crystals may form, making a correct diagnosis more difficult.

6.5.3.4 Steps for urine filtration

Before preparation:

- Use clean containers for urine collection.
- Ensure the filter holders and syringes are well cleaned and dry.
- Clearly and correctly label the stool containers to match the participant USIN. Step 1:

Unscrew the filter holder and insert a nucleopore filter between the two parts of the filter holder.

Make sure it is correctly held in place before screwing the unit together again.

Step 2: Thoroughly shake and mix the urine specimen before drawing a 10 mL

specimen into the syringe. Then attach the filter unit.

Step 3: Keeping the syringe and the unit in a vertical position, press the plunger down to push all the urine through the filter and out into a bucket.

Step 4: Carefully detach the syringe from the filter unit. Draw air into the syringe, reattach the syringe to the filter unit holder and expel the air again. This is important as it removes any excess urine and ensures that the eggs are firmly attached to the filter.

Step 5: Unscrew the filter holder and use a pair of tweezers to remove the filter and place it inverted onto the glass microscope slide labelled with a USIN. The top side of the filter, where the eggs were captured, should be face-up on the slide.

Step 6: Add one drop of Lugol's iodine and wait 15 seconds for the stain to penetrate the eggs. This makes the eggs more easily visible.

Step 7: Immediately examine the whole filter under a microscope at a low power (x 40). Schistosome eggs can be seen clearly because they stain orange. Record the *total number of eggs on the filter*.

Step 8: At the end of the day, wash all reusable equipment (forceps, filter holders, syringes, urine containers and glass slides) in 1.0% hypocholorite solution (domestic bleach) for use next day, discard used filters and clean the workbench.

Step 9: Around 10.0% of slides should be randomly selected and re-examined by a more experienced technician. Discrepancy in egg count should not be greater than 10.0%. *If discrepancy between readers is greater than 25.0% all slides should be reread.*

6.5.3.5 Quality control measure

Quality control was implemented to verify the consistency of the microscopic readings and to ensure compliance with the following: wearing protective gloves and clothing at all times when handling the specimen. Make sure all specimens are properly labelled.

A quality control officer, highly trained personnel on microscopy of SCH and

STH, cross-examined 10.0% of all positive and negative specimens to ensure the quality of the data. School and student personal data forms were properly filled and

USIN on the form matched with the specimen.

6.5.3.6 Survey data and processing

Standardised pre-tested, paper base form was used for data collection. Participant information included: name of schoolchild; assigning a USIN; date sample was collected; date of birth; and sex.

Data was entered using SPSS (IBM SPSS Statistics for Windows, Version 25.0.

Armonk, NY: IBM Corp.) directly from labelled forms that were collected during the

field survey, using chi square to determine the cross tabulation.

6.6 Results

6.6.1 Schools and districts

A total of 10 schools were randomly selected to participate in the study. The schools were selected from each of the eight health districts. Two districts had two schools selected from it due to the size of the districts (see Figure 6.3).





Table 6.1: Name of schools and number of participants per schoolSchool NameFrequency

| Barsee Kpangbai | 100 |
|-----------------------------------|-----|
| Community House Elementary School | 100 |
| David Fejue High School | 101 |
| E. J. Yancy Public School | 100 |

| Total | 1,003 | |
|--|-------|--|
| Sanoyea | 101 | |
| - , | | |
| Nyaquoi Bee Public School | 100 | |
| Mbelequah Public School | 100 | |
| Mawah Public School | 101 | |
| Jorkpemue Public School | 100 | |
| Faith Community AG Elementary and Junior High School | 100 | |

| Table 6.2: Sociodemographic characteristics of participants | | | | | | | |
|---|--------------|------------|-------|-------------|--|--|--|
| Level of education | on by gender | | | | | | |
| Level of | Female | Male | Total | | | | |
| education | | | | | | | |
| Grade 3 to Grade | 160 (31.7%) | 125 (25.19 | %) | 285 (28.4%) | | | |
| 6 | | | | | | | |
| Grade 7 and | 17 (3.4%) | 16 (3.2%) | | 33 (3.3%) | | | |
| above | | | | | | | |
| K1–Grade 2 | 327 (64.9%) | 358 (71.79 | %) | 685 (68.3%) | | | |
| Total | 504 | 499 | 1,003 | | | | |

Of the total 1,003 school-age children that participated in the study 504 (50.2%) were males and 499 (49.8%) were females. The majority of schoolchildren at 685 (68.3%) were in K1–Grade 2 (see Table 28). The mean age was 10.9 years and medium was 11 years (see Table 6.3).

| Table 6.3: Mean and median age of participants, by sex | | | | | | | |
|--|------------|------------|------------|--|--|--|--|
| Age | Female | Male | Overall | | | | |
| Mean (SD) | 10.9 (2.1) | 10.9 (2.0) | 10.9 (2.0) | | | | |
| Median (IQR) | 11 (9–12) | 11 (9–13) | 11 (9–12) | | | | |

IQR = interquartile range; SD = standard deviation.

6.6.3 Prevalence of S. mansoni, S. haematobium, and STH

Of the total 1,003 school-age children that had their stools examined using the single Kato-Katz method, 123 (12.3%) schoolchildren were infected with *S. mansoni*. For STH, hookworm infection was found in 2 (0.2%) schoolchildren and *T. trichiura* and *Ascaris* infections were found in 12 (1.2%) schoolchildren. Amongst STH, hookworm had the highest prevalence at 3.3% followed by *Ascaris* at 1.2%. The results showed that females had the highest prevalence of *S. haematobium* (58.0%) whereas males had the highest percentage of *Ascaris* (66.7%).

The urine filtration on *S. haematobium* showed a prevalence of 112 (11.2%) schoolchildren infected with single infections, for *S. mansoni* a prevalence of 123 (12.3%) schoolchildren infected with single infections, and a prevalence of 27 (2.7%) schoolchildren infected with double infections. There was no statistical significance between single infections, double infections and age (See Table 6.4).

| Age | Frequency | S. mansoni N (%) | Hookworm N (%) | T. trichiura N (%) | Ascaris N (%) | S. haematobium N (%) | double parasite |
|-------------|--------------|---------------------|-------------------|-----------------------|------------------|-------------------------|--------------------|
| | | | | | | | |
| 12-14 years | 352 (35.1%) | 59 (48.0%) | 15 (45.5%) | 0 (0.0%) | 3 (25.0%) | 42 (37.5%) | 12 (44.4%) |
| 15 years | 44 (4.4%) | 5 (4.0%) | 3 (9.1%) | 0 (0.0%) | 0 (0.0%) | 9 (8.0%) | 1 (3.7%) |
| Total | 1,003 (100%) | 123 (12.3%) | 33 (3.3%) | 2 (0.2%) | 12 (1.2%) | 112 (11.2%) | 27 (2.7%) |
| Sex | | | | | | | |
| Female | 504 (50.2%) | 57 (46.3%) | 13 (39.4%) | 1 (50.0%) | 4 (33.4) | 65 (58.0%) | 11 (40.7%) |
| Male | 499 (49.8%) | 66 (53.7%) | 20 (60.6%) | 1 (50.0%) | 8 (66.7) | 47 (42.0%) | 16 (59.3%) |
| Total | 1,003 (100%) | 123 (12.3%) | 33 (3.3%) | 2 (0.2%) | 12(1.2%) | 112 (11.2%) | 27 (2.7%) |

Table 6.4: Prevalence of S. mansoni, S. haematobium and STH

Table 6.5: Bivariate and multivariate logistical regression analysis of STH in relation to selected variables amongst school-age childrenin Bong County, 2017

| Variables | Presence of Helminths, n (%) | | COR (95% CI) | AOR(95% CI) | AOR(95% CI)** |
|------------|------------------------------|------------------|------------------|------------------|------------------|
| | Positive (N=47) | Negative (N=956) | | | |
| Female | 18 (38%) | 486(51%) | 1.00 | 1.00 | 1.00 |
| Male | 29 (62%) | 470(49%) | 1.85 (1.02–3.41) | 1.85 (1.03–3.42) | 1.84 (0.97–3.50) |
| Age (year) | | | | | |
| 7 – 11 | 26 (55%) | 581(61%) | 1.00 | 1.00 | 1.00 |
|--------|----------|----------|-------------------|---------------------|---------------------|
| | | | | | |
| 12-14 | 18(39%) | 334(35%) | 1.22 (0.71–3.07)* | 1.28 (0.69 – 3.46)* | 1.09 (0.78-3.52)* |
| 15 | 3(6%) | 41(4%) | 1.61 (0.79–3.25)* | 1.78 (0.38–5.75)* | 1.79 (0.56 – 5.84)* |

COR crude odds ratio, AOR adjusted odds ratio, CI confidence interval, *statistically significant at P < 0.05 in multivariate logistic regression analysis**Adjusting for clustering within schools – schools included as a fixed effect

As shown by Table 2, the prevalence of STH infection was 4.68% whereby a total of 47 school age tested positive for the soil-transmitted helminthiasis. This prevalence is relatively low compared to a study done by Novianty, et al., (2018) who investigated the risk factors for soiltransmitted helminthiasis in preschool children in North Sumatera and found a prevalence of 40.5%. Whereas Wang et al., (2012) reported that there was STH infection prevalence of 21.2% in school children.

The prevalence of soil-transmitted helminthiasis was high among male pupils 62% (29) compared with females 38% (18) showing a difference between sex and the infection of schistosomiasis. Male pupils were established to have a significant correlation with the prevalence of soiltransmitted helminthiasis (OR=1.85, 95% CI: 1.02–3.41). This shows male pupils were 1.85 times more likely to have soil-transmitted helminthiasis as compared to the female pupils. Similarly, studies conducted have also confirmed the prevalence of STH to be higher among male children as compared to the female children (Novianty, et al, 2018; Pullan, & Brooker, 2012; Speybroeck, 2017).

With regards to age, children aged between 7 and 11 years were the most infected with an infection rate of 55%, followed by those between 12 and 14 years (39%) and those aged 15 years the least at 6%. Children between 12 and 14 years, and those at 15 years were further found a

significant positive relationship with the number of soil-transmitted helminthiasis in relation infections (OR=1.22, 95% CI: 0.71-3.07 and OR=1.61, 95% CI: 0.79-3.25 respectively). This shows children between 12 and 14 years, and those at 15 years were 1.22 and 1.61 times respectively more likely to have soil-transmitted helminths as compared to the other age groups. In a similar manner, Alemu, et al, (2016) who investigated *Schistosoma mansoni* and soil-transmitted helminths among children in Chuahit, Dembia district, Ethiopia also established that prevalence intestinal helminthic infections showed significant association with age (p. value < 0.05).

Table 6.6: Bivariate and multivariate logistical regression analysis of *S. haematobium* and *S. mansoni* in relation to selected variables amongst school-age children in Bong County, 2017

| Variables | Presence of Schistosomiasis | | COR (95% CI) | AOR (95% CI) | AOR (95% CI)** |
|-------------|-----------------------------|------------|-------------------|-------------------|-------------------|
| | | | | | |
| | Yes (N=235) | No (N=768) | | | |
| Female | 122(52%) | 382(49%) | 1.00 | 1.00 | 1.00 |
| Male | 113(48%) | 386(51%) | 0.62 (0.22–1.84) | 0.60 (0.20–1.82) | 0.55 (0.19–1.78) |
| Age (years) | | | | | |
| 7 – 11 | 120(51%) | 487(63%) | 1.00 | 1.00 | 1.00 |
| 12 – 14 | 101(43%) | 251(33%) | 1.59 (1.20-2.75)* | 1.60 (1.21–2.78)* | 1.81 (1.40-2.98)* |
| 15 | 14(6%) | 30(4%) | 2.43(1.32–4.47) | 1.89(0.99–3.64) | 1.91(1.01-3.64) |
| | | | | | |

COR crude odds ratio, AOR adjusted odds ratio, CI confidence interval, *statistically significant at P < 0.05 in multivariate logistic regression analysis**Adjusting for clustering within schools – schools included as a fixed effect

As shown by Table 1, the prevalence of Schistosomiasis infection was 23.43% whereby a total of 235 school age tested positive for

Schistosomiasis. This shows a relatively high prevalence as compared to a study done by Hajissa, et al., (2018) among school children in UmAsher Area, Sudan and found that the prevalence of S. *haematobium* was 12.9%, whereas that of S. *mansoni* was 2.95%.

The prevalence of schistosomiasis was high among female pupils 122(52%) compared with males 113(48%) showing a significance between sex and the infection of schistosomiasis. However, the male pupils were established by bivariate and multivariate logistic regression analysis to have a significant correlation with the prevalence of *S. haematobium* and *S. mansoni* infections (OR=0.62, 95% CI: 0.22–1.84). This shows male pupils were 0.62 times more likely to have schistosomiasis as compared to the female pupils. This is confirmed by Omer, et al., (2016) who investigated schistosomiasis prevalence among School Age Children at Shendi Locality and also established a statistical significance between sex and the infection of schistosomiasis (P value = 0.006). With regards to age, children aged between 7 and 11 years were the most infected with an infection rate of 51%, followed by those between 12 and 14 years (43%) and those aged 15 years the least at 6%. Empirical studies conducted also found out that infections of schistosomiasis infections was statistically related to the age of the children between 12 and 14 years (OR=1.59, 95% CI: 1.20–2.75). This shows children between 12 and 14 years were 1.59 times more likely to have schistosomiasis as compared to the other age groups.

6.6.4 Prevalence of *S. mansoni, S. haematobium* and STH by school and district Table 33 shows the intensity of infection with SCH and STH. Out of the 123 schoolaged children positive for *S. mansoni* based on the Kato-Katz method, results showed that 75 (61.0%) had a light infection intensity, 25 (20.3%) had moderate infection intensity, and 23 (18.7%) had a heavy infection intensity. Out of the 112 school-aged children positive for *S. haematobium* based on the urine filtration method, results showed that 69 (61.6%) had a light infection intensity and 43 (38.4%) had a heavy infection intensity. For STH, out of the 12 school-aged children positive for *Ascaris* 7 (58.3%) had a light infection intensity. Out of 33 school-aged children positive for hookworm, 17 (51.5%) had a light infection intensity, 13 (39.4%) had a moderate infection intensity and 3 (9.1%) had a heavy infection intensity. Out of the two schoolaged children positive for *T. trichiura* both showed a light infection intensity (100.0%)

(see Table 33).

Table 6.7: Intensity of infection with SCH and STH

| | Intensity of | infection by cl | lass Par | Parasite | | |
|-----------------|--------------|-----------------|-----------------|------------|--|--|
| | Light Mod | lerate Hea | vy Total | | | |
| S. haematobium | 69 (61.6%) | 0 (0.0%) | 43 (38.4%) | 112 (100%) | | |
| S. mansoni | 75 (61.0%) | 25 (20.3%) | 23 (18.7%) | 123 (100%) | | |
| Ascaris | 7 (58.3%) | 4 (33.3%) | 1 (8.3%) | 12 (100%) | | |
| T. trichiura | 2 (100.0%) | 0 (0.0%) | 0 (0.0%) | 2 (100%) | | |
| <u>Hookworm</u> | 17 (51.5%) | 13 (39.4%) | <u>3 (9.1%)</u> | 33 (100%) | | |

| S. haematob | ium | S. mansoni | | | | <u>T. trichiur</u> | <u>a</u> | <u>Hookworm</u> | |
|--------------|--|--|---|---|--|--|---|---|---|
| | | | | <u>Ascaris</u> | | | | | |
| Male | Female | Male | Female | Male | Female | Male | Female | Male | Female |
| 13 (27.7%) | 28 (43.1%) | 42 (63.6%) | 33 (57.9%) | 5 (62.5%) | 2 (50.0%) | 1 (100%) | 1 (100%) | 11 (55.0%) | 6 (46.2%) |
| 13 (27.7%) | 15 (23.1%) | 14 (21.2%) | 11 (19.3%) | 2 (25.0%) | 2 (50.0%) | 0 (0.0%) | 0 (0.0%) | 7 (35.0%) | 6 (46.2%) |
| 21 (44.7%) | 22 (33.8%) | 10 (15.2%) | 13 (22.8%) | 1 (12.5%) | 0(0.0%) | 0(0.0%) | 0(0.0%) | 2 (10.0%) | 1 (7.6%) |
| 47 (100%) | 65 (100%) | 66 (100%) | 57 (100%) | 8 (100%) | 4 (100%) | 1 (100%) | 1 (100%) | 20 (100%) | 13 (100%) |
| | <i>S. haematob</i> Male 13 (27.7%) 13 (27.7%) 21 (44.7%) 47 (100%) | S. haematobium Male Female 13 (27.7%) 28 (43.1%) 13 (27.7%) 15 (23.1%) 21 (44.7%) 22 (33.8%) 47 65 (100%) (100%) | S. haematobium S. mansoni Male Female Male 13 (27.7%) 28 (43.1%) 42 (63.6%) 13 (27.7%) 15 (23.1%) 14 (21.2%) 21 (44.7%) 22 (33.8%) 10 (15.2%) 47 65 66 (100%) (100%) (100%) | S. haematobium S. mansoni Male Female Male Female 13 (27.7%) 28 (43.1%) 42 (63.6%) 33 (57.9%) 13 (27.7%) 15 (23.1%) 14 (21.2%) 11 (19.3%) 21 (44.7%) 22 (33.8%) 10 (15.2%) 13 (22.8%) 47 65 66 57 (100%) (100%) (100%) (100%) | S. haematobium S. mansoni Male Female Male Female Ascaris 13 (27.7%) 28 (43.1%) 42 (63.6%) 33 (57.9%) 5 (62.5%) 13 (27.7%) 15 (23.1%) 14 (21.2%) 11 (19.3%) 2 (25.0%) 13 (27.7%) 22 (33.8%) 10 (15.2%) 13 (22.8%) 1 (12.5%) 47 65 66 57 8 (100%) (100%) (100%) (100%) (100%) | S. haematobium S. mansoni Male Female Male Female Male Female 13 (27.7%) 28 (43.1%) 42 (63.6%) 33 (57.9%) 5 (62.5%) 2 (50.0%) 13 (27.7%) 15 (23.1%) 14 (21.2%) 11 (19.3%) 2 (25.0%) 2 (50.0%) 13 (27.7%) 22 (33.8%) 10 (15.2%) 13 (22.8%) 1 (12.5%) 0 (0.0%) 47 65 66 57 8 4 (100%) (100%) (100%) (100%) (100%) (100%) | S. haematobiumS. mansoniT. trichiumMaleFemaleMaleFemaleMaleFemale13 (27.7%)28 (43.1%)42 (63.6%)33 (57.9%)5 (62.5%)2 (50.0%)1 (100%)13 (27.7%)15 (23.1%)14 (21.2%)11 (19.3%)2 (25.0%)2 (50.0%)0 (0.0%)13 (27.7%)22 (33.8%)10 (15.2%)13 (22.8%)1 (12.5%)0 (0.0%)0 (0.0%)47656657841(100%)(100%)(100%)(100%)(100%)(100%)(100%) | S. haematobiumS. mansoniT. trichiuraMaleFemaleMaleFemaleMaleFemale13 (27.7%)28 (43.1%)42 (63.6%)33 (57.9%)5 (62.5%)2 (50.0%)1 (100%)13 (27.7%)15 (23.1%)14 (21.2%)11 (19.3%)2 (25.0%)2 (50.0%)0 (0.0%)0 (0.0%)13 (27.7%)22 (33.8%)10 (15.2%)13 (22.8%)1 (12.5%)0 (0.0%)0 (0.0%)0 (0.0%)476566578411(100%)(100%)(100%)(100%)(100%)(100%)(100%) | S. haematobiumS. mansoniT. trichiuraHookwormMaleFemaleMaleFemaleMaleFemaleMaleFemaleMalefemalefemaleMalefemalefemaleMalefem |

Table 6.8: Intensity of infection, by gender

| | | S. mansoni | Hookworm | Ascaris N | T. trichiura | S. haematobium |
|---|------------|-------------|------------|-----------|--------------|----------------|
| Schools | Districts | N (%) | N (%) | (%) | N (%) | N (%) |
| Mawah Public School | Fumah | 0 (0.0%) | 2 (6.1%) | 3 (25.0%) | 0 (0.0%) | 8 (7.1%) |
| Community House Elementary School | Jorquelleh | 19 (15.4%) | 1 (3.0%) | 1 (8.3%) | 0 (0.0%) | 10 (8.9%) |
| Jorkpemue Public School | Jorquelleh | 18 (14.6%) | 0 (0.0%) | 5 (41.7%) | 1 (50.0%) | 4 (3.6%) |
| Barsee Kpangbai | Kokoyah | 3 (2.4%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 18 (16.1%) |
| Nyaquoi Bee Public School | Panta | 11 (8.9%) | 4 (12.1%) | 3 (25.0%) | 1 (50.0%) | 6 (5.4%) |
| E. J. Yancy Public School | Salala | 1 (0.8%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 5 (4.5%) |
| Sanoyea | Sanoyea | 2 (1.6%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 1 (0.9%) |
| David Fejue High School | Suakoko | 0 (0.0%) | 1 (3.0%) | 0 (0.0%) | 0 (0.0%) | 27 (24.1%) |
| Faith Community AG Elementary and Junior High | Suakoko | 25 (20.3%) | 3 (9.1%) | 0 (0.0%) | 0 (0.0%) | 18 (16.1%) |
| Mbelequah Public School | Zota | 44 (35.8%) | 22 (66.7%) | 0 (0.0%) | 0 (0.0%) | 15 (13.4%) |
| Overall | | 123 (12.3%) | 33 (3.3%) | 12 (1.2%) | 2 (0.2%) | 112 (11.2%) |

| | 126 | |
|---|----------------------------------|-----------|
| Table 6.9: Prevalence of S. mansoni, S. haematobium | <i>i</i> and STH, by schools and | districts |

The intensity of SCH amongst the different age groups and gender showed that females had a higher prevalence of *S. haematobium* compared to males, but males had a higher prevalance of *S. mansoni*, *Ascaris* and hookworms. However, *T. trichiura* was prevalent in both males and females (see Table 34).

Of the total 10 schools surveyed from the eight health districts during the study, for *S. mansoni*, Mbelequah Public School in Zota district had highest positive cases with 44 (35.8%) followed by Faith Community AG Elementary and Junior High in Suakoko district with 25 (20.3%) cases. For *S. haematobium*, David Fejue High School in Suakoko district had the highest positive cases 27 (24.1%) followed by Barsee Kpangbai in Kokoya district and Faith Community AG Elementary and Junior High in Suakoko district with 18 (16.1%) each. For the STH, Mbelequah Public School had the highest positive cases of hookworm with 22 (66.7%) followed by Jorkpemue Public School with 5 (41.7%) positive cases of *Ascaris*.

6.6.5 Prevalence of haematuria using the urine strips

Total (N = 1,003)

There was no visible haematuria, however, reagent strips was dipped in the urine sample of each of the students for about 1 minute the strips were compared to the colorimetric scale on the cup of the strip. Of the 1,003 participants (111) 11.1% had positive haematuria. Amongst which 64 (57.7%) were female and 47 (42.3%) were male. The age range with the highest positivity was 7–11 years (see Table 36).

| dipstick | | |
|-------------|------------|--|
| Age | Haematuria | |
| 7–11 years | 60 (54.1%) | |
| 12–14 years | 42 (37.8%) | |
| 15 years | 9 (8.1%) | |

111 (11.1%)

Table 6.10: Percentage of participants with haematuria, by age and sex using the dipstick

| Sex | |
|--------|--------------|
| Female | 64 (57.7%) |
| Male | 47 (42.3%) |
| Total | 111 (100.0%) |

6.7 Discussion

Studies on the epidemiology of SCH in Liberia indicates high prevalence of SCH in northern Liberia, Bong, Lofa and Nimba Counties, with no snail hosts of *Schistosoma* found in the coastal regions although records on earlier scientific research reports and other relevant unpublished documents were lost during the war (Strickland, 2006). STH is the most prevalent parasitic disease in the world. These infections are most prevalent in tropical and subtropical regions of the developing world where adequate water and sanitation are lacking. These contribute to malnutrition and impaired physical and cognitive development in early childhood, poor school performance, reduced working capacity, and complication of pregnancy and birth outcome. According to a recent study, it has been noticed that even low infection intensities of multiple helminth infections enhance the risk of anaemia (Hotez, 2011).

The prevalence of *S. mansoni* infection amongst the 1,003 school-age children in Bong County, northern Liberia that participated in this study was 123 (12.3%). This is similar to a study on pre-schoolchildren in Sierra Leone that showed a prevalence rate of 11.2% (Hodges, et al., 2012) and another study on STH and SCH amongst preschool-age children in Ethopia also showed a prevalence of 11.2% (Alemu, Tegegne, Damte, & Melku, 2016). These results were higher than the nationwide study carried out in Liberia by the MoH and its partners in 2010, where the prevalence of *S. mansoni*

was 9.0% and S. haematobium was 6.0% (MoHSW, 2011b). A study on the

epidemiology of SCH in Bong County by Dennis et al. (1983) showed that there was a higher prevelance of S. mansoni (24.8%) compared to S. haematobium (22.0%). Their study also showed that the snail population in Bong County were Bulinus globosus and Biomphalaria pfeifferi, but the most prevalent snail population of the two was Bulinus globosus (Dennis, et al., 1983). The 1983 study by Dennis et al., collaborated with this study and showed that there is a higher prevalence of S. mansoni (12.3%) compared to S. haematobium (11.2%) in Bong County, Liberia. A further study conducted in Malawi on SCH in pre-school-age children observed that S. mansoni was 17.0% and S. haematobium was 45.1% (Poole, et al., 2014). These results are higher than that of this study. The prevalence of S. mansoni and S. haematobium observed in this study was lower than that of the overall prevalence of STH, where Ascaris had a prevalence at 1.2%, hookworm at 3.3% and T. trichiura at 0.2%. These results were similar to that of a study in Sierra Leone where there was a low prevalence of Ascaris (0.2%-17.2%), hookworm (8.4%–22.8%) and T. trichiura (0.9%–2.6%) infections (Hodges, et al., 2012). The low prevalence of STH can be attributed to the nationwide annual deworming programme by the Ministry of Education, MoH and their partners targeting all school-age children in the country.

Of the 10 schools that enrolled in the study, the Mbelequah Public School located in Zota district, had the highest positive cases of *S. mansoni* with 44 (35.8%) followed by Faith Community AG Elementary and Junior High in Suakoko district with 25 (20.3%). For *S. haematobium*, David Fejue High School in Suakoko district had the highest positive cases with 27 (24.1%) followed by Barsee Kpangbai in Kokoya district and Faith Community AG Elementary and Junior High in Suakoko district with 18 (16.1%) each. For STH, Mbelequah Public School had the highest positive cases of hookworm with 22 (66.7%) followed by Jorkpemue Public School with 5 (41.7%)

positive cases of *Ascaris* (see Table 35). Of the eight health districts, 10 schools located in 10 different communities were surveyed. This study revealed that the highest STH prevalence was in Zota district (35.8%) followed by Suakoko district (20.3%) and Jorquelleh (15.4%). A similar study by the MoH in Bong County, Liberia reported *Ascaris* at 21.0%, hookworm at 14.0%, and *T. trichiura* at 1.0% (MoHSW, 2011b). This study showed a decrease in the prevalence of STH, which could be attributed to the nationwide deworming programme that has been ongoing in schools around the country.

6.7.1 Intensity of SCH and STH

The intensity for *S. mansoni* based on the Kato-Katz method, results showed that 61.0% had a light infection intensity, 20.3% had moderate infection intensity, and 18.7% had a heavy infection intensity. The intensity for *S. haematobium* based on the urine filtration method showed a 61.6% had a light infection intensity and 38.4% had a heavy infection intensity. For STH, *Ascaris* had a light infection intensity of 58.3%, a moderate infection intensity of 33.3%, and a heavy infection intensity of 8.3%. Hookworm had a light infection intensity of 51.5%, a moderate infection intensity of 39.4%, and a heavy infection intensity of 9.1%. *T. trichiura* had 100.0% light infection intensity.

The intensity of SCH amongst the different age groups and gender showed that females had a higher prevalence of *S. haematobium* compared to males, but males had a higher prevalance of *S. mansoni*, *Ascaris* and hookworms. However, both males and females showed equal prevalence to *T. trichiura*.

6.8 Limitations

The study was limited to only one county as opposed to the entire country. However, the results obtained were valid and accurate, but is not a representation of the entire country. Also the intensity and prevalence of STH and SCH were obtained from a single stool specimen, which may have an influence on the accuracy of the egg count.

6.9 Conclusions

The study confirms urogenital and intestinal SCH are endemic in Bong County and that *S. haematobium* and *S. mansoni* were the two species of *Schistosoma* identified in the study. The study has shown that the prevalence of the parasitic disease varies across schools in the same county. The study also revealed that there is a low prevalence of STH across all participating schools and confirms that *T. trichiura* infection is generally low in the country. The decrease in the burden of STH could be attributed to the annual deworming programme in the country.

CHAPTER 7: KNOWLEDGE, ATTITUDES AND PRACTICES TOWARDS SCH, LF AND STH AMONGST SCHOOL-AGE CHILDREN IN BONG COUNTY, LIBERIA

7.1 Abstract

7.1.1 Background

There is considerable overlap in the geographical distribution and endemicity of LF, Oncho, SCH and STH in Liberia. Co-endemicity of the four NTDs occurs in 13 counties: Bong; Grand Gedeh; Grand Cape Mount; Grand Bassa; Grand Kru; Montserrado; Maryland; Margibi; Nimba; Lofa; Sinoe; Rivercess; and River Gee. Coendemicity of three NTDs excluding Oncho occurs in 14 counties (excluding Rivercess

County); while co-endemicity of three NTDs excluding LF occurs in 13 counties (excluding Gbarpolu and Bomi Counties); co-endemicity of two NTDs excluding LF and Oncho occurs in three counties: Bong; Nimba; and Lofa. However, there has never been a survey to determine the KAP in the population at risk to NTDs in the country.

7.1.2 Objective

To assess KAP of SCH, LF and STH amongst school-age children in Bong County, northern Liberia in order to prepare a health promotional material on the prevention and control of NTDs.

7.1.3 Methods

A cross-sectional survey was conducted in 10 primary schools from September to December 2018, using a structured and pre-tested questionnaire, to evaluate KAP amongst 1,003 school-aged children who met the inclusion criteria age of 5–14 years.

The survey was carried out on the second day following collection of samples from the schoolchildren to determine the prevalence of STH and SCH. Descriptive analysis was used for the data analysis.

7.1.4 Results

Of the 1,003 school-aged children, 90.0% had not heard about SCH, 86.0% had not heard of STH, and 92.0% had not heard of LF. For the water contact pattern only 9.0% had access to pipe water. Around 28.0% of schoolchildren had received medication during the MDA by the MoH, 21.4% slept under mosquito nets, and 72.0% used the bushes as latrines.

7.1.5 Conclusions

The study highlights the need for health education amongst schoolchildren in Bong County in order to raise awareness on the prevention and control of SCH, LF and STH and its related health risks. The study also highlights the need for improving sanitation and the use of mosquito nets.

7.2 Introduction

KAP in relation to NTDs are crucial in establishing effective disease control measures. In Liberia, data on the KAP of populations in endemic areas in Liberia with regards to NTDs are not available. Awareness of the community and their involvement are considered as one of the fundamental tools for the success and sustainability of any disease control programme (Kihara, et al., 2007). To remedy this weakness, this study assessed the KAP of school-age children on SCH, LF and STH in Bong County. It is anticipated that the findings may add new understandings about the prevention and control of NTDs in Liberia. There is considerable overlap in the geographical distribution and endemicity of LF, SCH and STH in Liberia. Co-endemicity of the four NTDs occurs in 13 out of 15 counties: Bong; Grand Gedeh; Grand Cape Mount; Grand Bassa; Grand Kru; Montserrado; Maryland; Margibi; Nimba; Lofa; Sinoe; Rivercess; and River Gee. Co-endemicity of three NTDs excluding Oncho occurs in 14 counties (excluding Rivercess County); while co-endemicity of three NTDs excluding LF occurs in 13 counties (excluding Gbarpolu and Bomi Counties); co-endemicity of two NTDs excluding LF and Oncho occurs in three counties: Bong; Nimba; and Lofa (MoHSW, 2017a). Bong County was selected for this survey due to the co-endemicity of the three NTDs under review; it has a high prevalence for SCH as compared to other counties and to be used as an exemplar to update the data on SCH for the NTD programme at the MoH after two rounds of MDA.

Universally, approximately 2 billion people are affected by schistosomes and STH, of which over 300 million suffer severe morbidity or permanent impairment. These worm infections affect the underprivileged, particularly children (Hotez, et al., 2006). At the fifty-fourth Word Health Assembly in 2001 a resolution was adopted for Member States to annually administer anthelmintic drugs to at-risk populations and to 75.0%–100.0% of school-age children that are at risk of morbidity due to STH and SCH (WHO, 2001). There is a need for integrating the control of STH with that of other NTDs (Stothard, et al., 2009b). In 2002, SCH control was launched with a grant from Bill and Melinda Gates. Since then millions of children in sub-Saharan Africa have received medication for the control for SCH and STH (Givewell, 2014).

Liberia has a population of 4.7 million. The NTDs MDA coverage index has reached 62/100. In 2016, data showed that there was 74.0% treatment coverage for LF, 31.0% treatment coverage for SCH, and 90.0% treatment coverage for STH (Uniting to Combat Neglected Tropical Diseases, 2016).

7.3 Objectives

• To assess the KAP on SCH, LF and STH amongst school-age children in Bong County, northern Liberia.

• To prepare a health promotional material on the prevention and control of NTDs.

7.4 Methodology

7.4.1 Ethical considerations

Before being interviewed, information about the questionnaire and objectives of the study were explained to the schoolchildren. They were also informed about their rights to withdraw from the study at anytime without any penalty. Written informed consent was obtained for each participant. During the entire period in each school, the principal and a teacher were present. Ethics was obtained from the MoH ethical committee, and the LSTM research and ethics committee. Recommendation for health education on the prevention and control of NTDs was made to the NTDs unit at the MoH.

7.4.2 Study design and study area

Liberia is located on the West Coast of Africa along the Atlantic Ocean. The country is divided into 15 political sub-divisions called counties. According to the World Bank development index 50.0% of the population of 4 million people are said to be under the national poverty line (World Bank, 2010). A cross-sectional survey was conducted in 10 primary schools from September to December 2018, using a structured, pre-tested and validated questionnaire, to evaluate KAP amongst 1,003 school-aged children who met the inclusion criteria (5–14 years old). The survey was carried out on the second day following collection of samples from the schoolchildren to determine the

prevalence of STH and SCH. The researcher under the supervision of a teacher and the school principle interviewed the students.

7.4.3 Study population

A total of 1,003 schoolchildren were recruited from 10 schools in Bong County. These schoolchildren had also participated in the studies on SCH and STH.

7.4.4 Questionnaire survey

A structured questionnaire for data collection was used to collect data. The questionnaire was pre-tested in a school, which was not part of those enrolled in the study. The questions were designed based on the research objectives of the larger study.

7.5 Results

7.5.1 Sociodemographic characteristics of participants

Using an integrated approach, the participants were also enrolled in the follow-up survey carried out to determine the intensity and prevalence of STH and SCH. A total of 1,003 school-aged children participated in the study where 504 (50.2%) were females and 499 (49.8%) were males. The mean age was 10.9 years and the medium age was 11 years. The majority of the schoolchildren 685 (68.3%) were in K1–Grade 2. The demographics are the same as in previous chapters as the participants were from the same study (see chapter 6).

Association of knowledge of SCH, STH and LF as it relates to gender and age are shown in Table 7.1.

| Do you know what I E is? | 7 11 yoong | 12 14 years | 15 yoong | Total |
|--------------------------|-------------|-------------|------------|-------------|
| Do you know what LF is: | 7–11 years | 12–14 years | 15 years | Total |
| Yes | 40 (6.77%) | 24 (6.79%) | 10 (16.7%) | 74 (7.4%) |
| No | 550 (93.2%) | 329 (93.2%) | 50 (83.3%) | 929 (92.6%) |
| Total | 590 | 353 | 60 | 1,003 |
| Do you know what SCH is? | 7–11 years | 12–14 years | 15 years | Total |
| Yes | 55 (9.3%) | 37 (10.5%) | 8 (13.3%) | 100 (10.0%) |
| No | 535 (90.7%) | 316 (89.5%) | 52 (86.7%) | 903 (90.0%) |
| Total | 590 | 353 | 60 | 1,003 |
| Do you know what STH is? | 7–11 years | 12-14 years | 15 years | Total |
| Yes | 71 (12.0%) | 53 (15.0%) | 11 (18.3%) | 135 (13.5%) |
| No | 519 (88.0%) | 300 (85.0%) | 49 (81.7%) | 868 (86.5%) |
| Total | 590 | 353 | 60 | 1,003 |
| GENDER | | | | |
| Do you know what STH is? | Female | Male | Total | |
| Yes | 68 (13.5%) | 67 (13.4%) | 135 (1 | 3.5%) |
| No | 436 (86.5%) | 432 (86.6%) | 868 (8 | 6.5%) |
| Total | 504 | 499 | 1,003 | |
| Do you know what LF is? | Female | Male | Total | |
| Yes | 37 (7.3%) | 37 (7.4%) | 74 (7.49 | %) |
| No | 467 92.7%) | 462 (92.6%) | 929 (92 | .6%) |
| Total | 504 | 499 | 1,003 | |
| Do you know what SCH is? | Female | Male | Total | |
| Yes | 54 (10.7%) | 46 (9.2%) | 100 (10 |).0%) |
| No | 450 (89.3%) | 453 (90.8% |) 903 (90 | 0.0%) |
| Total | 504 | 499 | 1,003 | |

 Table 7.1: Association of knowledge of STH, SCH and LF, by age and gender

 AGE

This study showed that majority of participants had not heard about SCH (90.0%), STH (86.5%) and LF (92.6%). However, 13.5% of participants had heard of STH compared to SCH (10.0%) and LF (7.4%).

The results on the knowledge of school-age children on the signs and symptoms of SCH, LF and STH are shown in Tables 40, 41 and 42. The results showed a low level of knowledge (>90.0%) on the signs and symptoms of LF, STH and SCH.

| Variables | Yes | No |
|------------------------|-----------|-------------|
| Swollen abdomen | 27 (2.7%) | 976 (97.4%) |
| Passing worms | 47 (4.5%) | 956 (95.3%) |
| Passing blood in stool | 54 (5.4%) | 949 (94.6%) |
| Lose of appetite | 48 (4.8%) | 955 (95.2%) |
| Loosing weight | 42 (4.2%) | 961 (95.8%) |
| | | |

Table 7.2: Knowledge on the signs and symptoms of STH (N = 1,003)

| Variables | Yes | No |
|------------------|-----------|---------------|
| Enlarged legs | 44 (4.4%) | 959 (95.6%) |
| Enlarged breasts | 3 (0.3%) | 1,000(99.7%) |
| Enlarged scrotum | 3 (0.3%) | 1,000 (99.7%) |
| Fever and chills | 35 (3.5%) | 968 (96.5%) |
| | | |

| Table 7.4: Knowledge on | the signs and symptoms | of SCH | <u>(N = 1,00</u> 3) |
|-------------------------|------------------------|--------|---------------------|
| Variables | Voc | No | |

| Variables | Yes | No |
|---------------------------------|------------------|--------------------|
| Abdominal pain | 43 (4.3%) | 960 (95.7%) |
| Vomitting | 44 (4.4%) | 959 (95.6%) |
| Blood in stool | 49 (4.9%) | 954 (95.1%) |
| Blood in urination | 14 (1.4%) | 989 (98.6%) |
| Painful urination | 29 (2.9%) | 974 (97.1%) |
| More frequent urination/urgency | 18 (1.8%) | 985 (98.2%) |
| Fever/headaches/chills | <u>44 (4.4%)</u> | <u>992 (98.9%)</u> |

_

| Variables | Yes | No |
|-------------------------------------|-------------|---------------|
| | | |
| Collect water from piped water | 9 (0.9%) | 994 (99.1%) |
| Collect water from wells | 412 (41.1%) | 591 (58.9%) |
| Collect water from creeks | 207 (20.6%) | 796 (79.4%) |
| Rivers | 29 (2.9 %) | 974 (97.1%) |
| Collect water from hand pumps | 491 (49.0%) | 512 (51.0%) |
| Others | 1 (0.1%) | 1,002 (99.9%) |
| Play in creeks | 492 (49.1%) | 511 (51.0%) |
| Pick up snails | 38 (3.8%) | 965 (96.2%) |
| Sand mining using pans (to be sold) | 86 (8.6%) | 917 (91.4%) |
| Wash hands before eating | 466 (46.5%) | 537 (53.5%) |
| Fishing | 264 (26.3%) | 739 (73.7%) |

Table 7.5: Water contact and sand mining behaviour

Table 7.8, shows the results on the attitude and practices of the participants. Of which, 3.8% collect snails, 8.6% were involved in sand mining, 49.1% play in creeks, and 46.5% wash their hands before eating.

On receiving drugs during MDA, of the 1,003 participants only 28.8% said they received medication for SCH, 96.9% said they received medication for deworming, but surprisingly none of the participants had received medication for LF.

| SCH | | |
|--|----------------|----------------|
| Knowledge of drug distribution | Yes | No |
| Have your received medication for SCH? | 289 (28.8%) | 714 (71.2%) |

Table 7.8: Knowledge on MDA

| 1 time 2 t | imes | 3 times | None |
|--|------------|-----------------|-----------|
| If yes how many 264 45 times? | | 13 (1.3%) | 699 |
| (24.5%) (4.5%) | | | (69.7%) |
| STH | | | |
| | Yes | No | |
| Have you taken worm tablets in the last | 972 | 31 (3.1%) | |
| 12 months? | (96.9%) | | |
| | School | Health facility | Home |
| Where did you get the deworming treatment? | 16 (51.6%) | 6 (19.4%) | 9 (29.0%) |
| Have you taken tablets for LF? | 0 (0%) | 1,003 (100.0%) | |

In this study only 1.7% of participants had access to indoor toilets (see Table

| Where do you go to Latrine | Yes | No | % Yes | |
|----------------------------|-----|-----|-------|--|
| Bush | 725 | 278 | 72.3 | |
| Pit latrine | 157 | 846 | 15.7 | |
| Behind the house | 104 | 899 | 10.4 | |
| Indoor toilet | 17 | 986 | 1.7 | |
| | | | | |

| Table 7 0. | Participant | knowladge | on conitation |
|------------------|----------------|--------------|----------------|
| Table 7.9 | I al ticipante | s kilowleuge | Ull Salitatiun |

45).

Table 7.10: Results from questions on malaria medication and test and sleeping under mosquito nets

| Variables | Ν | Percentage |
|-------------------|------------------------------|----------------|
| Have you taken m | alaria medication in the p | ast 12 months? |
| Yes | 418 | 41.7% |
| No | 585 | 58.3% |
| Have you had mal | aria test done in the last n | nonth? |
| No | 751 | 74.9% |
| Yes | 252 | 25.1% |
| Did you sleep und | er a bednet last night? | |
| Yes | 217 | 21.6% |
| No | 786 | 78.4% |
| | | |

Malaria is endemic in Liberia. Curative consultations across Liberia in 2014 were 2,347,631 visits and malaria accounts for 41.0% of all curative consultations. Inpatient admissions were 81,938, where malaria accounts for 32.0% of the total admissions. Therefore, participants were also surveyed for malaria. Of the 1,003 participants, 41.7% reported that they were treated for malaria in the past 12 months, while 74.9% reported that they were tested for malaria in the last month, and 21.6% slept under mosquito nets (see Table 46).

7.6 Discussion

This study carried out information on KAP of school-age children on SCH, LF and STH in Liberia. A parasitology follow-up survey was performed on the same 1,003 participants the day before.

The study was carried out in Bong County, which according to data from the MoH is co-endemic for STH, LF and SCH and is currently undergoing prevention and control measures by the NTD unit at the MoH. NTDs are a serious public health problem in Liberia that led the MoH to move from vertical programming to form a unit called the NTD unit. The NTD unit at the MoH has the following programmes: Oncho, which is prevalent in all 15 counties in Liberia; LF, which is prevalent in 13 counties in Liberia excluding (Gbarpolu and Bomi Counties); and STH has been recorded in all 15 counties in Liberia; Buruli ulcer and leprosy has also been recorded in all 15 counties in Liberia.

The NTD programme in the country has conducted three rounds of MDA for SCH and LF with a therapeutic coverage of 81.0%. However, results showed that more than 90.0% of the schoolchildren interviewed did not have any knowledge of LF and SCH, while 89.0% of the schoolchildren did not have knowledge of STH. In a KAP study conducted in rural Yemen it was found that 82.4% of the participants had heard of SCH, but their knowledge about the transmission and prevention of the disease was low (Sady, et al., 2015). In this present study, 90.0% of the participants had no knowledge of how the disease was transmitted and prevented, which shows that there is a great need for health education in Liberia. This is a surprise as there is active surveillance and treatment that is ongoing for LF, STH and SCH in Bong County. It was observed that many of the participants could not distinguish the name STH and SCH; therefore, this had to be explained in the local language by the research assistants. A study in Zambia reported that 53.0% of school-age children did not know how one was infected with the disease (Nyati-Jokomo & Chimbari, 2017). In this study, 90.0% of the participants did not know how one could be infected with SCH and LF. A study in sub-Saharan Africa also reported that the knowledge amongst community members about SCH was low (Sacolo, Chimbari, & Kalinda, 2018).

Majority of females did not know or had not heard about the disease, while only a few females confirmed to have knowledge of the disease. Similar findings were obtained in Tanzania where it was reported that male participants had more knowledge than female participants (Knopp, at al., 2013). For sourcing water at home only 9.0% of the schoolchildren had access to piped water, those who fetched water from wells were 41.0% and from creeks 20.6%. Up to 49.0% of the schoolchildren collected water from handpumps. This study found that the pattern of water contact in the schoolchildren could be a contributing factor for the transmission of SCH and the pattern in sand mining could be a contributing factor for being infected with STH. During this study, most of the schools were close to water bodies or sand mining, and picking up and selling snails could have attributed to the high burden of SCH in Bong County. A study in Mali in West Africa reported that schoolchildren who were close to breeding sites of snails were at risk of contracting SCH (Dabo, et al., 2015). Around 49.0% of the schoolchildren in this study admitted that they played in the creek (i.e., swimming). A study in three schools in Sudan reported that frequency of swimming was related to urogenital SCH (Ismail, et al., 2014).

When the schoolchildren were asked about MDA, 71.2% of them had no knowledge about MDA, 69.7% did not receive medication for SCH, and of those who admitted to receiving medication only 13.0% had received the three rounds of Praziquantel from the MoH during the MDA implementation. Of the 1,003 participants only 3.1% had received medication for Ivermectin and 97.0% had received anthelminthic drugs. Around 51.1% of participants admitted to taking the medication at school; this information correlated with the report from the MoH and the Ministry of Education that there is an annual deworming programme in all schools in the country. A similar study undertaken in Kenya reported that out of the 239 participants, 89.1% had heard of SCH and 87.4% had heard of STH (Gitaka, et al., 2019).

Malaria is endemic in Liberia and there have been various preventive measures put into place by the MoH, the Roll Back Malaria initiative and other partners ranging from indoor residual spraying and distribution of mosquitoes nets. In this study, 21.6% of the schoolchildren said that they slept under mosquito nets, about 99.0% heard of malaria and 25.0% had been tested for malaria within a month's time and 41.0% had taking antimalarial medication within the past 12 months. A study in northern Nigeria reported that majority of their respondents had good knowledge about malaria (Singh, Musa, Singh, & Ebere, 2014). A KAP study in Swaziland reported that a high number of the research participants had knowledge of malaria and 90.0% said they would seek treatment within 24 hours of the onset of malaria symptoms (HIongwana, Mabaso, Kunene, Govender, & Maharaj, 2006).

LF is endemic in Bong County, but 92.6% of the participants had not heard about LF, a substantial amount of them did not have knowledge of the signs and symptoms of LF, and only 3.1% of participants had taken treatment for LF. In a similar KAP study carried out in Indonesia, 89.0% of the respondents had heard of LF and while 21.0% had taken LF medication before MDA, 88.0% reported to have taken the medication after MDA (Krentel, et al., 2006).

7.7 Conclusions

This study reveals that the knowledge, symptoms and prevention of SCH, LF and STH was very low and this could be a major challenge to the achievement of the goals of the NTD programme in Liberia of: eliminating LF by 2020; achieving 100.0% treatment coverage of STH; and 75.0% therapeutic coverage for schoolchildren for SCH. This study revealed a great need for well-co-ordinated health education on NTDs amongst schoolchildren and community involvement, as well as prevention and control of these diseases. If a community is well informed about a disease the prevention measures will be practised by communities, which will lead to a decrease in prevalence of the disease.

7.8 Limitations

The parents of the schoolchildren in this study were not interviewed. It is possible that the parents may have knowledge of the diseases, but these may not be discussed with their children due to cultural practices. Also, the accuracy in recall by the participants could be low as children are unable to give accurate dates of events.

7.9 Recommendations

The MoH should strengthen the NTD programme. Health education is urgently needed in order to accelerate the elimination of NTDs. Annual social mobilisation and community sensitisation on the prevention and control of NTDs in Liberia is essential. The NTD programme in Liberia should work in collaboration with the health promotion unit and the community health unit in order to achieve their objectives.

CHAPTER 8: ASSESSMENT OF SCH-RELATED MORBIDITY AMONGST SCHOOL-AGE CHILDREN IN BONG COUNTY, LIBERIA BY THE USE OF ULTRASOUND: A CROSS-SECTIONAL STUDY

8.1 Abstract

8.1.1 Background

S. mansoni and *S. haematobium* are endemic in three counties in Liberia, despite the intermittent MDA by the MoH and its partners the disease is still prevalent. Ultrasound is a very important tool for assessing morbidty related to SCH, we can observe hepatomegaly, splenomegaly and urinary tract abnormalities. Ultrasound is accessible, mobile, cheap, and can be used on the field.

8.1.2 Objectives

To assess pathological changes of the bladder caused by S. haematobium.

To assess pathological changes of the liver caused by S. mansoni.

To access pathology changes of the spleen caused by S. mansoni.

8.1.3 Methods

A Mindray diagnostic system ultrasound Model MS MR–17003578, which has a 3.5 megahertz convex transducer was used to access the abdomen and urinary tract of 272 school-aged children from 10 different communities for SCH-related morbidity. A single sonographer scanned the participants to avoid intra-observer variability. The Niamey protocol was used as the guide for this study.

8.1.4 Results

Of the 272 school-aged children examined, only one (0.4%) had periportal fibrosis, one (0.4%) had hepatomegaly, three (1.1%) had splenomegaly, and 37 (13.6%) had starry

sky liver pattern. The urinary bladder pathology examination revealed that 53 (19.5%) had distorted bladder shape, 49 (18.0%) had focal bladder thickening where the multifocal was 19 (7.0%). Irregular bladder was found in 69 (25.4%) schoolchildren, the multifocal was found in 7 (2.6%) schoolchildren, and 2 (0.7%) of the schoolchildren had single pseudopolyp. There was no bladder mass noted.

8.1.5 Conclusions

The study revealed that *S. mansoni-* and *S. haematobium*-related morbidities such as hepatomegaly, splenomegaly and urinary tract abnormalities are prevalent amongst school-age children in Bong County, Liberia. The use of ultrasound should be encouraged in endemic areas as it helps to detect *Schistosoma*-related morbidities.

8.2 Introduction

Although SCH is controlled in some countries, it still remains a major public health problem in Liberia, especially in the Central region; this maybe due to farminig in swamps practised in this region. In Liberia, the intermediate hosts for a snail are *Bulinus globosus* and *Biomphalaria pfeifferi* (Dennis, et al., 1983).

There are more than 230 million people worldwide that are infected with SCH. Of which, 120 million are symptomatic, 20 million have severe disease and around 600 million are at risk of infection. It has been estimated that approximately 192 million (93.0% of the world total) are from sub-Saharan Africa (Kramer, Zhang, Sinclair, & Olliaro, 2014). The three species of schistosome that affect humans are *S. haematobium, S. mansoni* and *S. japonicum* (McKerrow & Salter, 2002).

The female and male adult worms live in the mesenteric veins of their host at different locations, where they mate and produce eggs. The female is about 7 mm to 20 mm long and the male is slightly smaller about 6 mm to 11 mm long. The eggs produced as a result

of mating are released either through the urine or faeces in fresh waters. The eggs then are hatched under ideal conditions and release larvae, and then environmental transmission can ensue. However, eggs die after a period of 1 to 2 weeks after they leave the worm. Upon successful hatching, the miracidium enters the compatibile snail, which is the intermediate host. Thousands of cercariae are later produced when the mother and daughter sporocytes undergo replications and their produce are released into the fresh water. The infected cercariae may penetrate the skin of any person who enters the water especially those who do swamp rice farming, swimming, snail picking or sand mining (Lawson & Wilson, 1980).

After entering the human host, the parasite lives for about 3–20 years in its host and in rare cases about 40 years (van Oordt, van den Heuvel, Tielens, & van den Bergh, 1985). In regions where SCH is endemic, research has showed that approximately 20.0%–40.0% of school-age children remain actively infected (Stothard, et al., 2011).

8.2.1 Pathogenesis and morbidity

The intensity of the infection depends on continuing exposure, the age of first exposure, and whether or not the person develops immunity against the infection (Gryseels, et al., 2006). It has been noted that the intensity of the infection is mostly seen in the first two decades of life, but declines in adults. This could be attributed to acquisition of antibodies or antigen from the disease due to continuous exposure (Buttterworth, 1998).

Not all infected individuals develop signs and symptoms of SCH. If symptoms do appear, it usually takes 4 to 6 weeks from the time of infection. The first symptom of the disease may be a general ill feeling. Within 12 hours of infection, an individual may complain of a tingling sensation or light rash, commonly referred to as 'swimmer's itch', where this is due to irritation at the point of entrance (Charnock,

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1980). The rash that may develop can mimic scabies and other types of rashes. Other symptoms can occur 2 to 10 weeks later and can include fever, aching, cough, diarrhoea or gland enlargement. These symptoms can also be related to avian SCH, which does not cause any further symptoms in humans. The manifestations of schistosomal infection vary over time as the cercariae, and later adult worms and their eggs migrate through the body (Cheng, 2014).

Signs and symptoms may include abdominal pain, diarrhoea, bloody stool or blood in the urine. Those who have been infected for a long time may experience liver damage, kidney failure, infertility or bladder cancer (squamous cell carcinoma). In children, it may cause poor growth and learning difficulties (Cheng, 2014).

The first potential reaction is an itchy, popular rash that results from cercariae penetrating the skin, often when a person is first infected. The round bumps are usually 1 cm to 3 cms in size. As people living in affected areas have often been repeatedly exposed, acute reactions are more common in tourists and migrants. The rash can occur between the first few hours and a week after exposure and lasts for several days. A similar, more severe reaction called 'swimmer's itch' can also be caused by cercariae from animal trematodes that often infect birds (Rutherford, 2000).

8.2.2 Intestinal SCH

In intestinal SCH, eggs become lodged in the intestinal wall and cause an immune system reaction called a granulomatous reaction. This immune response can lead to obstruction of the colon and blood loss. The infected individual may have what appears to be a swollen or pot-belly (Charnock, 1980). Eggs can also become lodged in the liver, leading to high blood pressure through the liver, enlarged spleen, the buildup of fluid in the abdomen, and potentially life-threatening dilations or swollen areas in the

oesophagus or gastrointestinal tract can rupture and bleed profusely (oesophageal varices). In rare instances, the central nervous system is affected (Grimes, et al., 2014).

8.2.3 Genitourinary disease

The worms of *S. haematobium* migrate to the veins around the bladder and ureters. This can lead to blood in the urine 10 to 12 weeks after infection. Over time, fibrosis can lead to obstruction of the urinary tract, hydronephrosis and kidney failure. Bladder cancer diagnosis and mortality are generally elevated in affected areas. In Egypt efforts to control SCH have led to decreases in the bladder cancer rate. The risk of bladder cancer appears to be especially high in male smokers, perhaps due to chronic irritation of the bladder lining allowing it to be exposed to carcinogens from smoking. In women, genitourinary disease can also include genital lesions that may lead to increased rates of HIV transmission (Cheng, 2014).

8.2.4 Central nervous system disease

Central nervous system lesions occur occasionally. Cerebral granulomatous disease may be caused by *Schistosoma* eggs in the brain. Communities in China affected by *S. japonicum* have been found to have rates of seizures eight times higher than the baseline. Similarly, granulomatous lesions from *S. mansoni* and *S. haematobium* eggs in the spinal cord can lead to transverse myelitis with flaccid paraplegia. Eggs are thought to travel to the central nervous system via embolisation (Oliveira, Rodrigues, Romanha, & Bahia, 2004).

8.3 Objectives

- To assess pathological changes of the kidney due to *S. haematobium*.
- To assess pathological changes of the liver due to S. mansoni.

• To access pathology changes of the spleen due to S. mansoni.

8.4 Justification of the study

Liberia is a post-conflict country, which following a number of studies have produced evidence of the presence of multiple NTDs where three NTDs are widespread, amenable to MDA and could be controlled on the same platform. However, there is no study in the country for the assessment of SCH-related morbidity amongst children.

8.5 Methodology

8.5.1 Ethical considerations

Ethics clearance was obtained from the MoH ethical committee, and the ethics committee of LSTM. The objectives of the ultrasound study was explained to the students and their parents with verbal and written consent obtained from them. They were also informed about their rights to withdraw from the study at anytime without any penalties. Informed consent was obtained for each participant. During the ultrasound examination most of the parents were present.

8.5.2 Study design and study area

Liberia is situated on the West Coast of Africa between Sierra Leone on the West, Côte d'Ivoire on the East, and Guinea on the North. The study was conducted in Bong County, one of the 15 counties in Liberia and located in the northern region. Bong County was selected due its co-endemicity with STH, SCH and LF. According to the World Bank development index 50.0% of the population of 4 million people are said to be under the national poverty line (World Bank, 2010).

8.5.3 Study design and population

The study assessed the mobidity of *S. haematobium* and *S. mansoni* by using a portable ultrasound. The bladder, liver and spleen were examined. Physical examination of the schoolchildren was performed in addition to the examination of the abdomen and urinary tract by the use of ultrasound. About 28.8% of the children admitted to having taken Praziquantel treatment. The baseline study to determine the prevalence of *S. haematobium* and *S. mansoni* was conducted 2 days before the scanning of the abdominal and urinary bladder.

The schoolchildren were aged from 5–15 years. A total of 123 schoolchildren examined by the Kato-Katz method, were found to be positive for S. *mansoni*, while 112 schoolchildren examined by urine filtration were found to be infect. 37 students not positive for *S. haematobium* and *S. mansoni* were recruited for the study.

8.5.4 Study procedures

The schoolchildren that were positive for *S. mansoni* and *S. haematobium*, their parents were informed that an ultrasound was going to be performed to determine SCH morbidity. Upon arrival at the school on the morning of the survey a room was given to the research team to be used for the ultrasound procedure.

A list was given to the school principal of those schoolchildren who were positive for SCH and who needed to be scanned. The schoolchildren were called to a room where approximately 500 mL of water was given to each schoolchild to drink 30 minutes to 1 hour before the examination. They were then scanned only if they felt the urge to urinate. Once the student entered the ultrasound room accompanied by their parents the procedure was explained to them. The schoolchild was than placed on the bed and gel placed on the abdomen to enable better resolution. A convex ultrasound probe was used to check the liver and kidney, looking for morbidities related to *S. haematobium* and *S. mansoni*. A makeshift bed was placed in the room and a portable ultrasound placed on a table. A portable generator was used to provide electricity to the machine. The ultrasound used was a Mindray diagnostic system ultrasound Model MS MR–17003578, which has a 3.5 megahertz convex transducer for the abdominal scan.

8.5.5 Kidney ultrasound

The ultrasound was performed using the Niamey protocol (Richter, Campagne, Berquist, & Jenkins, 2000). Each schoolchild was placed in a supine position, and an ultrasound gel and a linear ultrasound probe was placed on the abdomen to assess the kidneys and bladder. This procedure was carried out in each school. Bladder wall irregularity is considered when thickeness of the wall is about 5 mm. Two or more lesions are recorded as multiple lesions. Bladder wall thickness was recorded and measured in the proximity of the trigone. Normal thickness of the bladder wall is ≤ 5 mm. When there is a localised thickening of the bladder wall protruding in the lumen a score of 2 is given for the mass. The intermediate score for urinary bladder is indicated if the participant has SCH or not (Richter, et al., 2000). Scores are as follows: 0-1 = SCH unlikely; 2 = SCH likely; and >3 = SCH very likely.

8.5.6 Hepatic ultrasound

The ultrasound was performed using the Niamey protocol (Richter, et al., 2000). Each schoolchild was placed in a supine position and an ultrasound gel was placed on the abdomen. A linear convex ultrasound probe was placed on the abdomen showing the liver to check for any abnormalities as a result of SCH. This procedure was carried out

in each school. The procedure was performed early in the morning. The standard view as described in the protocol was used, but at times different views were also used. The image pattern of liver parenchyma + IP score. If the liver appears normal (pattern A) score = 0 then no further investigation is needed. If there is any echogenic periportal thickening then the score ranges from 1-8 depending on severity.

8.5.7 Spleen

The spleen was measured in the left oblique view. Each schoolchild was placed in a supine position, tilting a little towards the sonographer and ultrasound gel was placed at the anatomical level of the spleen. A linear convex ultrasound probe was used to check for abnormality (splenomegaly) as a result of SCH. This procedure was carried out in each school.

The maximum length of the spleen was measured through the splenic hilus: 0 = splenomegaly absent (mean + 2 SD) ; 1 = moderate splenomegaly (size >2 to 4 SD above mean); and 2 = marked splenomegaly (>4 SD above mean).

The following are the normal lengths of the spleen, taking into consideration that in malaria endemic areas like Liberia, the spleen may also be enlarged in the absence of SCH: 9.0 cms at 4 years; 9.5 cms at 6 years; 10.0 cms at 8 years; 11.0 cms at 10 years; 11.5 cms at 12 years; 12.0 cms at 15 years or older for girls; and 13.0 cms at 15 years or older for boys (Rosenberg, et al., 1991).

8.6 Results

Of the 272 school-age children examined by ultrasound for periportal fibrosis, only one female had periportal fibrosis (see Table 47).
| Age | No. examined | | Total % with | |
|-------------|--------------|------------|---------------------|--|
| | Male | Female | periportal fibrosis | |
| 7–11 years | 85 (59.9%) | 79 (60.8%) | 0 (0.0%) | |
| 12–14 years | 52 (36.6%) | 43 (33.1%) | 1 (0.4%) | |
| 15 years | 5 (3.5%) | 8 (6.2%) | 0 (0.0%) | |
| Total | 142 | 130 | 1 (0.4%) | |

Table 8.1: Prevalence of organomegaly and periportal fibrosis of participants, by age and sex (N = 272)

Of the 272 school-age children examined by ultrasound and clinical examination for hepatomegaly and splenomegaly, only one male (0.4%) was found to have hepatomegaly and three schoolchildren (1.1%) were found to have splenomegaly, one male (33.3%) and two females (66.7%) (see Table 48).

| by age and sex $(N = 2/2)$ | | | | | |
|----------------------------|--------------------------------------|---|---|---|--|
| No. examined | Liver enlargement | | Starry sky liver | Splenomegaly | |
| | Right lobe | Left lobe | - | | |
| | | 1 (0.4%) | | | |
| 142 | 0 (0.0%) | × / | 20 (54.1%) | 1 (33.3%) | |
| 130 | 0 (0.0%) | 0 (0.0%) | 17 (46.0%) | 2 (66.7%) | |
| 272 | 0 (0.0%) | 1 (0.4%) | 37 (13.6%) | 3 (1.1%) | |
| | No. examined 142 130 272 | No. Liver enlarg examined Right lobe 142 0 (0.0%) 130 0 (0.0%) 272 0 (0.0%) | No. examined Liver enlargement Right lobe Left lobe 142 0 (0.0%) 130 0 (0.0%) 272 0 (0.0%) | No. examinedLiver enlargement Right lobeStarry sky liver1420 (0.0%)1 (0.4%) 20 (54.1%)1300 (0.0%)0 (0.0%)17 (46.0%)2720 (0.0%)1 (0.4%)37 (13.6%) | |

Table 8.2: Prevalence of organomegaly and periportal fibrosis of participants, by age and sex (N = 272)

Figure 8.1: Participant being scanned during examination



Of the total 272 school-age children examined by ultrasound and clinical examination for urinary bladder pathology, 53 (19.5%) had distorted bladder shape, 49 (18.01%) had bladder thickening that was focal, and multifocal was 19 (7.0%). Those with irregular bladder, 69 (25.4%) were focal and 7 (2.6%) were multifocal. There was no mass noted during the process, but two schoolchildren (0.73%) were noted to have single pseudopolyp (see Table 49 and Figures 31 and 32).

| Urinary bladder pathology | | Classification | N (272) | Frequency |
|---------------------------|----------------------|----------------|---------|-----------|
| | | | | |
| Bladder shape | Normal (rectangular) | 0 | 219 | 80.5% |
| | distorted (round) | 1 | 53 | 19.5% |
| Thickening | Normal | 0 | 204 | 75.0% |
| C | Focal | 1 | 49 | 18.0% |
| | Multifocal | 2 | 19 | 7.0% |
| Irregularity | Normal | 0 | 196 | 72.1% |
| | Focal | 1 | 69 | 25.4% |
| | Multifocal | 2 | 7 | 2.6% |
| | | | | |

Table 8.3: Classification of urinary track diseases of participants

| Mass | None | 0 | 272 | 100.0% |
|-------------|----------|-----|-----|--------|
| | Single | 2 | 0 | 0.0% |
| | Multiple | n+2 | 0 | 0.0% |
| Pseudopolyp | None | 0 | 270 | 99.3% |
| | Single | 2 | 2 | 0.7% |
| | Multiple | n+2 | 0 | 0.0% |

Figure 8.2: A scan showing an irregular shaped bladder in a participant



8.7 Discussion

The patology due to *Schistosoma* may heal after the PQZ treatment, provided treatment is started early before organic complications such as, bladder cancer and hepatic scaring. There are many methods for the examination of *Schistosoma* induced pathology of the liver, kidney and spleen. The changes are usually due to the result of the *Schistosoma* eggs lodging in different tissues causing inflammation, which leads to fibrosis as a result of the host response. One of such methods is ultrasound, which is now widely used to follow-up patients. Ultrasonography of the liver, spleen and kidney for the diagnosis of pathology due to SCH usually correlate with the clinical status of the person with the disease (Abdel-Wahab & Strickland, 1993).

Ultrasound is non-invasive, has low radiation, is portable and can be used in the

field. Ultrasound has been used for over 50 years as a key method for monitoring SCH morbidity and has been reported to be accepted in communities (Hatz, 1992). The main limitations have been reported as operator skill and inter-observer differences (King, et al., 2003). In the 10 communities, there were objections by the community for the use of ultrasound on their children; this was due to the recent end of the Ebola crisis that terrified the nation making them lose confidence in health workers. They asked if adults could be scanned instead of the children. After series of meetings with the communities and the built confidence of the team, the research was agreed to continue.

Other imaging methods that can be used to detect *Schistosoma* pathology in humans and experimental animals are intravital microscopy, fluorescence molecular tomography and positron emission tomography (Skelly, 2013).

Globally, ultrasound is now frequently used to assess the morbidity of *S*. *haematobium* and *S. mansoni*. In some endemic areas morbidity has been observed in children from 7–14 years of age, and pathological changes identified by the use of ultrasound usually directly correlates with the intensity of the *Schistosoma* infection (Mostafa, Sheweita, & O'Connor, 1999). It has been reported that there is a relationship between *S. mansoni* and carcinoma of the colorectal and between *S. haematobium* and squamous cell carcinoma of the urinary bladder (Khurana, et al., 1992). In contrast, another study reported that there was no cause effect relation with the parasite in relation to tumour of the urinary bladder (Dimmette, Sproat, & Sayegh, 1956).

In this study, no histopathology was performed to determine if there was carcinoma due to SCH. Histopathology can detect if there is inflammatory granulomata due to eggs and in such cases would require barium enema or colposcopy.

The measurement of periportal fibrosis can be conducted in two ways: (i) by using the qualitative method by using image classification; and (ii) the quantitative

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method by measuring the diameter of the secondary portal branches (DoehringSchwerdtfeger, et al., 1989). In this study, both qualative and quantitative methods were used to assess periportal fibrosis.

In this study, both abdominal and urinary track ultrasounds were performed on the 272 school-age children from 10 schools in Bong County.

The results from this study showed that of the 272 participants only one schoolchild (0.4%) was noted to have periportal fibrosis, which may have been due to infection with *S. mansoni*. Periportal fibrosis lesion is usually seen after years of infection with the disease, but this was present in a schoolchild in this study. The eggs that are produced by the female worms go into the bowel, bladder and the environment. In the human body some of the *Schistosoma* eggs go into the portal veins from the mesenteric venules where some get entrapped and cause inflammation, granulomas of the liver and then fibrosis. The fibrosis of the liver could lead to obstruction of the portal veins, portal hypertension and futher leads to oesophageal varices (Cheever & Andrade, 1967).

A study in Tanzania amongst 1,840 children and adults reported that only 458 participants were invited to have extended examination. The study used the Mangagil protocol for classification of their results and reported that 65.0% of the participants were classified with periportal fibrosis. The study concluded that the ultrasound scan usually does not correlate very well with the staging of SCH and was not found to be of significance in classifying periportal fibrosis and therefore, not useful in follow-up studies on *S. mansoni* as it relates to periportal fibrosis (Asztely, Ericksson, Gabone, & Nilsson, 2016). In this study, as mentioned earlier, there was only one (0.4%) male schoolchild with periportal fibrosis compared to 36.0% in the Tanzanian study.

Furthermore, of the 458 participants invited for extended examination, 1.0% of participants had right lobe of the liver enlargement compared to 17.0% with left lobe enlargement in the Tanzanian study.

Hepatomegaly and splenomegaly in this study were defined using the Niamey protocol. For normal size left liver lobe less than or equal to 2 SD, and the right liver lobe greater than or equal to 2 SD, but less than or equal to 4 SD were classified as hepatomegaly, and splenomegaly was defined as the length of the spleen greater then 2 SD above the normal reference.

In a similar cross-sectional study, in north-western Tanzania amongst 412 participants aged 18–89 years, 29.6% had left hepatomegaly and 58.5% had right hepatomegaly (Mazigo, et al., 2017). A similar study in China, by Wiest, et al. (1992), to assess morbidity induced by *S. japonicum* in 825 participants, found hepatomegaly (75.0%) using ultrasound and physical examination. In these participants 20.0% were found to have Symmer's clay fibrosis as compared to this study where 37 schoolchildren (13.6%) were noted to have starry sky liver (Wiest, et al., 1992).

The use of ultrasound to assess pathology of the kidney associated with infection due to *S. haematobium*, has also become widely used. Studies have shown that there is a strong association between *S. haematobium* and squamous cell carcinoma of the urinary bladder. Therefore, it is important to use ultrasound to record bladder wall thickening in endemic areas for the purpose of follow-up. If a person was found to have bladder lesions and treated, this should lead to regression. If there is no change after taking medications for STH, then malignancy of the bladder should be ruled out (Abdoulaye, et al., 2015).

In this study, we found that of the 272 school-age children 53 (19.5%) had distorted bladder shape, 49 (18.0%) had focal bladder thickening, 69 (25.4%) had

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bladder wall irregularity, and 2 (0.7%) had pseudopolyp. This compares favourably with a similar study in Mali using the WHO protocol to assess *S. haematobium* pathology from 29 randomly selected schools (Abdoulaye, et al., 2015). This result was higher compared to another study in Nigeria where a total of 2,822 schoolchildren were examined by ultrasound. The results revealed that 6.0% of the schoolchildren had bladder wall thickening and irregularities, masses and pseudopolyps with an estimated upper urinary tract pathology of 3.7% (Koukounari, et al., 2006). A similar study also carried out in Nigeria amongst 138 participants (both adult and children) showed that on ultrasound 55.8% had bladder wall thickness, 69.8% was abnormal and 4.7% had pseudopolyps (Nmorsi, Ukwandu, Ogoinja, & Blackie, 2007). A cross-sectional study in Kenya amongst 1,014 participants of all ages living in *S. mansoni* endemic areas, on urine cytology examination found that amongst those without therapy for *S. haematobium* the prevalence of cellular inflammation, hyperkeratosis and squamous cell metaplasia was greater than 30.0% as compared to those treated for *S. haematobium* (Hodder, et al., 2000).

A randomised controlled trial was conducted amongst 162 pre-school-age children and 141 school-age children in Côte d'Ivoire, West Africa, to assess morbidity due to *S. haematobium*, using ultrasound examination. The participants were positive with *S. haematobium* who were compared with a placebo group. The result revealed that 67.0% of the school-age children had bladder morbidities that were mainly moderate, 4.0% had pseudopolyps, and 6.0% were found to have dilatation of the renal pelvis. Of the pre-school-age children examined, 43.0% were found to have urinary tract pathology, of which 7.0% were found to have hydronephrosis. The study reported that those in the placebo group had worse pathology compared with those treated with Praziquantel (Barda, Coulibaly, Hatz, & Keiser, 2017).

Some reasons communities reject to the study will include if there were no pror senstitition, if they think the study may have harmful effect on them and if they think there is no financial benfit.

8.8 Conclusions

This study revealed that *S. mansoni* and *S. haematobium* and their associated morbidities such as hepatomegaly, splenomegaly and urinary tract abnormalities are endemic amongst school-age children in Bong County, but not common. According to the MoH report there have been two rounds of MDA of Praziquental in Liberia, in 2013 and 2015 (MoHSW, 2017b). However, results from our study indicate that, despite the intermittent treatment, morbidity due to *S. mansoni* and *S. haematobium* are still a public health problem in Liberia. Ultrasound is very much important in assessing morbidities due to SCH as it is accessible, mobile, cheap and can be used on the field with reliable results. Its use should therefore be encouraged to access SCH-related morbidities.

8.9 Recommendations

- 1. Follow-up ultrasound study to be performed after MDA to assess progression of pathology of the disease in the participating school-age children.
- 2. Ultrasonography study to be undertaken by the MoH in all endemic counties.
- 3. MDA for SCH should be distributed to both adults and children in endemic counties.
- 4. Continuous and vigilant health education to enhance compliance and to note any challenges from participants.
- 5. Revison of pathology after Praziquantel treatment.

8.10 Limitations

This study did not take into consideration adult participants as the focus was on schoolage children.

CHAPTER 9: GENERAL DISCUSSIONS, RECOMMENDATIONS AND CONCLUSIONS

9.1. General discussions

Neglected Tropical Diseases (NTDs) continue to form a huge health burden in majority of low income countries. According to Mitjà, et al. 2017 every low income country is affected by at least one NTD while others by up to five. Due to the increasing morbidity and mortality associated with unmanaged NTDs, there is a need for urgent and effective control of NTDs, to prevent hindering of the attainment of the sustainable development goals for eliminating NTDs by 2030. As many of the NTDs have the same aetiology, epidemiology and treatments, integrated efforts in the management have the potential to control NTDs in an accountable, efficient and economical manner (Standley et al.2018). Efforts should be geared towards enhancing the detection and surveillance as recommended by the World Health Organization (WHO, 2013).

An integrated approach therefore provides an avenue for tracking the progress, promoting accountability and easy verification of the success of the management of the disease by improving the efficacy and quality of service delivery in managing NTDs. It also reduces disparities in the control programmes (WHO, 2017). Therefore, the World Health Assembly emphasises the need to intensify and integrate the programs aimed at controlling NTDs and improving the health of those affected by these diseases. Efforts have been made by several countries that have high burden of NTDs, including Liberia, to integrate the management programs of several of the diseases into a single program.

In Africa, NTDs integration has been embraced and implemented in several countries. For example, in West Africa, Sierra Leone a post conflict and post Ebola country like its neighbouring country Liberia, has an integrated some of its NTDs program vertical programs examples, LF, SCH, Oncho, and STH since 2007, but have is still having challenges like ,budget constraints to fully implement the integration and relay on health partners for support.(NTDs Masterplan Sierra Leone (2016-2020) has and Nigeria has an integrated plan for the management of tuberculosis and leprosy (Walsh et al.2015). Togo and Benin have integrated programmes for the control of yaws, leprosy and Buruli ulcers. Cameroon has integrated strategic plans for the management of yaws, Buruli ulcers and leishmaniasis (WHO, 2010). Sudan has integrated strategies for controlling vector-borne diseases that are arthropod associated (Ministry of Health report Sudan, 2015). In post-conflict Côte d'Ivoire integration was focused on the elimination of dracunculiasis instead of all of the NTDs.(Standley, et al., 2018). In Liberia, strategic plans have been designed and put into place to transfer most of NTDs programmes from vertical to integrated for the control of Schistosomiasis (SCH), Soil transmitted Helminths (STH), Onchocerchiasis

(ONCHO), Buruli ulcer and Lymphatic Filariasis (LF).

In Liberia, NTDs related prolong morbidities and in extension concomitant socioeconomic burden posed on the population with an already poor health infrastructures, and lack of logistics and funding have necessitated the embracing of the integrated program for the control of these diseases. Recent data obtained through a survey by the Ministry of Health from the mapping of NTDs in the regions in this study revealed high prevalence and overlapping of the following NTDs: onchocerciasis; lymphatic filariasis; schistosomiasis; soil-transmitted helminthiasis; Buruli ulcer; and leprosy (Ministry of Health, Liberia, 2016). Notably, more than a million people are at risk of contracting these specific NTDs. Among these at-risk group is a great proportion of school-age children that are affected by soil-transmitted helminths namely, *Ascaris lumbricoides, Trichuris trichiura* and hookworms, This class of NTDs are prevalent in

all of the 15 counties. Therefore, WHO in collaboration with the Ministry of Health has set up the integrated program for the management of these NTDs in the through preventive chemotherapy by Mass Drug Administration (MDA) and management of attendant morbidities.

Accordingly, a 5-year master plan was introduced in 2011 through the NTDs programme. This formed the basis for the implementation of the integrated form of management of these diseases. The goal of the programme is to reduce the burden of targeted NTDs, to a level that it no longer poses a public health problem, whilst simultaneously contributing to the socioeconomic development of the country.

In Liberia, the advantage of the program lies in its efficiency, time saving, cost effectiveness through its optimisation and greater control of limited resources, and labour saving. Its socioeconomic benefits are through decrease morbidity, improve health and increase productivity and therefore the alleviation poverty. However, the program implementation started regionally and is yet to cover the entire country. Even at that there are challenges experienced in these selected regions that hinder the full realization of the potential benefits of the Integrated program. Some of these challenges are either institutional, technical, resource limitation and lack of roads and trained manpower capacity. Others include Natural disasters, civil unrest, interregional migration mismanagement and lack of political will. These have to be addressed in order for the country to realize the full benefits of integration.

These challenges prevent sustained regular MDA and monitoring and evaluation and therefore leads to resurgence of the diseases, depletion of megar resources and donor fatigue. In order to succeed the integrated approach to the control of NTDs must be multidepartment, to include example, the department of education,

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and agriculture. Multi-partnership with Tropical medical institutions example, Liverpool School of Tropical Medicine, and NGOs like Sight-Savers.

The challenges enumerated including the Ebola crisis were very interruptive to the health sector of the country as well as to the integrated control of NTDs. This study however has shown for the first recorded time in Liberia the use of ultrasound to diagnose bladder and liver pathologies due to Schistosomiasis.

The study also showed demographic distribution of the respective NTDs under study in Liberia based on the vector habitat and geographic landscape, social practices as well as age groups. The absence of substantive resource local records, literatures and data base to compare were specific challenges.

The present study was conducted looking at the integrated approach to the control of three NTDs: LF; SCH; and STH. Liberia a post-conflict country is endemic for these NTDs.

The objectives of the NTD programme in the country are as follows:

- 1. To strengthen the government's ownership, advocacy, co-ordination and partnership.
- 2. To establish and sustain partnerships for the control, elimination and eradication of targeted NTDs at central, county, district and community levels.
- 3. To enhance high level reviews of NTD programme performance and the use of lessons learned to enhance advocacy, awareness and effective implementation.
- 4. To strengthen advocacy, visibility and profile of NTD control, elimination and eradication interventions at all levels.

- 5. To strengthen service delivery systems for the prevention, control and management of NTDs.
- 6. To build the capacity of service providers on NTD prevention, control and management.

To get a clearer understanding of the epidemiology and distribution of *W*. *bancrofti* in human and vector populations in Liberia, a literature review of historical documents was undertaken and results from the study showed that *A. gambiae* and *A. melas* were the prevalent vectors in the country and were found mainly in the coastal regions. This result also collaborates with the only nationwide mapping survey for LF conducted in 2010 by the MoH and its partners, the results which have been referenced in WHO publications, indicates that 13 out of the 15 counties in Liberia are endemic for LF. The MoH and its partners conducted a pre-MDA baseline survey for microfilaria in 2012. The result showed an overall 6.3% prevalence rate with males having higher prevalence compared to females.

After the establishment of the NTD programme, a nationwide survey was conducted amongst 1,489 males, they were examined to determine the morbidity of the LF. Results from the survey indicated that 12.8% of those examined had hydrocele and 6.3% had lymphoedema. Not much has been done on the morbidity control of the disease. To date, only 10 cases of hydrocelectomy has been recorded by the national health management information system unit of the MoH. The programme has since conducted three rounds of MDA for LF in 2013, 2015 and 2017 in all affected counties with therapeutic coverage of 83.0%.

There is currently an MDA plan and budget for the implementation activities in the counties including coverage and a sentinel or spot check sites survey. Based on the NTDs master plan 2016–2020, an annual implementation plan for the programme has been developed. The programme has also developed a 1-year operational plan, which is a subset of the master plan. It describes short-term ways of achieving milestones and explains how, or what portion of the master plan will be put into operation during a given operational period (MoHSW, 2016).

9.1.1 SCH

SCH is endemic in Liberia. Unfortunately, in all 15 counties in the country that have not been mapped for the disease, records on earlier scientific research, reports and other relevant documents were lost during the civil war. In 2010, there was an estimated nationwide prevalence of 24.0% for SCH (Jones, 2015). In some unpublished studies it was reported that SCH was found in mainly Bong, Nimba and Lofa Counties. However, a study by Dennis and colleagues (1983) on the epidemiology of SCH in Liberia indicated high prevalence of the disease in Central Liberia (Bong, Lofa and Nimba Counties) where the snail host responsible for the transmission was *Bulinus globosus* (for urinary SCH) and *Biompjalaria pfeifferi* (for intestinal SCH). There were no snail hosts of the disease in coastal regions (Dennis, et al., 1983).

In 2012, the MoH in collaboration with its partners conducted an integrated mapping survey for STH and SCH using the Kato-Katz method. The results indicated that in Bong County the prevalence rate for *S. mansoni* was 63.0% and for *S. haematobium* was 56.0%, in Nimba County the prevalence rate was 38.0% for *S. mansoni* and 20.0% for *S. haematobium*, and in Lofa County, the prevalence rate was 32.0% for *S. mansoni* and 10.0% for *S. haematobium*. Analysis of these results indicates that Margibi County showed a prevalence of 9.0% and 7.0% for *S. mansoni* and *S. haematobium*, respectively. The remaining counties had SCH prevalence rates ranging

from 0.0% to 3.0%. The distribution of SCH was 11.6% and 11.2% for *S. haematobium* and *S. mansoni* respectively in schoolchildren aged 5–8 years. The age group of 9–12 years had the highest prevalence rate of 58.0% for *S. haematobium* and 57.0% for *S. mansoni*; age group 13–15 years followed with prevalence of 28.2% and 30.6% for *S. haematobium* and *S. mansoni* respectively; while age group >15 years had a low prevalence rate of 2.2% for *S. haematobium* and 1.2% for *S. mansoni*. These results showed that Bong County has the highest prevalence rate of SCH in Liberia, schoolchildren aged 9–12 years are the most vulnerable, and schoolchildren above 15 years are the least affected (MoHSW, 2013).

The MoH in 2013 conducted the study again at 38 sentinel schools in the three endemic counties. Results from the 2013 study showed a marked reduction in SCH prevalence compared to the 2011 study. Since the establishment of the NTD programme in the country, the MoH has conducted two rounds of MDA for STH in the three endemic counties and had a therapeutic coverage of 81.0% (MoHSW, 2017b).

This integrated study on SCH and STH was conducted in Bong County amongst school-age children in 10 schools representing the eight health districts of Bong County. The study selected Bong County as an exemplar to provide up-to-date information for the national control programme after two rounds of MDA because of its high prevalence of STH. The study was a cross-sectional parasitological study conducted amongst 1,003 schoolchildren aged 5–15 years. The results indicated that 12.3% of the schoolchildren were infected with S. *mansoni* while 11.2% were infected with S. *haematobium*. The result in this follow-up study conducted after two rounds has shown that there is approximately a 50.0% decrease in the prevalence of the disease in Bong County as compared to the 1980s and 2011 survey results. This is attributed to:

(i) the two MDA rounds conducted in that county; and (ii) vast migration of citizens from rural areas to urban areas – evident by the overpopulation in the capital in

Monrovia and the scarcity of the population in rural areas.

9.1.2 STH

STH are widely distributed in Liberia and are prevalent in all 15 counties. A nationwide mapping survey was conducted for STH conducted in 2011 by the MoH and its partners on 59 sampled schools totalling 3,144 schoolchildren. The results indicated the prevalence rates of the disease as: *Ascaris* at 20.0%; hookworm at 9.0%; and *T. trichiura* at 3.0%. This study found generally a low prevalence of STH in Bong County and observed that hookworm had the highest prevalence at 3.3%. The current decrease in the prevalence of STH can be attributed to the annual deworming programme by the combined efforts of the Ministries of Education and Health as well as the inclusion of the deworming programme in the MoH polio campaigns.

9.1.3 KAP towards SCH, LF and STH

The present study observed the KAP towards SCH, LF and STH amongst school-age children in Bong County, northern Liberia. The results highlighted the need for health education and improved sanitation for the control of NTDs. Additional significant information gathered from this study on 1,003 school-age children was that 92.0% had not heard of LF, 86.0% had never heard of STH, and 90.0% had never heard of SCH. Furthermore, this study observed that only 9.0% of participants that came from the communities had access to pipe water; this is a major reason why these school-age children go to vector snails infested streams for their domestic water needs in addition to swamp farming.

9.1.4 SCH-related morbidity using ultrasound

This study assessed SCH-related morbidity amongst school-age children in Liberia by use of ultrasound. This study was necessary to access the pathological changes of the kidney, liver and spleen after MDA. Results from this study showed that of the 272 school-age children examined, the following pathologies were observed: hepatomegaly at 0.4%, splenomegaly at 1.1% and periportal fibrosis at 0.4%. There was no urinary bladder mass noted.

9.2 A brief analysis of the county health system and the Ebola outbreak

9.2.1 Health system situational analysis

Liberia is a low-income country with an estimated GDP per capita of US\$454 in 2013. Although the real GDP growth in 2014 had been projected at 5.8%, it was estimated to have declined to 2.5% or less by the end of 2014 due to the EVD crisis.

During the 14-year period (1989 to 2003), the health system was dysfunctional as a result of the destruction of the infrastructure and the mass exodus of the workforce. Since 2005, the country has made great strides in rebuilding the health system through reform, an introduction of the 'Basic Package of Health Services (BPHS)' under the 'National Health Policy, National Health Plan 2007–2011' (MoHSW, 2007); and later the 'Essential Package of Health Services (EPHS)' under the 'National Health Policy and Plan 2011–2012', all of which defined the type of services to be delivered at every level of care, in the face of the minimum levels of resources required to provide the package namely, infrastructure, equipment, essential medicines and human resources (MoHSW, 2016).

The main health policy document, which is the 'National Health and Social Welfare Policy' (MoHSW, 2011c), is the blue print of the strategy that identifies the priority areas including decentralisation, access to basic services, increasing health workforce and expanding the package of health services. The strategy enables transformation from a highly centralised to a decentralised client-centred health care delivery system, focusing on the essential package of health. The service delivery system is pluralistic with a variety of direct service providers (government, faith-based organisations, local and international non-governmental organisations and the private sector). The government abolished user fees in 2006 towards more equitable access to health care. However, communities frequently reported informal payments as a common practice.

There are challenges relating to facility functionality. From the 2014–2015 health facility assessment, 13.0% of all facilities did not have access to safe drinking water, 43.0% had no functional incinerators, while 45.0% did not have a primary power source for emergency lighting. This significantly limits the readiness of facilities to provide the health services required in the EPHS (MoHSW, 2016).

9.2.2 The Ebola outbreak (NTD)

The outbreak of the EVD is a zoonotic tropical disease, of which should be classified as an NTD. Ebola is usually found in countries with low economic status and in a poorly prepared health sector as that of the NTDs.

In Liberia, the outbreak was first detected in a person from Lofa County bordering on 22 March 2014, confirmed on 30 March 2014, and officially declared an EVD case by WHO on 31 March 2014. By April 2014, documented cases of EVD had spread from Lofa County to Margibi and Montserrado Counties. Because of the the risk involved both to the study participants and the researcher during the Ebola outbreak my research was suspended.

Liberia was declared EVD transmission free for the second time and certificated by WHO on 3 September 2015

9.3 Conclusions

The multiple burdens of different NTDs in Liberia are impediments to socioeconomic development of an already resource limited impoverished society with a poor health sector. This study showed the nationwide mapping of LF and has therefore, provided accurate knowledge on the distribution and epidemiology of LF and the associated morbidities in Liberia. This study has further quantifiably, for the first time, revealed the socioeconomic benefits derived from integrated MDA by significantly reducing the financial, logistics, manpower and time consuming burdens previously involved in the individual control methods used for these NTDs. This is graphically demonstrated in the pre- and post-MDA surveys by a significant decrease in SCH and STH after MDA.

9.4 Recommendations

- The integrated NTD programme at the MoH should endeavour to collaborate with other health service-related programmes at the MoH, the Ministry of Education, the Liberia water and sewer company, the National Drug Service and other relevant stakeholders in order to maximise programme impact.
- The establishment of a Centre of Excellence for Case Management and Disability Prevention of NTDs.
- Mapping for NTD-related morbidities, such as hydrocele and lymphoedema.

• Annual community-based health education programmes on NTDs to be carried out amongst all counties in the country.

9.5 Future study

If further funding was available, in the future I would wish to carry out future studies related to NTDs in Liberia. The study of priority would be: 'Using colposcopy to diagnose Female Genital Schistosomiasis (FGS) in Liberia: a cross-sectional study'.

In Liberia, doctors as well as health workers are unaware of FGS, which has gone undiagnosed or misdiagnosed for decades. This may also be due to the similarity of FGS to sexually transmitted diseases (STDs). FGS is caused by *S. haematobium*, where the patient will present with vague signs, which are similar to that of any of the STDs such as: vaginal discharge; bloody discharge; bleeding after intercourse; spotting, itching or burning sensations; pelvic pain; and pain during or after intercourse.

In my future study, I will enrol females aged 15 years and older who live in the three endemic counties (Bong, Lofa and Nimba) for SCH. Using the 28 images that are in the pocket atlas for clinical health care workers, I will use the colposcopy and carry out visual inspections of the cervix of the participants to help diagnose FGS.

The results from this study will be used to sensitise the medical community in Liberia on FGS, to raise awareness that it is a public health concern, and to highlight that FGS is one of the causes of infertility amongst women who live in endemic areas, especially in a country where infertility is a rising problem. According to observations by doctors in the gynaecological outpatient clinic, a great percentage of the clinic attendees come with complaints of primary or secondary infertility.

REFERENCES

- Abdel-Wahab, M. F., & Strickland, G. T. (1993). Abdominal ultrasonography for assessing morbidity from schistosomiasis. 2. Hospital studies. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 87(2), 135–157.
- Adenowoa, A. F., Oyinloyea, B. E., Ogunyinkaa, B. I., & Kappo, A. P. (2015). Impact of human schistosomiasis in sub-Saharan Africa. *The Brazilian Journal of Infectious Diseases*, 19(2), 196–205.
- Albonico, M., Engels, D., & Savioli, L. (2004). Monitoring drug efficacy and early detection of drug resistance in human soil-transmitted nematodes: a pressing public health agenda for helminth control. *International Journal of Parasitology*, 34(11),1205–1210.
- Al-Abd, N. M., Mohamed Nor, Z. M., Ahmed, A., Al-Adhroey, A. H., Mansor, M., & Kassim, M. (2014). Lymphatic filariasis in Peninsular Malaysia: a crosssectional survey of the knowledge, attitudes, and practices of residents. *Parasites & Vectors*, 7, 545.
- Albonico, M., Montresor, A., Crompton, D. W., & Savioli, L. (2006). Intervention for the control of soil-transmitted helminthiasis in the community. *Advances in Parasitology*, 61, 311.
- Alemu, A., Tegegne, Y., Damte, D., & Melku, M. (2016). Schistosoma mansoni and soil-transmitted helminths among preschool-aged children in Chuahit, Dembia district, Northwest Ethiopia: prevalence, intensity of infection and associated risk factors. BMC Public Health, 16, 422.
- Amazigo, U. (1999). Community selection of Ivermectin distributors. *Community Eye Health*, *12*(31), 39–40.
- Appawu, M. A., Dadzie, S. K., Baffoe-Wilmot, A., & Wilson, M. D. (2001). Lymphatic filariasis in Ghana: entomological investigation of transmission dynamics and intensity in communities served by irrigation systems in the Upper East Region of Ghana. *Tropical Medicine & International Health*, 6(7), 511–516.
- Asztely, M. S., Eriksson, B., Gabone, R. M., & Nilsson, L. Å. (2016). Is ultrasonography useful for population studies on schistosomiasis mansoni? An evaluation based on a survey on a population from Kome Island, Tanzania. *Acta Radiologica Open*, 5(12), 1–8.
- Babu, B. V., & Mishra, S. (2008). Mass drug administration under the programme to eliminate lymphatic filariasis in Orissa, India: a mixed-methods study to identify factors associated with compliance and non-compliance. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 102(12), 1207–1213.
- Bah, Y. M., Paye, J., Bah, M. S., Conteh, A., Saffa, S., & Zhang, Y. (2019). Schistosomiasis in School Age Children in Sierra Leone After 6 Years of Mass Drug Administration With Praziquantel. *Frontiers in public health*, 7, 1.
- Barber, M. A., Rice, J. B., & Brown, J. Y. (1932). Malaria studies on the firestone rubber plantation in Liberia, West Africa. *American Journal of Epidemiology*, 15(3), 601–633.
- Barda, B., Coulibaly, J. T., Hatz, C., & Keiser, J. (2017). Ultrasonographic evaluation of urinary tract morbidity in school-aged and preschool-aged children infected with *Schistosoma haematobium* and its evolution after praziquantel treatment:

a randomized controlled trial. *PLoS Neglected Tropical Diseases*, 11(2), e0005400.

- Beau de Rochars, M. V., Direny, A. N., Roberts, J. M., Addiss, D. G., Radday, J., Beach, M. J., et al. (2004). Community-wide reduction in prevalence and intensity of intestinal helminths as a collateral benefit of lymphatic filariasis elimination programs. *The American Journal of Tropical Medicine and Hygiene*, 71(4), 466–470.
- Beau de Rochars, M. V., Milord, M. D., St Jean, Y., Désormeaux, A. M., Dorvil, J. J., Lafontant, J. G., et al. (2007). Geographic distribution of lymphatic filariasis in Haiti. *The American Journal of Tropical Medicine and Hygiene*, 71(5), 598– 601.
- Bethony, J., Brooker, S., Albonico, M., Geiger, S. M., Loukas, A., Diemert, D., et al. (2006). Soil-transmitted helminth infections: ascariasis, trichuriasis, and hookworm. *The Lancet*, 367(9521):1521–1532.
- Biritwum, N.-K., de Souza, D. K., Marfo, B., Odoom, S., Alomatu, B., Asiedu, O., et al. (2017). Fifteen years of programme implementation for the elimination of Lymphatic Filariasis in Ghana: impact of MDA on immunoparasitological indicators. *PLoS Neglected Tropical Diseases*, 11(3), e0005280.
- Bockarie, M., & Molyneux, D. (2009). The end of Lymphatic filariasis? *The British Medical Journal*, 338, b1686.
- Bogus, J., Gankpala, L., Fischer, K., Krentel, A., Weil, G. J., Fischer, P. U., et al. (2016). Community attitudes toward mass drug administration for control and elimination of neglected tropical diseases after the 2014 outbreak of Ebola virus disease in Lofa County, Liberia. *The American Journal of Tropical Medicine* and Hygiene, 94(3), 497–503.
- Brinkmann, U. K. (1972). [Infections with Wuchereria bancrofti in Marshall Territory, a coastal region of Liberia]. Zeitschrift fur Tropenmedizin und Parasitologie, 23(4), 369–386.
- Brinkmann, U. K. (1977). Epidemiological investigations of Bancroftian filariasis in the coastal zone of Liberia. *Tropenmedizin und Parasitologie*, 28(1), 71–76.
- Briscoe, M. S. (1948). Insect reconnaissance in Liberia, West Africa. *Psyche*, 54(4), 246–255.
- Briscoe, M. S. (1952). The relation of insects and insect-borne diseases to the vegetation and environment in Liberia. *Ecology*, 33(2), 187–214.
- Brooker S. (2007). Spatial epidemiology of human schistosomiasis in Africa: risk models, transmission dynamics and control. *Transactions of the Royal Society* of *Tropical Medicine and Hygiene*, 101(1), 1–8.
- Brooker, S., Clements, A. C., & Bundy, D. A. (2006). Global epidemiology, ecology and control of soil-transmitted helminth infections. *Advances in Parasitology*, 62, 221–261.
- Burch, T. A., & Greenville, H. J. (1955). Filariasis in Liberia. *The American Journal of Tropical Medicine and Hygiene*, *4*, 47–51.
- Butterworth, A. E. (1998). Immunological aspects of human schistosomiasis. *British Medical Bulletin*, 54(2), 357–368.
- Cabral, S., Bonfim, C., Oliveira, R., Oliveira, P., Guimarães, T., Brandão, E., et al. (2017). Knowledge, attitudes and perceptions regarding lymphatic filariasis: study on systematic noncompliance with mass drug administration. *Revista do Instituto de Medicina Tropical de Sao Paulo, 59*, e23.

- Cano, J., Rebollo, M. P., Golding, N., Pullan, R. L., Crellen, T., Soler, A., et al. (2014). The global distribution and transmission limits of lymphatic filariasis: past and present. *Parasites & Vectors*, 7, 466.
- Cantey, P. T., Rout, J., Rao, G., Williamson, J., & Fox, L. M. (2010). Increasing compliance with mass drug administration programs for lymphatic filariasis in India through education and lymphedema management programs. *PLoS Neglected Tropical Diseases*, 4(6), e728.
- Centers for Disease Control and Prevention (CDC). (2014) Neglected Tropical Diseases: Diseases. Retrieved March 19, 2014, from http://www.cdc.gov/globalhealth/ntd/diseases.
- Centers for Disease Control and Prevention (CDC). (2017). *DPDx: Trichuriasis*. Retrieved February 6, 2020, from <u>https://www.cdc.gov/dpdx/trichuriasis</u>.
- Centers for Disease Control and Prevention (CDC). (2019a). DPDx: Lymphatic filariasis. Retrieved January 10, 2020, from <u>http://www.cdc.gov/dpdx/lymphaticFilariasis/index.html</u>.
- Centers for Disease Control and Prevention (CDC). (2019b). *DPDx: Schistosomiasis*. Retrieved February 6, 2020, from https://www.cdc.gov/dpdx/schistosomiasis/index.html.
- Centers for Disease Control and Prevention (CDC). (2019c). *DPDx: Hookworm* (*intestinal*). Retrieved February 6, 2020, from https://www.cdc.gov/parasites/hookworm/disease.html.
- Centers for Disease Control and Prevention (CDC). (2019d). *DPDx: Ascariasis*. Retrieved February 6, 2020, from <u>https://www.cdc.gov/dpdx/ascariasis</u>.
- Chandra, G., Chatterjee, S. N., Das, S., & Sarkar, N. (2007). Lymphatic filariasis in the coastal areas of Digha, West Bengal, India. *Tropical Doctor*, *37*(3), 136–139.
- Charnock, A. (1980, August 7). Taking Bilharziasis out of the irrigation equation. *New Civil Engineer*. Bilharzia caused by poor civil engineering design due to ignorance of cause and prevention.
- Cheever, A. W., & Andrade, Z. A. (1967). Pathological lesions associated with *Schistosoma mansoni* infection in man. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 61, 626–639.
- Cheng. M. (2014, June 20). Ancient parasite egg found in 6,200-year-old child skeleton gives earliest evidence of a modern disease. *National Post*. Retrieved January 10, 2020, from <u>https://nationalpost.com/news/ancient-parasite-egg-found-in6200-year-old-child-skeleton-gives-earliest-evidence-of-a-modern-disease</u>.
- Chernin, E. (1983). Sir Patrick Manson's studies on the transmission and biology of filariasis. *Reviews of Infectious Diseases*, 5(1), 148–166.
- Chitsulo, L., Engels, D., Montresor, A., & Savioli, L. (2000). The global status of schistosomiasis and its control. *Acta Tropica*, 77(1), 41.
- Chlebowsky, H. O., & Zielke, E. (1979). Studies on Bancroftian filariasis in Liberia, West Africa. II. Changes in Microfilaremia in a Rural population some years after first examination. *Tropenmedezin und Parasitologie*, *30*(2), 153–156.
- Chlebowsky, H.O., & Zielke, E. (1980). Studies on Bancroftian filariasis in Liberia, West Africa. III. Efficacy of repeated treatment with diethylcarbamazine and vector control on the microfilarial reservoir in a rural population. *Tropenmedezin und Parasitologie*, *31*(2), 181–193.

- Clements, A. C. A. Moyeed, R., & Brooker, S. (2006). Bayesian geostatistical prediction of the intensity of infection with *Schistosoma mansoni* in East Africa. *Parasitology*, *133*(Pt 6), 711–719.
- Clerinx, J., Bottieau, E., Wichmann, D., Tannich, E., & Van Esbroeck, M. (2011). Acute schistosomiasis in a cluster of travelers from Rwanda: diagnostic contribution of schistosome DNA detection in serum compared to parasitology and serology. *Journal of Travel Medicine*, *18*(6):367–372.
- Conteh, L., Engels, T., & Molyneux, D. H. (2010). Socioeconomic aspects of neglected tropical diseases. *The Lancet*, *375*(9710), 239–247.
- Coulibaly, Y. I., Dao, S., Traoré, A. K., Diallo, A., Sacko, M., & Traoré, S. F. (2006). Presence and risk of transmission of Wuchereria bancrofti is a reality in rural Mali: the case of the town of Bariambani in the Circle of Kati]. *Le Mali Medical*, 21(1), 12–17.
- Dabo, A., Diarra, A. Z., Machault, V., Touré, O., Niambélé, D. S. Kanté, A., et al. (2015). Urban schistosomiasis and associated determinant factors among school children in Bamako, Mali, West Africa. *Infectious Diseases of Poverty*, 4, 4.
- de Silva, N. R., Brooker, S., Hotez, P. J., Montresor, A., Engels, D., & Savioli, L. (2003). Soil-transmitted helminth infections: updating the global picture. *Trends* in Parasitology, 19(12), 547–551.
- de Souza, D. K., Koudou, B., Kelly-Hope, L. A., Wilson, M. D., Bockarie. M. J., & Boakye, D. A. (2012). Diversity and transmission competence in lymphatic filariasis vectors in West Africa, and the implications for accelerated elimination of Anopheles-transmitted filariasis. *Parasites & Vectors, 5*, 259.
- de Souza, D. K., Sesay, S., Moore, M. G., Ansumana, R., Narh, C. A., Kollie, K., et al. (2014). No evidence for Lymphatic Filariasis transmission in big cities affected by conflict related rural-urban migration in Sierra Leone and Liberia. *PLoS Neglected Tropical Diseases*, 8(2), e2700.
 - Dennis, E., Vorkpor, P., Holzer, B., Hanson, A., Saladin, B., Saladin, K., et al. (1983). Studies on the epidemiology of schistosomiasis in Liberia: the prevalence and intensity of schistosomal infections in Bong County and the bionomics of the snail intermediate hosts. *Acta Tropica*, 40(3), 205–229.
 - Despommier, D. D., Gwadz, R. W., & Hotez, P. J. (1995). Lymphatic Filariae: Wuchereria bancrofti (Cobbold 1877) and Brugia malayi (Brug 1927). In D. D. Despommier, R. W. Gwadz, & P. J. Hotez, (Eds.), *Parasitic Diseases* (3rd ed., pp. 40–47). New York, NY: Springer.
 - Diller, W. F. (1947). Notes on filariasis in Liberia. *Journal of Parasitology, 33*, 363–366.
 - Dimmette, R. M., Sproat, H. F., & Sayegh, E. S. (1956). The classification of carcinoma of the urinary bladder associated with schistosomiasis and metaplasia. *The Journal of Urology*, 75(4), 680–686
 - Doehring-Schwerdtfeger, E., Mohamed-Ali, G., Abdel-Rahim, I. M., Kardorff, R., Franke, D., Kaiser, C., et al. (1989). Sonomorphological abnormalities in Sudanese children with *Schistosoma mansoni* infection: a proposed staging system for field diagnosis of periportal fibrosis. *The American Journal of Tropical Medicine and Hygiene*, 41(1), 63–69.
 - Dunyo, S. K., Appawu, M., Nkrumah, F. K., Baffoe-Wilmot, A., Pedersen, E. M., & Simonsen, P. E. (1996). Lymphatic filariasis on the coast of Ghana.

Transactions of the Royal Society of Tropical Medicine and Hygiene, 90(6), 634–638.

- Eberhard, M. L., Walker, E. M., Addiss, D. G., & Lammie, P. J. (1996). A survey of knowledge, attitudes, and perceptions (KAPs) of lymphatic filariasis, elephantiasis, and hydrocele among residents in an endemic area in Haiti. *The American Journal of Tropical Medicine and Hygiene*, 54(3), 299–303.
- Fenwick, A. (2006). New initiatives against Africa's worms. *Transactions of the Royal* Society of Tropical Medicine and Hygiene, 100(3), 200–207.
- Fenwick, A. (2011). Raising the international profile of schistosomiasis. *Tropical Medicine & International Health*, 16, 23–24.
- Fenwick, A. (2017). Schistosomiasis research and control since the retirement of Sir Patrick Manson in 1914. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 111(5), 191–198.
- Fenwick, A., & Jourdan P. (2016). Schistosomiasis elimination by 2020 or 2030? International Journal for Parasitology, 46, 385–388.
- Fischer, P., Erickson, S. M., Fischer, K., Fuchs, Rao, R. U., Christensen, B. M., et al. (2007). Persistence of Brugia Malayi DNA in vector and non-vector mosquitoes: implications for xenomonitoring and transmission monitoring of lymphatic filariasis. *The American Journal of Tropical Medicine and Hygiene*, 76(3), 502–507.
- Fox, R. M. (1957). Anopheles Gambiae in relation to malaria and filariasis in coastal Liberia. *The American Journal of Tropical Medicine and Hygiene*, 6, 598–620.
- Fox, R. M. (1958). Man-biting mosquitoes in coastal Liberia. *The American Journal of Tropical Medicine and Hygiene*, 7(2), 215–220.
- Freitas, A. R., Oliveira, A. C., & Silva, L. J. (2010). Schistosomal myeloradiculopathy in a low-prevalence area: 27 cases (14 autochthonous) in Campinas, São Paulo, Brazil. *Memórias do Instituto Oswaldo Cruz, 105*(4): 398–408.
- Gbakima, A. A., Sahr, F. (1996). Filariasis in the Kaiyamba Chiefdom, Moyamba District Sierra Leone: an epidemiological and clinical study. *Public Health*, *110*(3), 169-174.
- Gbakima, A. A., Appawu, M. A., Dadzie, S., Karikari, C., Sackey, S. O., BaffoeWilmot, A., et al. (2005). Lymphatic filariasis in Ghana: establishing the potential for an urban cycle of transmission. *Tropical Medicine & International Health*, 10(4), 387–392.
- Gelfand, H. M. (1955a). Studies on the vectors of Wuchereria bancrofti in Liberia. Americal Journal of Tropical Medicine, 4(1), 52–60.
- Gelfand, H. M. (1955b). Anopheles gambiae giles and Anopheles melas Theobald in a coastal area of Liberia, West Africa. Transactions of the Royal Society of Tropical Medicine and Hygiene, 49(6), 508–527.
- Gelfand H. M. The Anopheline mosquitoes of Liberia, West Africa 1954 W. Afr. Medj, (N. S) 3 pp80-88
- Gitaka, J., Mwaura, P., Oware, K., Kongere, J., Wasonga, J., & Matendechero, S. (2019). Evaluating community's knowledge on integrated malaria, schistosomiasis and soil transmitted helminth (STH) infections in a Lake Victoria island, Kenya: A mixed method approach [version 1; peer review: 1 approved, 1 approved with reservations]. AAS Open Research, 2, 8 Retrieved February 4, 2020, from https://doi.org/10.12688/aasopenres.12897.1.
- GiveWell. (2019). SCI Foundation. Retrieved February 4, 2020, from

https://www.givewell.org/charities/schistosomiasiscontrolinitiative/November-2014.

- Global Alliance to Eliminate Lymphatic Filariasis (GAELF). Global alliance to eliminate lymphatic filariasis—progress as of January 2005. Retrieved July 27, 2009, from http://www.filariasis.org/pdfs/GAELFnumbers6_13.pdf.
- Global Programme to Eliminate Lymphatic Filariasis: progress report on mass drug administration, Weekly epidemiological record
- 2010 https://www.who.int/lymphatic_filariasis/resources/who_wer8635/en/ Global programme to eliminate lymphatic filariasis: progress report, Weekly

epidemiological record 2014

https://www.who.int/lymphatic_filariasis/resources/who_wer9038/en/ Global Programme to Eliminate Lymphatic Filariasis: progress report on mass drug

administration, Weekly epidemiological record 2016

https://www.who.int/lymphatic_filariasis/resources/who_wer9240/en/

- Gnielinski, S. V. (1972). *Liberia in maps*. London, UK: University of London Press Ltd.
- Goodman, D. S., Orelus, J. N., Roberts, J. M., Lammie, P. J., & Streit, T. G. J. (2003). PCR and mosquito dissection as tools to monitor filarial infection levels following mass treatment. *Filaria Journal*, 2(1), 11.
- Govella, N. J., Chaki, P., Mpangile, J., & Killeen, G. F. (2011). Monitoring mosquitoes in urban Dar es Salaam: evaluation of resting boxes, window exit traps, CDC light traps, Ifakara tent traps and human landing catches. *Parasites & Vectors*, 4(1), 40.
- Government of the Republic of Liberia. (2008). 2008 national population and housing census: preliminary results. Monrovia, Liberia: Government of the Republic of Liberia. Retrieved October 14, 2008, from https://www.emansion.gov.lr/doc/census_2008provisionalresults.pdf
- Gratama, S. (1966). Onchocerciasis in the south-eastern territories of Liberia. With studies on the role of Onchocerca volvulus and Wuchereria bancrofti in the pathogenesis of hydrocele and elephantiasis.. *Acta Leiden, 35*, 1–35. Retrieved February 3, 2020 from <u>https://www.ncbi.nlm.nih.gov/m/pubmed/5333986/.</u>
- Grimes, J. E., Croll, D., Harrison, W. E., Utzinger, J., Freeman, M. C., & Templeton, M. R. (2014). The relationship between water, sanitation and schistosomiasis: a systematic review and meta-analysis. *PLoS Neglected Tropical Diseases*, 8(12), e3296.
- Gryseels, B. (1996). Uncertainties in the epidemiology and control of schistosomiasis. *The American Journal of Tropical Medicine and Hygiene*, 55(5 Suppl), 103–108.
- Gryseels, B., Polman, K., Clerinx, J., & Kestens, L. (2006). Human schistosomiasis. *The Lancet*, 368(9541), 1106–1118.
- Gyapong, J. O. (2000). Lymphatic filariasis in Ghana: from research to control. Transactions of the Royal Society of Tropical Medicine and Hygiene, 94(6), 599–601.
- Gyapong, J. O., Adjei, S., Gyapong, M., & Asamoah, G. (1996). Rapid community diagnosis of lymphatic filariasis. *Acta Tropica*, *61*(1), 65–74.
- Gyapong, J. O., Kyelem, D., Kleinschmidt, I., Agbo, K., Ahouandogbo, F., Gaba J., et al. (2002). The use of spatial analysis in mapping the distribution of Bancroftian

filariasis in four West African countries. Annals of Tropical Medicine and Parasitology, 96(7), 695–705.

- Gyapong, J. O., Webber, R. H., & Bennett, S. (2002). The potential role of peripheral health workers and community key informants in the rapid assessment of community burden of disease: the example of lymphatic filariasis. *Tropical Medicine & International Health*, *3*(7), 522–528.
- Gyapong, J. O., Webber, R. H., Morris, J., & Bennett, S. (1998). The prevalence of hydrocele as a rapid diagnostic index for lymphatic filariasis. *Transactions of the Royal Society Society of Tropical Medicine and Hygiene*, 92, 40–43.
- Hajissa, K., Muhajir, A. E. M., Eshag, H. A., Alfadel, A., Nahied, E., & Mohamed, Z. (2018). Prevalence of schistosomiasis and associated risk factors among school children in Um-Asher Area, Khartoum, Sudan. *BMC research notes*, 11(1), 779.
- Hamlin, K. L., Moss, D. M., Priest, J. W., Roberts, J., Kubofcik, J., Gass, K., et al. (2012). Longitudinal monitoring of the development of antifilarial antibodies and acquisition of Wuchereria bancrofti in a highly endemic area of Haiti. *PLOS Neglected Tropical Diseases*, 6(12), e1941.
- Haque, R. (2007). Human intestinal parasites. *Journal of Health, Population and Nutrition, 25*(4), 387–391.
- Hatz, C., Murakami, H., & Jenkins, J. M. (1992). A review of the literature on the use of ultrasonography in schistosomiasis with special reference to its use in field studies: 3. *Schistosoma japonicum. Acta Tropica*, *51*(1), 29–36.
- Hatza, C., Jenkinsa, J. M., Alib, Q. M., Abdel-Wahab, M. F., Cerrid, G. G., & Tannera, M. (1992). A review of the literature on the use of ultrasonography in schistosomiasis with special reference to its use in field studies: *Schistosoma mansoni*. Acta Tropica, 51(1), 15–28.
- Hawking, F. (1957). The distribution of Bancroftian filariasis in Africa. *Bulletin of the World Health Organization*, *16*(3), 581–592.
- Hawking, F. (1977). The distribution of human filariasis throughout the world. Part III. Africa. *Tropical Diseases Bulletin*, 74(8), 649–679.
- Hlongwana, K. W., Mabaso, M. L., Kunene, S., Govender, D., & Maharaj, R. (2006).
 Community knowledge, attitudes and practices (KAP) on malaria in Swaziland:
 a country earmarked for malaria elimination. *Tropical Medicine & International Health*, 11(11), 1731–1740.
- Hodder, S. L., Mahmoud, A. A., Sorenson, K., Weinert, D. M., Stein, R. L., Ouma, J. H., et al. (2000). Predisposition to urinary tract epithelial metaplasia in Schistosoma haematobium infection. The American Journal of Tropical Medicine and Hygiene, 63(3–4), 133–138.
- Hodges, M. H. (2012). High level of *Schistosoma mansoni* infection in pre-school children in Sierra Leone highlights the need in targeting this age group for praziquantel treatment. *Acta Tropica*, *124*(2):120–125.
- Hodges, M. H., Paye, J., Koroma, M. M., Nyorkor, E. D., Fofonah, I., & Zhang, Y. (2012). Mass drug administration significantly reduces infection of *Schistosoma mansoni* and hookworm in school children in the national control program in Sierra Leone. *BMC Infectious Diseases*, 12, 16.
- Hodges, M. H., Smith, S. J., Fussum, D., Koroma, J. B., Conteh, A., Sonnie M., et al. (2010). High coverage of mass drug administration for lymphatic filariasis in rural and non-rural settings in the western area, Sierra Leone. *Parasites & Vectors*, 3(1), 120.

- Horton, J., Witt, C., Ottesen, E. A., Lazdins, J. K., Addiss, D. G., Awadzi, K., et al. (2000). An analysis of the safety of the single dose, two drug regimens used in programmes to eliminate lymphatic filariasis. *Parasitology*, 121(Suppl):S147– S160.
- Hotez P. J. Molyneux, D. H., Fenwick, A., Kumaresan, J., Sachs, S. E., Sachs, J. D., et al. (2007). Control of neglected tropical diseases. *New England Journal of Medicine*, 357(10), 1018–1027.
- Hotez, P. (2011). Enlarging the 'audacious goal': elimination of the world's highest prevalence neglected tropical diseases. *Vaccine*, 295, D104–D110.
- Hotez, P. J., & Kamath, A. (2009). Neglected tropical diseases in sub-saharan Africa: review of their prevalence, distribution, and disease burden. *PLoS Neglected Tropical Diseases*, 3(8), e412.
- Hotez, P. J., Brooker, S., Bethony, J. M., Bottazzi, M. E., Loukas, A., & Xiao, S. (2004). Hookworm infection. *New England Journal of Medicine*, *351*(8), 799–807.
- Hotez, P. J., Fenwick, A., Savioli, L., & Molyneux, D. H. (2009). Rescuing the bottom billion through control of neglected tropical diseases. *The Lancet*, 373, 1570– 1575.
- Hotez, P. J., Molyneux, D. H., Fenwick, A., Kumaresan, J., Sachs, S. E., Sachs, J. D., et al. (2007). Control of neglected tropical diseases. *New England Journal of Medicine*, 357, 1018–1127.
- Hotez, P. J., Ottesen, E., Fenwick, A., & Molyneux, D. (2006). The neglected tropical diseases: the ancient afflictions of stigma and poverty and the prospects for their control and elimination. *Advances in Experimental Medicine and Biology*, 582, 23–33.
- Hotez PJ, Molyneux DH, Fenwick A, Ottesen E, Ehrlich Sachs S, Sachs JD. Incorporating a rapid-impact package for neglected tropical diseases with programs for HIV/AIDS, tuberculosis, malaria. PLoS Med 2006;3:e102-e102
- Ichimori, K., & Graves, P. M. (2017). Overview of PacELF—the Pacific Programme for the Elimination of Lymphatic Filariasis. *Tropical Medicine and Health*, 45, 34.
- Ismail, H. A. H. A., Hong, S. T., Babiker, A. T. E. B., Hassan, R. M. A. E., Sulaiman, M. A. Z., Jeong, H. G., et al. (2014). Prevalence, risk factors, and clinical manifestations of schistosomiasis among school children in the White Nile River basin, Sudan. *Parasites & Vectors*, 7(1), 478.
- Itoh, M., Weerasooriya, M. V., Qiu, G., Gunawardena, N. K., Anantaphruti, M. T., Tesana, S., et al. (2001). Sensitive and specific enzyme-linked immunosorbent assay for the diagnosis of Wuchereria bancrofti infection in urine samples. *The American Journal of Tropical Medicine and Hygiene*, 65(4), 362–365.
- Jenkins, J. M., & Hatz, C. (Eds.). (1992). The use of diagnostic ultrasound in schistosomiasis—attempts at standardization of methodology. Cairo Working Group. *Acta Tropica*, *51*(1), 45–63.
- Jones, I. (2015). *Liberia*. Stanford, CA: Stanford University. Retrieved February 4, 2020, from https://schisto.stanford.edu/pdf/Liberia.pdf.
- Karmakar, P. R., Mitra, K., Chatterjee, A., Jana, P. K., Bhattacharya, S., & Lahiri, S. K. (2011). A study on coverage, compliance and awareness about mass drug administration for elimination of lymphatic filariasis in a district of West Bengal, India. *Journal of Vector Borne Diseases*, 48(2), 101–104.

- Keenan, J. D., Hotez, P. J., Amza, A., Stoller, N, E., Gaynor, B. D., Porco, T. C., et al. (2013). Elimination and eradication of neglected tropical diseases with mass drug administrations: a survey of experts. *PLoS Neglected Tropical Diseases*, 7(12), e2562.
- Kelly-Hope, L.A., Diggle, P. J., Rowlingson, B. S., Gyapong, J. O., Kyelem, D., Coleman, M., et al. (2006). Short communication: negative spatial association between lymphatic filariasis and malaria in West Africa. *Tropical Medicine & International Health* 11(2), 129–135.
- Khurana, P., Morad, N., Khanm A. R., Shettym S., Ibrahim, A., & Patil, K. (1992). Impact of schistosomiasis of urinary bladder cancer in the southern province of Saudi Arabia: review of 60 cases. *Journal of Tropical Medicine and Hygiene*, 95, 149–151.
- Kihara, J., Muhoho, N., Njomo, D., Mwobobia, I., Joslyne, K., Mitsui, Y., et al. (2007). Drug efficacy of praziquantel and albendazole in school children in Mwea, Central Province, Kenya. *Acta Tropica*, *102*, 165–171.
- King, C. H. (2010). Parasites and poverty: the case of schistosomiasis. *Acta Tropica*, *113*, 95–104.
- King, C. H., Dickman, K., & Tisch, D. J. (2005). Reassessment of the cost of chronic helmintic infection: a meta-analysis of disability-related outcomes in endemic schistosomiasis. *The Lancet*, 365(9470), 1561–1569.
- King, C. H., Magak, P., Salam, E. A., Ouma, J. H., Kariuki, H. C., Blanton, R. E., et al. (2003). Measuring morbidity in schistosomiasis mansoni: relationship between image pattern, portal vein diameter and portal branch thickness in large-scale surveys using new WHO coding guidelines for ultrasound in schistosomiasis. *Tropical Medicine & International Health*, 8, 109–117.
- King, C. L., Suamani, J., Sanuku, N., Cheng, YC., Satofan, S., Mancuso, B., et al. (2018). A trial of a triple-drug treatment for lymphatic filariasis. *The New England Journal of Medicine*, 379(19), 1801–1810.
- Kiszewski, A., Mellinger, A., Spielman, A., Malaney, P., Sachs, S. E., & Sachs, J. (2004). A global index representing the stability of malaria transmission. *The American Journal of Tropical Medicine and Hygiene*, 70(5), 486–498.
- Klion, A. D., Weller, P. F., & Baron, E. L., (Eds.). (2014). Lymphatic filariasis: epidemiology, clinical manifestations, and diagnosis. Reprinted from World Health Organization. (2015). Global programme to eliminate lymphatic filariasis: progress report, 2014. Weekly Epidemiological Record, 38(90), 489504.
- Knight, R. (1980). Current status of filarial infections in The Gambia. Annals of Tropical Medicine and Parasitology, 74(1), 63–68.
- Knopp, S., Person, B., Ame, S. M., Mohammed, K. A., Ali, S. M., I. Khamis, S., et al. (2013). Elimination of schistosomiasis transmission in Zanzibar: baseline findings before the onset of a randomized intervention trial. *PLoS Neglected Tropical Diseases*, 7(10), e2474.
- Knott, J. (1935). The periodicity of the microfilaria of Wuchereia bancrofti. Preliminary report of some injection experiments. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 29(1), 59–64.
- Koroma, J. B., Bangura, M. M., Hodges, M. H., Bah, M. S., Zhang, Y., & Bockarie, M. J. (2012). Lymphatic filariasis mapping by Immunochromatographic Test cards

and baseline microfilaria survey prior to mass drug administration in Sierra Leone. *Parasites & Vectors*, 5, 10.

- Koroma, J. B., Peterson, J., Gbakima, A. A., Nylander, F. E., Sahr, F., Soares Magalhães, R. J., et al. (2010). Geographical distribution of intestinal schistosomiasis and soil-transmitted helminthiasis and preventive chemotherapy strategies in Sierra Leone. *PLoS Neglected Tropical Diseases*, 4(11), e891.
- Koroma, J., & Karsor, K. (2012). Report on Microfilaria prevalence and density data collection before MDA in Liberia. Ministry of Health and Social Welfare report NTD unit.
- Kouassi, B. L., de Souza, D. K., Goepogui, A., Narh, C. A., King, S. A., Mamadou, B. S., et al. (2015). Assessing the presence of Wuchereria bancrofti in vector and human populations from urban communities in Conakry, Guinea. *Parasites & Vectors*, 8, 492.
- Koukounari, A., Sacko, M., Keita, A. D., Gabrielli, A. F., Landouré, A., Dembelé, R., et al. (2006). Assessment of ultrasound morbidity indicators of schistosomiasis in the context of large-scale programs illustrated with experiences from Malian children. *The American Journal of Tropical Medicine and Hygiene*, 75(6), 1042–1052.
- Kramer, C. V., Zhang, F., Sinclair, D., Olliaro, P. L. (2014). Drugs for treating urinary schistosomiasis. *The Cochrane Database of Systematic Reviews*, 6(8), CD000053.
- Krentel, A., Fischer, P., Manoempil, P., Supali, T., Servais, G., & Rückert, P. (2006). Using knowledge, attitudes and practice (KAP) surveys on lymphatic filariasis to prepare a health promotion campaign for mass drug administration in Alor District, Indonesia. *Tropical Medicine & International Health*, 11(11), 1731– 1740.
- Krentel, A., Peter U. Fischer, P. U., & Weil, G. J. (2013). A review of factors that influence individual compliance with mass drug administration for elimination of lymphatic filariasis. *PLoS Neglected Tropical Diseases*, 7(11), e2447.
- Kuhlow, F., & Zielke, E. (1976). Distribution and prevalence of Wuchereria bancrofti in various parts of Liberia. *Tropenmedizin und Parasitologie*, 27, 93–100.
- Kuhlow, F., & Zielke, E. (1978). Dynamics and intensity of Wuchereria bancrofti transmission in the Savannah forest regions of Liberia. *Tropenmedizin und Parasitologie*, 29(3), 371–381.
- Kumaraswami, V. (2000). The clinical manifestations of lymphatic filariasis. In T. B. Nutman (Ed.), *Lymphatic filariasis* (pp. 103–125). London, UK: Imperial College Press.
- Kyelem, D., Biswas, G., Bockarie, M. J., Bradley, M. H., El-Setouhy, M., Fischer, P. U., et al. (2008). Determinants of success in national programs to eliminate lymphatic filariasis: a perspective identifying essential elements and research needs. *The American Journal of Tropical Medicine and Hygiene*, 79(4), 480–484.
- Lamberton, P. H., & Jourdan, P. M. (2015). Human ascariasis: diagnostics update. *Current Tropical Medicine Reports*, 2(4), 189–200.
- Lardeux, F. & Cheffort, J. (2001). Ambient temperature effects on the extrinsic incubation period of Wuchereria bancrofti in Aedes polynesiensis: implications

for filariasis transmission dynamics and distribution in French Polynesia. *Medical and Veterinary Entomology*, 15(2), 167–176.

- Lawson, J. R., & Wilson, R. A. (1980). The survival of the cercariae of *Schistosoma mansoni* in relation to water temperature and glycogen utilization. *Parasitology*, 81(2):337–348.
- Liberianhistory.org. (nd). A Liberian Journey: history, memory and the making of a nation. The Havard African expedition's arrival in Liberia. Retrieved February 6, 2020 from https://liberianhistory.org/exhibits/show/chiefsuahkoko1926/th e-harvard-african-expedition.
- Madinga, J., Polman, K., Kanobana, K., van Lieshout, L., Brienen, E., Praet, N., ... & Speybroeck, N. (2017). Epidemiology of polyparasitism with Taenia solium, schistosomes and soil-transmitted helminths in the co-endemic village of Malanga, Democratic Republic of Congo. Acta tropica, 171, 186-193.
- Mangal, T. D., Paterson, S., & Fenton, A. (2008). Predicting the impact of long-term temperature changes on the epidemiology and control of schistosomiasis: A mechanistic model. *PLoS ONE*, *3*(1), e1438.
- Mangal, T. D., Paterson, S., & Fenton, A. (2010). Effects of snail density on growth, reproduction and survival of Biomphalaria alexandrina exposed to *Schistosoma mansoni*. Journal of Parasitology Research, 2010, 6.
- Manhenje, I., Galán-Puchades, M. T., & Fuentes, M. V. (2013). Socio-environmental variables and transmission risk of lymphatic filariasis in central and northern Mozambique. *Geospatial Health*, 7(2), 391–398.
- Mazigo, H. D., Nuwaha, F., Dunne, D. W., Kaatano, G. M., Angelo, T., Kepha, S., et al. (2017). *Schistosoma mansoni* infection and its related morbidity among adults living in selected villages of Mara region, north-western Tanzania: a cross-sectional exploratory study. *The Korean Journal of Parasitology*, 55(5), 533–540.
- McKerrow, J. H., & Salter, J. (2002). Invasion of skin by *Schistosoma* cercariae. *Trends in Parasitology*, 18(5), 193–119.
- McMahon, J. E., Marshall, T. F., Vaughan, J. P., & Kolstrup, N. (1979). Tanzania filariasis project: a provocative day test with diethylcarbamazine for the detection of microfilariae of nocturnally periodic Wuchereria bancrofti in the blood. *Bulletin of the World Health Organization*, 57(5), 759–765.
- Melrose, W. (2004). *Lymphatic filariasis: a review 1862-2002*. Killarney, Australia: Warwick Educational Publishing Inc.
- Melrose, W. D., Turner, P. F., Pisters, P., & Turner, B. (2000). An improved Knott's concentration test for the detection of microfilariae. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, *94*(2), 176.
- Meurs, L., Mbow, M., Vereecken, K., Menten, J., Mboup, S., & Polman, K. (2012). Bladder morbidity and hepatic fibrosis in mixed *Schistosoma haematobium* and S. mansoni infections: a population-wide study in northern Senegal. *PLoS Neglected Tropical Diseases*, 6, e1829.
- Meurs, L., Mbow, M., Vereecken, K., Menten, J., Mboup, S., & Polman, K. (2012). Epidemiology of mixed *Schistosoma mansoni* and *Schistosoma haematobium* infections in northern Senegal. *International Journal for Parasitology*, 42(3), 305–311.

- Michael, E., & Bundy, D. A. (1997). Global mapping of lymphatic filariasis. *Parasitology Today*, 13(12), 472–476.
- Michael, E., Malecela-Lazaro, M. N., Simonsen, P. E., Pedersen, E. M., Barker, G., & Kumar, A., et al. (2004). Mathematical modeling and the control of lymphatic filariasis. *The Lancet*, 4(4), 223–234.
- Micro-Richter, J. (2003). The impact of chemotherapy on morbidity due to schistosomiasis. *Acta Tropica*, 86(2–3), 161.
- Ministry of Health and Social Welfare (MoHSW). (2007). National Health Policy, National Health Plan 2007–2011. Monrovia, Liberia: MoHSW.
- Ministry of Health and Social Welfare (MoHSW). (2010). Report on lymphatic filariais mapping in Liberia using ICT cards. Monrovia, Liberia: MoHSW, Epidemiology Unit.
- Ministry of Health and Social Welfare (MoHSW). (2010). 2010 annual report. Monrovia, Liberia: MoHSW.
- Ministry of Health and Social Welfare (MoHSW). (2011a). *Country situtational analysis report*. Monrovia, Liberia: MoHSW. Retrieved February 4, 2020, from <u>https://www.medbox.org/liberia-country-situational-analysis-report-july2011/preview</u>.
- Ministry of Health and Social Welfare (MOHSW). (2011b). 2011 annual report. Monrovia, Liberia: MoHSW.
- Ministry of Health and Social Welfare (MoHSW). (2011c). *National Health and Social Welfare Policy 2011–2012*. Monrovia, Liberia: MoHSW.
- Ministry of Health and Social Welfare (MoHSW). (2013). 2013 annual report. Monrovia, Liberia: MoHSW. Retrieved February 6, 2020, from <u>https://www.medbox.org/liberia-ministry-of-health-and-social-welfare-</u>2013annual-report/download.pdf
- Ministry of Health and Social Welfare (MoHSW). (2016). NTDs Operational Plan 2016. Monrovia, Liberia: MoHSW.
- Ministry of Health and Social Welfare (MoHSW). (2017a). *Master plan for neglected tropical diseases 2016–2020*. Monrovia, Liberia: MoHSW. Retrieved February 4, 2020, from http://espen.afro.who.int/system/files/content/resources/LIBERIA_NTD_Mast er Plan 2016 2020.pdf.
- Ministry of Health and Social Welfare (MOHSW). (2017b). 2017 annual report. Monrovia, Liberia: MoHSW.
- Ministry of Health, Liberia. (2016). *Master Plan for Neglected Tropical Diseases* 20162020. Monrovia, Liberia: Ministry of Health.
- Ministry of Health, India. (2010). *Kala-azar Elimination Programme*. New Delhi: Ministry of Health. Retrieved February 12, 2020 from https://www.nhp.gov.in/kala-azar-elimination-programme.
- Ministry of Health, Sierra Leone. (2015). Master plan for Neglected tropical disease elimination in Sierra Leone 2016-2016. http://espen.afro.who.int/system/files/content/resources/SIERRA_LEONE_N TD_Master_Plan_2016_2020.pdf
- Ministry of Health, Sudan. (2015). *Integrated vector management strategic plan for Sudan 2014–2018*. Khartoum: Federal Ministry of Health, Sudan.

- Mitjà, O., Marks, M., Bertran, L., Kollie, K., Argaw, D., Fahal, A. H, et al. (2017). Integrated control and management of neglected tropical skin diseases. *PLoS Neglected Tropical Diseases*, 11(1), e0005136.
- Molyneux, D. (2003). Lymphatic filariasis (elephantiasis) elimination: a public health success and development opportunity. *Filaria Journal*, *2*, 13.
- Molyneux, D. H. (2004). Neglected diseases but unrecognized successes challenges and opportunities for infectious disease control. *The Lancet, 364*, 380–383.
- Molyneux, D. H. (2010). Neglected tropical diseases—beyond the tipping point? *The Lancet*, 375, 3–4.
- Molyneux, D. H., Hopkins, D. R., & Zagaria, N. (2004). Disease eradication, elimination and control: the need for accurate and consistent usage. *Trends in Parasitology*, 20(8), 347–351.
- Molyneux, D. H., Hotez, P. J., & Fenwick, A. (2005). 'Rapid-impact interventions': how a policy of integrated control for Africa's neglected tropical diseases could benefit the poor. *PLoS Medicine*, *2*, e336–e336.
- Molyneux, D. H., Hotez, P. J., Fenwick, A., Newman, R. D., Greenwood, B., & Sachs, J. (2009). Neglected tropical diseases and the Global Fund. *The Lancet*, 373(9660):296–297.
- Mostafa, M. H., Sheweita, S. A., & O'Connor P. J. (1999). Relationship between schistosomiasis and bladder cancer. *Clinical Microbiolgy Reviews*, 12(1), 97–111.
- Mutero, C. M., Mbogo, C., Mwangangi, J., Imbahale, S., Kibe, L., Orindi, B., et al. (2015). An assessment of participatory integrated vector management for malaria control in Kenya. *Environmental Health Perspectives*, 123(11), 1145– 1151.
- Mwakitalu, M. E., Malecela, M. N., Pedersen, E. M., Mosha, F. W., & Simonsen, P. E. (2013). Urban lymphatic filariasis in the metropolis of Dar es Salaam, Tanzania. *Parastites & Vectors, 6,* 286.
- Namewanje, H., Kabateriene, N. B., & Olson, K. (2011). The acceptability and safety of praziquantel alone and in combination with mebendazole in the treatment of *Schistosoma mansoni* and soil-transmitted helminthiasis in children aged 1-4 years in Uganda. *Parasitology*, 138(12), 1586–1592.
- Nazeh M Al-Abd, Zurainee Mohamed Nor, Abdulhamid Ahmed, Abdulelah H Al-Adhroey, Marzida Mansor, and Mustafa Kassim Lymphatic filariasis in Peninsular Malaysia: a cross-sectional survey of the knowledge, attitudes, and practices of residents2014,Parasit Vectors. 2014; 7: 545.
- Ngwira, B. M. M., Tambala, P., Perez, A. M., Bowie, C., & Molyneux, D. H. (2007). The geographical distribution of lymphatic filariasis infection in Malawi. *Filaria Journal*, 6, 12.
- Nicolls, D. J., Weld, L. H., Schwartz, E., Reed, C., von Sonnenburg, F., Freedman, D. O., et al. (2008). Characteristics of schistosomiasis in travelers reported to the GeoSentinel Surveillance Network 1997–2008. *The American Journal of Tropical Medicine and Hygiene*, 79(5), 729–734.
- Njenga, S. M., Mwandawiro, C. S., Muniu, E., Mwanje, M. T., Haji, F. M., & Bockarie, M. J. (2011a). Adult population as potential reservoir of NTD infections in rural villages of Kwale district, Coastal Kenya: implications for preventive chemotherapy interventions policy. *Parasites & Vectors*, 4, 175

- Njenga, S. M., Mwandawiro, C. S., Wamae, C. N., Mukoko, D. A., Omar, A. A., Shimada, M., et al. (2011b). Sustained reduction in prevalence of lymphatic filariasis infection in spite of missed rounds of mass drug administration in an area under mosquito nets for malaria control. *Parasites & Vectors*, 4, 90.
- Nmorsi, O. P. G., Ukwandu, N. C. D., Ogoinja, S., & Blackie, H. O. T. (2007). Urinary tract pathology in some *Schistosoma haematobium* infected Nigerians. *Journal of Biotechnology*, 6(2), 123-127.
- Norões, J., Dreyer, G., Santos, A., Mendes, V. G., Medeiros, Z., & Addiss, D. (1997). Assessment of the efficacy of diethylcarbamazine on adult Wuchereria bancrofti in vivo. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 91, 78.
- Novianty, S., Dimyati, Y., Pasaribu, S., & Pasaribu, A. P. (2018). Risk factors for soiltransmitted helminthiasis in preschool children living in farmland, North Sumatera, Indonesia. *Journal of tropical medicine*, 2018.
- Nutman, T. B., & Kazura, J. W. (2011). Lymphatic filariasis. In R. L. Guerrant, D. H. Walker, & P. F. Weller, (Eds.), *Tropical infectious diseases: principles*, *pathogens and practice* (3rd ed., pp. 729–734). Philadelphia, PA: Saunders.
- Nuwaha, F., Okware, J., & Ndyomugyenyi, R. (2005). Predictors of compliance with community-directed ivermectin treatment in Uganda: quantitative results. *Tropical Medicine & International Health*, *10*(7), 659–667.
- Nyati-Jokomo, Z., & Chimbari, M. J. (2017). Risk factors for schistosomiasis transmission among school children in Gwanda district, Zimbabwe. *Acta Tropica*, 175, 84–90.
- Oliveira, G., Rodrigues, N. B., Romanha, A. J., & Bahia, D. (2004) Genome and genomics of schistosomes. *Canadian Journal of Zoology*, 82(2), 375–390.
- Omer, E. O., Goja, A. M., Elhag, K. I., & Gadalrap, O. M. (2016). Haematobium Schistosomiasis Prevalence Among School Age Children In Irrigated Schemes At Shendi Locality, River Nile State, Sudan: Implication Of Behavior And Risk Factors. *Journal of Science* 6(4) 219-222.
- Ondigo, B. N., Muok, E. M. O., Oguso, J. K., Njenga, S. M., Kanyi, H. M., Ndombi, E. M., et al. (2018). Impact of mothers' schistosomiasis status during gestation on children's IgG antibody responses to routine vaccines 2 years later and antischistosome and anti-malarial responses by neonates in Western Kenya. *Frontiers in Immunolology*, 9, 1402.
- Otsuji, Y. (2011). History, epidemiology and control of filariasis. *Tropical Medicine* and Health, 39(1 Suppl 2), 3–13. Retrieved February 6, 2020, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3153148/
- Ottesen, E. A. (2000). The global programme to eliminate lymphatic filariasis. *Tropical Medicine & International Health*, 5(9), 591–594.
- Ottesen, E. A. (2006). Lymphatic filariasis: treatment, control and elimination. *Advances in Parasitology*, 61:395–441.
- Ottesen, E. A., Duke, B. O., Karam, M., & Behbehani, K. (1997). Strategies and tools for the control/elimination of lymphatic filariasis. *Bulletin of the World Health Organization*, 75(6), 491–503.
- Over, M., Bakote'e, B., Velayudhan, R., Wilikai, P., & Graves, P. M. (2004). Impregnated Nets or DDT Residual Spraying? Field effectiveness of malaria prevention techniques in Solomon Islands, 1993–1999. In J. G. Breman, M. S. Alilio, & A. Mills A, (Eds.), *The intolerable burden of malaria II: What's new*,

what's needed: supplement to volume 71(2) of The American Journal of Tropical Medicine and Hygiene. Northbrook, IL: The American Society of Tropical Medicine and Hygiene.

- Pani, S. P., & Srividya, A. (1995). Clinical manifestations of Bancroftian filariasis with special reference to lymphoedema grading. *Indian Journal of Medical Research*, 102, 114–118.
- Pani, S. P., Yuvaraj, J., Vanamail, P., Dhanda, V., Michael, E., Grenfell, B. T., et al. (1995). Episodic adenolymphangitis and lymphoedema in patients with Bancroftian filariasis. *Transactions of the Royal Society of Tropical Medicine* and Hygiene, 89(1), 72–74.
- Partono, F., Maizels, R. M., & Purnomo. (1989). Towards a filariasis-free community: evaluation of filariasis control over an eleven year period in Flores, Indonesia. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 83(6), 821–826.
- Perera, M., Whitehead, M., Molyneux, D., Weerasooriya, M., & Gunatilleke, G. (2007). Neglected patients with a neglected disease? A qualitative study of lymphatic filariasis. *PLoS Neglected Tropical Diseases*, 1(2), e128.
- Peters, W. (1956). Mosquitoes of Liberia (Diptera: Culicidae): a general survey. Bulletin of Entomological Research, 47(3), 525–551.
- Poindexter, H. A. (1950). Filariasis bancrofti studies in Liberia. *The American Journal* of Tropical Medicine and Hygiene, 30(4), 519–523.
- Pollack HM, Banner MP, Martinez LO, Hodson CJ. Diagnostic considerations in urinary bladder wall calcification AJR Am J Roentgenol. 1981 Apr;136(4):791-7.
- Poole, H., Terlouw, D. J., Naunje, A., Mzembe, K., Stanton, M., Betson, M., et al. (2014). Schistosomiasis in pre-school-age children and their mothers in Chikhwawa district, Malawi with notes on characterization of schistosomes and snails. *Parasites & Vectors 7*, 153.
- Proffitt, R. D., & Walton, B. C. (1962). Ascaris pnuemonia in a two-year-old girl. The New England Journal of Medicine, 266, 931–934.
- Puchner, K. P., Parisi, S., Schwienhorst-Stich, E.-M., Kasang, C., Salah, M., & Tanyous, E. (2017) Trends and patterns in leprosy in nine states of the Republic of the Sudan 7 years after the introduction of routine contact screening. *Transactions of The Royal Society of Tropical Medicine and Hygiene*, 111(8), 354–359. <u>https://doi.org/10.1093/trstmh/trx063</u>
- Pullan, R. L., & Brooker, S. J. (2012). The global limits and population at risk of soiltransmitted helminth infections in 2010. *Parasites & vectors*, 5(1), 81.
- Raghavan, N. G. S. (1961). The vectors of human infections by Wuchereria species in endemic areas and their biology. *Bulletin of the World Health Organization*, 24(2), 177–195.
- Ramaiah, K. D., & Kumar, K. N. (2000). Effect of lymphatic filariasis on school children. *Acta Tropica*, *76*, 197–199.
- Ramaiah, K. D., & Ottesen, E. A. (2014). Progress and impact of 13 years of the global programme to eliminate lymphatic filariasis on reducing the burden of filarial disease. *PLoS Neglected Tropical Diseases*, 8(11), e3319.
- Rebollo, M. P., Sambou, S. M., Thomas, B., Biritwum, N.-K., Jaye, M. C., Kelly-Hope, L., et al. (2015). Elimination of lymphatic filariasis in The Gambia. *PLoS Neglected Tropical Diseases*, 9(3), e0003642.
- Ribbands C. R. (1994a). Differences between Anopheles melas (A. gambiae var. melas) and Anopheles gambiae. I. The larval pecten. Annals of Tropical Medicine & Parasitology, 38(2), 85–86.
- Ribbands, C. R. (1944b). Differences between *Anopheles melas* and *Anopheles gambiae*. II. Salinity relations of larvae and maxillary palp banding of adult females. *Annals of Tropical Medicine and Parasitology*, 38(2), 87–99.
- Richter, J., Hatz, C., Campagne, G., Bergquist, N. R., & Jenkins, J. M. (2000). Ultrasound in schistosomiasis: a practical guide to the standard use of ultrasonography for assessment of schistosomiasis-related morbidity. Geneva: World Health Organization.
- Rosenberg, H. K., Markowitz, R. I., Kolberg, H., Park, C., Hubbard, A., & Bellah, R.
 D. (1991). Normal splenic size in infants and children: sonographic measurements. *American Journal of Roentgenology*, 157(1), 119–121.
- Rudge, J. W., Stothard, J. R., Basáñez, M. G., Mgeni, A. F., Khamis, I. S., Khamis, A. N., et al. (2008). Micro-epidemiology of urinary schistosomiasis in Zanzibar: local risk factors associated with distribution of infections among schoolchildren and relevance for control. *Acta Tropica*, 105(1), 45–54.
- Rudge, J. W., Webster, J. P., Lu, D. B., Wang, T. P., Fang, G. R., & Basáñez, M. G. (2013). Identifying host species driving transmission of schistosomiasis japonica, a multihost parasite system, in China. *Proceedings of the National Academy of Sciences of the United States of America*, 110(28), 11457–11462.
- Rutherford, P. (2000). The diagnosis of schistosomiasis in modern and ancient tissues by means of immunocytochemistry. *Chungará* (Arica), 32(1), 127–131.
- Sacolo, H., Chimbari, M., & Kalinda, C. (2018). Knowledge, attitudes and practices on Schistosomiasis in sub-Saharan Africa: a systematic review. *BMC Infectious Diseases*, 18, 46.
- Sady, H., Al-Mekhlafi, H. M., Atroosh, W. M., Al-Delaimy, A. K., Nasr, N. A., Dawaki, S., et al. (2015). Knowledge, attitude, and practices towards schistosomiasis among rural population in Yemen. *Parasites & Vectors*, 8(1), 436.
- Schwartz, D. A. (1981). Helminths in the induction of cancer. II. Schistosoma haematobium and bladder cancer. Tropical and Geographical Medicine, 33, 1– 7.
- Shi, Z. J. (1994). [Study on the transmission threshold of filariasis. Collaborating research group on the transmission threshold of filariasis]. *Chinese Journal of Parasitology & Parasitic Diseases*, 12(1), 1–6.
- Shi, Z. J. Shumbej, T., Belay, T., Mekonnen, Z., Tefera, T., & Zemene E. (2015). Soiltransmitted helminths and associated factors among pre-school children in Butajira town, South Central Ethiopia: a community-based cross-sectional study. *PLoS ONE*, 10(8), e0136342.
- Simonsen, P. E., & Mwakitalu, M. E. (2013). Urban lymphatic filariasis. *Parasitology Research*, *112*(1), 35–44.
- Singh, R., Musa, J., Singh, S., & Ebere, U. V. (2014). Knowledge, attitude and practices on malaria among the rural communities in Aliero, northern Nigeria. *Journal of Family Medicine and Primary Care*, 3(1), 39–44.
- Skelly, P. (2013). The use of imaging to detect schistosomes and diagnose schistosomiasis. *Parasite Immunology*, 35(0): 295–301.

- Sokolow, S. H., Wood, C. L., Jones, I. J., Swartz, S. J., Lopez, M., Hsieh, M. H., ... & De Leo, G. A. (2016). Global assessment of schistosomiasis control over the past century shows targeting the snail intermediate host works best. *PLoS neglected tropical diseases*, 10(7).
- Standley, C., Boyce, M. R., Klineberg, A., Essix, G., & Katz, R. (2018). Organization of oversight for integrated control of neglected tropical diseases within Ministries of Health. *PLoS Neglected Tropical Diseases*, 12(11), e0006929.
- Steinmann, P., Keiser, J., Bos, R., Tanner, M., & Utzinger, J. (2006). Schistosomiasis and water resources development: systematic review, meta-analysis, and estimates of people at risk. *The Lancet Infectious Diseases*, 6(7), 411–425.
- Stelma, F. F., Talla, I., Verle, P., Niang, M., & Gryseels, B. (1994). Morbidity due to heavy Schistosoma mansoni infections in a recently established focus in northern Senegal. The American Journal of Tropical Medicine and Hygiene, 50(5), 575–579.
- Stothard, J. R., Chitsulo, L., Kristensen, T. K., & Utzinger, J. (2009). Control of schistosomiasis in sub-Saharan Africa: progress made, new opportunities and remaining challenges. *Parasitology*, 136(13), 1665–1675.
- Stothard, J. R., French, M. D., Khamis, I. S., Basáñez, M. G., & Rollinson, D. (2009). The epidemiology and control of urinary schistosomiasis and soil-transmitted helminthiasis in schoolchildren on Unguja Island, Zanzibar. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 103(10), 1031–1044.
- Stothard, J. R., Kabatereine, N. B., Tukahebwa, E. M., Kazibwe, F., Mathieson, W., Webster, J. P., et al. (2005). Field evaluation of the Meade Readview handheld microscope for diagnosis of intestinal schistosomiasis in Ugandan school children. *The American Journal of Tropical Medicine and Hygiene*, 73(5), 949– 955.
- Stothard, J. R., Rollinson, D., Imison, E., & Khamis, I. S. (2009). A spot-check of the efficacies of albendazole or levamisole, against soil-transmitted helminthiases in young Ungujan children, reveals low frequencies of cure. *Annals of Tropical Medicine and Parasitology*, 103(4), 357–603.
- Stothard, J. R., Sousa-Figueiredo, J. C., Betson, M., Green, H. K., Seto, E. Y., Garba, A., et al. (2011). Closing the praziquantel treatment gap: new steps in epidemiological monitoring and control of schistosomiasis in African infants and preschool-aged children. *Parasitology*, 138(12), 1593–1606.
- Strickland, G. T. (2006). Liver disease in Egypt: hepatitis C superseded schistosomiasis as a result of iatrogenic and biological factors. *Hepatology*, *43*(5), 915–922.
- Strong, R. P., & Shattuck, G. C. (1930). Splenomegaly. In R. P. Strong (Ed.), *The African Republic of Liberia and the Belgian Congo* (pp.214–223). Chapter 16, Volume 1. Cambridge, MA: Harvard University Press.
- Sturrock, R. F. (2001). The schistosomes and their intermediate hosts. In A. A. F. Mahmoud, (Ed.), *Schistosomiasis* (pp.7–83). London, UK: Imperial College Press.
- Takagi, H., Yahathugoda, T. C., Tojo, B., Rathnapala, U. L., Nagaoka, F., Weerasooriya, M. V., et al. (2019). Surveillance of Wuchereria bancrofti infection by anti-filarial IgG4 in urine among schoolchildren and molecular xeno monitoring in Sri Lanka: a post mass drug administration study. *Tropical Medicine and Health*, 47, 39.

- Talbot, J. T., Viall, A., Direny, A., de Rochars, M. B., Addiss, D., Mathieu, E. et al. (2008). Predictors of compliance in mass drug administration for the treatment and prevention of lymphatic filariasis in Leogane Haiti. *The American Journal* of Tropical Medicine and Hygiene, 78, 283–288.
- Tan, S Y., & Ahana, A. (2007). Theodor Bilharz (1825-1862): discoverer of schistosomiasis. Singapore Medical Journal, 48(3), 184–185.
- Theodor Bilharz as ethnographer and geographer. (1968). Bulletin of the New York Academy of Medicine, 44(3), 373–374.
- Thomas, J. E., Bassett, M. T., Sigola, L. B., & Taylor, P. (1990). Relationship between bladder cancer incidence, *Schistosoma haematobium* infection, and geographical region in Zimbabwe. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 84,551–553.
- Tukahebwa, E. M., Magnussen, P., Madsen, H., Kabatereine, N. B., Nuwaha, F., Wilson, S., et al. (2013). A very high infection intensity of *Schistosoma mansoni* in a Ugandan Lake Victoria fishing community is required for association with highly prevalent organ related morbidity. *PLoS Neglected Tropical Diseases*, 7(7), e2268.
- Ultroska, J. A., Chen, M. G., Dixon, H., Yoon, S., Helling-Borda, M., Hogerzeil, H. V., et al. (1989). An estimate of global needs for praziquantel within schistosomiasis control programmes. WHO/SCHISTO/89.102. Geneva: World Health Organization.
- Uniting to Combat Neglected Tropical Diseases. (2016). *Reaching the unreached: fourth progress report of the London Declaration*. Haywards Heath, UK: Uniting to Combat NTDs.
- Utzinger, J., Du, Z. W., Jiang, J. Y., Chen, J. X., Hattendorf, J., Zhou, H., et al. (2011). Efficacy of single-dose and triple-dose albendazole and mebendazole against soil-transmitted helminths and Taenia spp.: a randomized controlled trial. *PLoS ONE*, 6(9), e25003.
- Vandemark, L. M., Jia, T. W., & Zhou, X. N. (2010). Social science implications for control of helminth infections in Southeast Asia. Advances in Parasitology, 73, 137–170
- Van Oordt, B. E., van den Heuvel, J. M., Tielens, A. G., & van den Bergh, S. G. (1985). The energy production of the adult *Schistosoma mansoni* is for a large part aerobic. *Molecular and Biochemical Parasitology*, *16*(2), 117–126.
- Walsh, D. S., De Jong, B. C., Meyers, W. M., & Portaels, F. (2015). Leprosy and Buruli ulcer: similarities suggest combining control and prevention of disability strategies in countries endemic for both diseases. *Leprosy Review*, 86(1), 1–5.
- Webber, R. (1977). The natural decline of Wuchereria bancrofti infection in a vector control situation in the Solomon Islands. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 71(5),396–400.
- Weil, G. J. (1990). Parasite antigenemia in lymphatic filariasis. *Experimental Parasitology*, 71(3), 353–356.
- Weil, G. J., & Ramzy, R. M. (2007) Diagnostic tools for filariasis elimination programs. *Trends in Parasitology*, 23(2), 78–82.
- Weil, G. J., Kastens, W., Susapu, M., Laney, S. J., Williams, S. A., King, C. L., et al. (2008). The impact of repeated rounds of mass drug administration with diethylcarbamazine plus albendazole on Bancroftian filariasis in Papua New Guinea. *PLoS Neglected Tropical Diseases*, 2(12): e344.

- Weil, G. J., Lammie, P. J., & Weiss, N. (1997). The ICT Filariasis Test: a rapid-format antigen test for diagnosis of Bancroftian filariasis. *Parasitology Today* (*Personal Ed.*), 13(10), 401–404.
- Whitty, C. J., Mabey, D. C., Armstrong, M., Wright, S. G., & Chiodini, P. L. (2000). Presentation and outcome of 1,107 cases of schistosomiasis from Africa diagnosed in a non-endemic country. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 94(5), 531–534.
- Wiebe, A., Longbottom, J., Gleave, K., Shearer, F. M., Sinka, M. E., Massey, N. C., et al. (2017). Geographical distributions of African malaria vector sibling species and evidence for insecticide resistance. *Malaria Journal*, 16(1), 85.
- Wiest, P. M., Wu, G., Zhang, S., Yuan, J., Peters, P A. S., McGarvey, S. T., et al. (1992). Morbidity due to schistosomiasis japonica in the People's Republic of China. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 86, 47.
- Wijers, D. J. B. (2016). Bancroftian Filariasis in Kenya. I. Prevalence survey among adult males in the Coast Province. Annals of Tropical Medicine and Parasitology, 71(3), 313–331.
- Witt, C., & Ottesen, E. A. (2001). Lymphatic filariasis: an infection of childhood. *Tropical Medicine & International Health*, 6(8):582–606.
- World Bank. (2010). World Development Indicators 2010. Washington, DC: World Bank;
- World Health Organization (WHO), & Global Programme to Eliminate Lymphatic Filariasis (GPELF). (2005). *Monitoring and epidemiological assessment of the programme to eliminate lymphatic filariasis at implementation unit level.* Geneva: World Health Organization.
- World Health Organization (WHO). (1997). *Elimination of lymphatic filariasis as a public health problem*. Fiftieth World Health Assembly, WHA50.29. Geneva: World Health Organization.
- World Health Organization (WHO). (1999) Building partnerships for lymphatic filariasis: strategic plan. Geneva: World Health Organization. Retrieved January 14, 2014, from http://apps.who.int/iris/bitstream/10665/66252/1/WHO_FIL_99.198.
- World Health Organization (WHO). (2000a). Preparing and implementing a national plan to eliminate lymphatic filariasis (in areas where onchocerciasis is not coendemic): a guide for programme managers.
 WHO/CDSA/CPE/CEE/2000.15. Geneva: World Health Organization.
- World Health Organization (WHO). (2000b). *Operational guidelines for rapid mapping of Bancroftian filariasis in Africa*. WHO/CDS/CPE/CEE/20009. Geneva: World Health Organization.
- World Health Organization (WHO). (2001). Lymphatic filariasis. Weekly *Epidemiological Record*, 20(76), 149–156.
- World Health Organization (WHO). (2002). Defining the role of vector control and xenomonitoring in the global programme to eliminate lymphatic filariasis: report of the informal consultation held at WHO, Geneva. WHO/CDS/CPE/PVC/2002.3. Geneva: World Health Organization.
- World Health Organization (WHO). (2003). *Control of lymphatic filariasis in China*. Manila, Philippines: World Health Organization for the Western Pacific.

- World Health Organization (WHO). (2005). *Monitoring and epidemiological* assessment of the programme to eliminate lymphatic filariasis at implementation unit level. WHO/CDS/CPE/CEE/2005.50. Geneva: World Health Organization.
- World Health Organization (WHO). (2006). *Guidelines for the safe use of wastewater, excreta and greywater: Volume 4: excreta and greywater use in agriculture* (3rd Ed.). Geneva: World Health Organization.
- World Health Organization (WHO). (2006a). *Preventive chemotherapy in human helminthiasis: coordinated use of anthelminthic drugs in control interventions: a manual for health professional and programme managers.* Geneva: World Health Organization.
- World Health Organization (WHO). (2008). Training for Mid-level managers (MLM): Module 7: The EPI coverage survey. WHO/IVB/08.07. Geneva: World Health Organization. Retrieved January 10, 2020, from <u>https://apps.who.int/iris/bitstream/handle/10665/70184/WHO_IVB_08.07_eng</u>.pdf?sequence=7.
- World Health Organization (WHO) (2009). Distribution of Schistosomiasis worldwide. <u>https://www.google.co.in/search?q=gamapserver.who.int/maplibrary/files/map</u> <u>s/global_schistosomiasis_2009</u>
- World Health Organization (WHO). (2010a). Global programme to eliminate lymphatic filariasis: progress report on mass drug administration in 2009. *Weekly Epidemiological Record*, 38(85), 365–337.
- World Health Organization (WHO). (2010b). Lymphatic filariasis: eliminating one of humanity's most devastating diseases: action against worms. Newsletter 14. Geneva: World Health Organization.
- World Health Organization (WHO). (2010). Progress report 2000–2009 and strategic plan 2010–2020 of the global programme to eliminate lymphatic filariasis: halfway towards eliminating lymphatic filariasis. Geneva: World Health Organization. Retrieved January 14, 2014, from http://whqlibdoc.who.int/publications/2010/9789241500722_eng.pdf
- World Health Organization (WHO). (2010). Schistosomiasis: number of people treated, 2008. Weekly Epidemiological Record, 85(18), 157–164. Retrieved December 7, 2015, from <u>http://www.who.int/wer/en/</u>
- World Health Organization (WHO). (2011a). Accelerating work to overcome the global impact of neglected tropical diseases. Geneva: WHO. Retrieved February 4, 2020, https://www.who.int/neglected_diseases/NTD_RoadMap_2012_Fullversion.p df
- World Health Organization. (2011b). Lymphatic filariasis: monitoring and epidemiological assessment of mass drug administration: a manual for national elimination programmes. WHO/HTM/ NTD/PCT/2011 4: 1–79. Geneva: World Health Organization.
- World Health Organization (WHO). (2011c). Position statement on managing morbidity and preventing disability in the global programme to eliminate lymphatic filariasis. Geneva: World Health Organization. Retrieved January 14, 2014, from

http://whqlibdoc.who.int/hq/2011/WHO_HTM_NTD_2011.8_eng.pdf

- World Health Organization (WHO). (2012a). Accelerating work to overcome the global impact of neglected tropical diseases: a roadmap for implementation. Geneva: WHO. Retrieved June 20, 2015, from http://www.who.int/entity/neglected_diseases/NTD_RoadMap_2012_Fullvers_ion.pdf
- World Health Organization (WHO). (2012). Global programme to eliminate lymphatic filariasis: progress report, 2011. Weekly Epidemiological Record, 87, 346–356 Retrieved January 14, 2014, from <u>http://www.who.int/wer/2012/wer8737.pdf</u>
- World Health Organization (WHO). (2012). Soil-transmitted helminthiases: eliminating as public health problem soil-transmitted helminthiases in children: progress report 2001-2010 and strategic plan 2011-2020. Geneva: World Health Organization.
- World Health Organization (WHO). (2012). Transmission assessment surveys in the global programme to eliminate lymphatic filariasis: WHO position statement. Geneva: World Health Organization. Retrieved January 14, 2014, from <u>http://apps.who.int/iris/bitstream/10665/77690/1/WHO HTM NTD PCT 20</u> <u>12_9_eng.pdf</u>
- World Health Organization (WHO). (2013a). *Sixty-sixth World Health Assembly: resolutions and decisions*. Agenda item 16.2. WHA66/2013/REC/1. Geneva: World Health Organization.
- World Health Organization (WHO). (2013b). Lymphatic filariasis: training in monitoring and epidemiological assessment of mass drug administration for eliminating lymphatic filariasis: TAS learners' guide. Geneva: World Health Organization.
- World Health Organization (WHO). (2013). Lymphatic filariasis: a handbook for national elimination programmes. Geneva: World Health Organization.
- World Health Organization (WHO). (2013). Lymphatic filariasis: fact sheet No. 102.RetrievedAugust30,2013,http://www.who.int/mediacentre/factsheets/fs102/en/
- World Health Organization (WHO). (2013). Schistosomiasis: progress report 2001–2011, strategic plan 2012–2020. Geneva: World Health Organization.
- World Health Organization (WHO). (2014). Status of vaccine research and development of vaccines for schistosomiasis. Geneva: World Health Organization.
- World Health Organization (WHO) (2015) global programme to eliminate lymphatic filariasis: progress report, 2014. *Weekly Epidemiological Record, 38*(2015), 489–504.
- World Health Organization (WHO). (2016a). WHO global programme to eliminate lymphatic filariasis: progress report, 2015. *Weekly Epidemiological Record*, *39*(91), 441–460.
- World Health Organization (WHO). (2016) *African Programme for Onchocerciasis* control (APOC): Community-directed distributors (CDDs). Retrieved February 4, 2020 from https://www.who.int/apoc/cdti/cdds/en/.
- World Health Organization (WHO). (2017a). *Report of the Tenth Meeting of the WHO Strategic and Technical Advisory Group for Neglected Tropical Diseases.* Geneva: World Health Organization.

- World Health Organization (WHO). (2017b). Lymphatic filariasis fact sheet Updated March 2017, The World Health Organization
- World Health Organization (WHO). (2017c). Validation of elimination of lymphatic filariasis as a public health problem. Geneva: World Health Organization.
- World Health Organization (WHO). (2019). *Lymphatic filariasis*. Retrieved January 10, 2020, from <u>http://www.who.int/news-room/fact-sheets/detail/lymphaticfilariasis</u>
- Xiao S, H. (2013). Mefloquine, a new type of compound against schistosomes and other helminthes in experimental studies. *Parasitology Research*, *112*(11), 3723–3740.
- X. Wang, L. Zhang, R. Luo et al., (2012). Soil-transmitted helminth infections and correlated risk factors in preschool and school aged children in rural southwest China," *PLoS ONE*, vol. 7, no. 9, Article ID e45939.
- Yirga, D., Deribe, K., Woldemichael, K., Wondafrash, M., & Kassahun, W. (2010). Factors associated with compliance with community directed treatment with ivermectin for onchocerciasis control in south-western Ethiopia. *Parasites & Vectors*, 3, 48.
- Yosry, A. (2006). Schistosomiasis and neoplasia. *Contributions to Microbiology*, 13, 81–100.
- Young, M. D. (1953). Microfilariae and Trypansomes found in a blood survey of Liberia. Transactions of the Royal Society of Tropical Medicine and Hygiene, 47(4), 346–349.
- Zielke, E., & Chlebowsky, H. O. (1979). Studies on Bancroftian filariasis in Liberia, West Africa. I. Distribution and prevalence in the north-western savanna area. *Tropenmedizin und Parasitologie*, 30(1), 91–96.

APPENDICES

Appendix A: Questionnaire for the Knowledge, Attitudes and Practices towards SCH amongst school-age children in Bong County,

Liberia

| Name of School |
|--|
| Community |
| District |
| Initials of person doing the survey |
| GPS co-ordinate: LongitudeLatitude |
| Demographic characteristics |
| Date |
| ID number |
| Name of the respondent |
| Gender Male { } Female { } |
| Age Level of education: a) K1-Grade 2 b) Grade 3 to Grade 6 c) Grade 7 and above |
| 1. Knowledge of Schistosomiasis Yesif yes, what is your source of information? School Health workers Clinic/hospital Relatives Parents Town chiefs I don't have knowledge of schistosomiasis Others specify 2. Knowledge of sign and symptoms of Schistosomiasis Bloody urine (blood in your pee pee) Bloody stool (blood in your pop poo) |

| Abdominal pain |
|---|
| Diarrhoea |
| Headache |
| Fever |
| Chills |
| Itching skin/rash |
| 3. What germ causes schistosomiasis? |
| Worm |
| Fresh water |
| Snail |
| Other specify |
| Don't know |
| 4. Water contact pattern |
| Where do you get your water from a well/creek/river/hand pump? |
| Do you play in fresh water ponds or streams? |
| Where do you wash your cloths in the river/creek? |
| Do you pick up snails from the swamp? |
| Have you helped your parents with their rice farm? |
| Do you wash your cloths in the fresh water? |
| Where do you get your drinking water from? |
| 5. Presence of latrine Toilet in the house? |
| Go to the nearby toilet house? |
| Go to the bush to toilet? |
| Go to the waterside to toilet? |
| 6. Do you know how to prevent Schistosomiasis? |
| Avoiding unsafe drinking water |
| Washing hands before having a meal |
| Washing hands when leaving the bathroom Not |
| playing and swimming in unsafe water |
| Don't eat uncooked snail |
| Wash fruits and vegetables before eating |
| Taking the medications for schistosomiasis during MDA |
| 7. Mass Drug Administration |
| Do you know of any drug distribution in this community where people's height were |
| taken? |
| How many drugs did you receive? |
| Have you received the medication for schistosomiasis? |
| How many times have you received the schistosomiasis medication? Who |
| gave you the medication? |
| |

Appendix B: Assessment of Knowledge Awareness and Attitude to

MDA intervention for the control of LF, northern Liberia (Bong, Lofa,

and Nimba Counties): A post-Ebola survey

Name: Dr Louise Mapleh Kpoto, Tel: 0777702609, Health worker Name: Tel: ID number:

GIS co-ordinates: Latitude: Longitude:

A) Sociodemographic characteristics

1. Age

| 2. | Sex | Male | Female |
|----|--|-----------|--------|
| 3. | Marital status | | |
| | a) Married | | |
| | b) Single | | |
| | c) Divorced | | |
| | d) Widowed | | |
| | e) Cohabiting | | |
| 4. | Occupation | | |
| | a) Farmer | | |
| | b) Fisherman | | |
| | c) Hunter | | |
| | d) House wife | | |
| | e) Business personf) Others | | |
| 5. | Religion | | |
| | a) Christian | | |
| 6. | b) Muslim Level of education | | |
| | a) Have never had formal e | education | |

- b) Elementary
- c) Junior high

- d) High school
- e) Completed high school

B) Knowledge of the disease (you can tick more than one)

- 1. Have you heard of the disease LF a) Yes b) No
- 2. If yes, how does it present?
 - a) Enlarged leg
 - b) Enlarged scrotum
 - c) Enlarged breast
 - d) Fever and chills
 - e) Mumps
- 3. How does one get the disease?
 - a) Through traditional rituals
 - b) Through drinking dirty water
 - c) Through farming
 - d) Through mosquito bites
 - e) Don't know
 - d) Others

C) Prevention

- 1. Do you know how the disease is prevented? Yes NO
- a) By taking medication
- b) By drinking clean water
- c) By using mosquito nets
- d) By spraying your house
- e) Others

D) Drug distribution/compliance

- a) Has there been drug distribution in your community for LF? Yes or NO
- b) Did you take some of the medications? Yes or No

- c) How many times was the drug distributed? a) one b) two c) three d) four
- d) Did everyone in your house take the medications? Yes or No
- e) If no, why did they refuse the medication?
- f) How many tablets were given per person? a) one b) two c) three
- 1. What are the side effects of the drug?
 - a) Nausea
 - b) Vomiting
 - c) Fever and chills
 - d) Itching of skin
 - e) Impotency
 - f) Weakness/dizziness
 - g) Abdominal pains
 - h) Diarrhoea
 - i) Sweating
 - j) Blood in faeces
- 2. If these drugs were distributed would you take it again? Yes or No

Question on the Ebola

- 1. Were you or your family affected by the Ebola? Yes or No
- 2. If yes, were you in the Ebola Treatment Unit? Yes or No
- 3. Did you have a family member in the Ebola Treatment Unit? Yes or No
- 4. Did you have a friend in the Eboal Treatment Unit? Yes or No
- 5. Did you lose a friend to the Ebola? Yes or No
- 6. Did you lose a family member to the Ebola? Yes or No
- 7. During the Ebola did you have assess to health care? Yes or No
- 8. Do you know how Ebola is prevented? Yes or No
- 9. Select any answer
 - a) By washing hand
 - b) By avoiding others
 - c) By praying

- d) Don't know
- e) Others

10 Do you know how Ebola is acquired?

- a) Through food
- b) Through witchcraft
- c) Through hand shake
- d) By bodily fluids
- e) Don't know
- f) By touching others

Appendix C: A practical guide to the standardised use of ultrasonography for the assessment of SCH-related morbidity

See: www.who.int/schistosomiasis/resources/tdr_str_sch_00.1/en/

Appendix D: Informed Consent Form for lymphatic filariasis,

schistosomiasis prevalence study

Study participants ID#

Informed Consent

This Informed Consent Form is for men and women living in the targeted study villages who we are inviting to participate in research on the effectiveness of integrated mass drug administration in an environment that has not previously received treatment through community health workers. The title of our research project is: 'Integrated Approach to the Control of Lymphatic Filariasis, Schistosomiasis and Soil-Transmitted Helminthiasis in Liberia, West Africa'.

Principal investigators: Louise Kpoto

Liverpool School of Tropical Medicine, Liverpool, UK

This Informed Consent Form has two parts:

- Information Sheet (to share information about the research with you)
- Certificate of Consent (for signatures if you agree to take part)

You will be given a copy of the full Informed Consent Form

PART I: Information Sheet Introduction

Hello, my name is ______ and I am in from the Ministry of Health and Social Welfare. We are conducting a study about a disease called lymphatic filariasis (Guah), which are very common in this county. I am going to give you some information about the study and invite you to be part of this research. We would very much appreciate your participation; however, your participation is completely voluntary. Before you decide, you can talk to anyone you feel comfortable with about the research.

There may be some words that you do not understand. Please ask me to stop as we go through the information and I will take time to explain. If you have questions later, you can ask them of me or any of the other members of the research team.

Purpose of the research

Lymphatic filariasis is a common disease in this region, spread by mosquitoes. The infection is usually acquired in childhood; however, the painful and disfiguring visible manifestations are normally visible later on in life. Men suffering from the disease are commonly found with hydrocoeles, while both men and women will suffer with

enlarged and inflamed limbs. The treatments for this disease are two tablets that will be provided by your community health volunteer once a year for everyone in the village, including children. For effective treatment it is important to take the tablets every year.

We want to determine if you are affected with this disease. Results from the research will then be used by the Ministry of Health in providing treatment for children in this district.

Participant selection

We are inviting healthy adults living in your village to participate in this study.

Voluntary participation

Your participation in this research is entirely voluntary. It is your choice whether to participate or not. You may change your mind later and stop participating even if you agreed earlier.

Procedures and protocol

If you agree to participate, we will first ask you some basic questions about yourself and the people living in your household, such as your age, the location of your home, how long you have lived in this community, and the type of methods you use in your household to control mosquitoes.

Lymphatic filariaisis

We will take blood from your finger using a lancet. The blood will be tested to determine if you have been exposed to lymphatic filariasis and if you have active disease.

We will first clean the finger to be pricked with an alcohol swab and allow the finger to dry. We will then prick the finger with a sterile lancet and a small amount of your blood will be placed on a test card (show a test card). We will share the results with you.

We will also be conducting an additional test for detecting lymphatic filariasis; where a very small volume of blood will be collected from another finger approximately 2 mL taken at night between 22:00 hours and 02:00 hours will be mixed with approximately 10 times its volume of distilled water in a simple slide counting chamber and examine the slide immediately. We will also share these results with you.

The blood obtained during this research procedure will be used only for this research. At the end of the research, in 2 years, any leftover blood sample will be destroyed.

Duration

Our research study will take about 2 years to complete. During this time, we will return to your community to repeat the blood testing on a random sample of the community about once per year. When we return, you may or may not be included again in the study. Your participation at this time will take place over the next 2 days. During this time, we will ask you some simple questions about your household using a questionnaire. This should take approximately 10 minutes. We will then conduct the blood test.

Side effects

Potential side effects are limited. You may experience some mild discomfort at the site of the blood drawn from your finger.

Benefits

There will not be any monetary benefit for you but your participation is likely to help us find the answer to the research question and if people in this county are found with the disease they will receive treatment through Mass Drug Administration.

Confidentiality

With this research, something out of the ordinary is being done in your community. It is possible that if others in the community are aware that you are participating, they may ask you questions. We will not be sharing the identity of those participating in the research.

The information that we collect from this research project will be kept confidential. Information about you that will be collected during the research will be put away and no-one but the researchers will be able to see it. Any information about you will have a number on it instead of your name. Only the researchers will know what your number is and we will lock that information up with a lock and key. It will not be shared with or given to anyone except the senior members of our research team.

Sharing the results

The knowledge that we get from doing this research will be shared with you through community meetings before it is made widely available to the public. Confidential information will not be shared. There will be small meetings in the community and these will be announced. After these meetings, we will publish the results in order that other interested people may learn from our research.

Right to refuse or withdraw

You do not have to take part in this research if you do not wish to do so. You may also stop participating in the research at any time you choose. It is your choice and all of your rights will be respected.

Who to contact

If you have any questions you may ask them now or later, even after the study has started. If you wish to ask questions later, you may contact any of the following: Dr Louise Kpoto MoH&SW, Tel: 06514469; Mr Jerry Diaboi Tel: 06543567.

This proposal has been reviewed and approved by the University of Liberia, which is a committee whose task it is to make sure that research participants are protected from harm. If you wish to find about more about the University of Liberia Institutional Review Board, contact: Mr Jamee, University of Liberia Tel: 06583774.

PART II: Certificate of consent

I have read the foregoing information, or it has been read to me. I have had the opportunity to ask questions about it and any questions that I have asked have been answered to my satisfaction. I consent voluntarily to participate as a participant in this research.

| Print Name of Participant | |
|--|--|
| Signature of Participant | |
| Date | |
| $D_{a} = \sqrt{M_{a}} + 41 \sqrt{M_{a}}$ | |

Day/Month/Year

If illiterate

I have witnessed the accurate reading of the consent form to the potential participant, and the individual has had the opportunity to ask questions. I confirm that the individual has given consent freely.

| Print name of witness | AND | Thumb print of | |
|-----------------------|-----|----------------|--|
| participant | | | |
| Signature of witness | | | |

Date _____

Day/Month/Year

Statement by the researcher/person taking consent

I have accurately read out the information sheet to the potential participant, and to the best of my ability made sure that the participant understands the study procedures.

I confirm that the participant was given an opportunity to ask questions about the study, and all the questions asked by the participant have been answered correctly and to the best of my ability. I confirm that the individual has not been coerced into giving consent, and the consent has been given freely and voluntarily.

A copy of this Informed Consent Form has been provided to the participant.

| Print Name of researcher/person taking the consent |
|--|
| Signature of researcher /person taking the consent |
| Date |

Day/Month/Year