# Drug-specific T-cell responses in patients with liver injury following treatment with the BACE inhibitor atabecestat.

**Short title:** T-cell responses to drugs

**Authors:** Paul J Thomson1, Laila Kafu1, Xiaoli Meng1, Jan Snoeys2, An De Bondt,2 Dries De Maeyer,2 Hans Wils,2 Laurent Leclercq2, Petra Vinken3, Dean J Naisbitt1\*.

1MRC Centre for Drug Safety Science, Department of Molecular and Clinical Pharmacology, Sherrington Building, Ashton Street, University of Liverpool, Liverpool, UK;

2Drug Metabolism and Pharmacokinetics, Janssen R&D, Belgium;

3Non-Clinical Safety, Janssen R&D, Belgium

**\*Corresponding author:**  Professor Dean J. Naisbitt, MRC Centre for Drug Safety Science, Department of Molecular and Clinical Pharmacology, Sherrington Building, Ashton Street, The University of Liverpool, Liverpool L69 3GE, England

Telephone: 0044 151 7945346; Fax: 0044 151 7945540; e-mail: dnes@liverpool.ac.uk

**Word count:** 3499

## Abstract

**Background:** Atabecestat is an orally administered BACE inhibitor developed to treat Alzheimer’s disease. Elevations in hepatic enzymes were detected in a number of in trial patients, which resulted in termination of the drug development programme. Immunohistochemical characterization of liver tissue from an index case of atabecestat-mediated liver injury revealed an infiltration of T-lymphocytes in areas of hepatocellular damage. This, coupled with the fact that liver injury had a delayed onset suggests that the adaptive immune system may be involved in the pathogenesis. The aim of this study was to generate and characterise atabecestat(metabolite)-responsive T-cell clones from patients with liver injury.

**Methods:** Peripheral blood mononuclear cells were cultured with atabecestat and its metabolites (diaminothiazine [DIAT], *N*-acetyl DIAT & epoxide) and cloning was attempted in a number of patients. Atabecestat(metabolite)-responsive clones were analysed in terms of T-cell phenotype, function, pathways of T-cell activation and cross-reactivity with structurally-related compounds.

**Results:** CD4+ T-cell clones activated with the DIAT metabolite were detected in 5 out of 8 patients (up to 4.5% cloning efficiency). Lower numbers of CD4+ and CD8+ clones displayed reactivity against atabecestat. Clones proliferated and secreted IFN-γ, IL-13 and cytolytic molecules following atabecestat or DIAT stimulation. Certain atabecestat and DIAT-responsive clones cross-reacted with *N*-acetyl DIAT; however, no cross-reactivity was observed between atabecestat and DIAT. CD4+ clones were activated through a direct, reversible compound-HLA class II interaction with no requirement for protein processing.

**Conclusion.** The detection of atabecestat metabolite-responsive T-cell clones activated via a pharmacological interactions pathway in patients with liver injury is indicative of an immune-based mechanism for the observed hepatic enzyme elevations.

**Key words:** Drug-induced liver injury, immune system, T-lymphocytes, human.

## Introduction

Alzheimer’s Disease (AD) is a chronic neurodegenerative disease which is characterised by an impairment in cognitive function beyond the scope of the normal ageing process. An imbalance exists between production and clearance of amyloid-β in patients with AD and a common disease characteristic is the presence of neurofibrillary tangles comprised of hyperphosphorylated tau proteins and amyloid plaques (1). Amyloid-β is produced from amyloid precursor protein via a cleavage process by BACE-1 in the first and rate limiting step. Therefore, it is hypothesised that inhibition of the BACE-1 enzyme can reduce the formation of the toxic amyloid plaques, thereby having a positive impact on the progression of AD. Development of atabecestat (JNJ-54861911), an orally administered BACE-1 inhibitor, for the treatment of AD was recently stopped due to elevations in hepatic enzymes in clinical studies (2).

Elevated hepatic enzymes were observed in patients participating in two atabecestat studies with 24% of patients exhibiting >1.5 upper limit of normal (ULN) levels of alanine transaminase (ALT), including 11% with ALT > 3x ULN. These were commonly associated with increases in aspartate aminotransferase (AST), but only 1 patient had a concomitant increase of bilirubin > 2x ULN in the presence of a normal alkaline phosphatase, indicative of severe hepatocellular injury. Importantly, increases in the level of ALT did not correlate with the concentration of atabecestat (metabolites) in patient plasma. A swift decline in ALT levels was observed in most patients following withdrawal of atabecestat; however, ALT levels continued to increase in the plasma of a limited number of patients. In particular, one patient reported an ALT level of 28.8x ULN one month after cessation of therapy. The delayed onset of liver injury and immuno-histological detection of T-cell infiltrates in a liver biopsy specimen (3) suggests that the adaptive immune system, might be involved in the pathogenesis of the atabecestat-induced adverse event.

Until recently, the involvement of the adaptive immune system in drug-induced liver injury (DILI) was controversial. Maria and Victorino detected drug-responsive T-cells within peripheral blood mononuclear cells (PBMC) from certain patients with DILI (4). However, the drug-responsive T-cells were never characterized in terms of phenotype or function. More recently, we have shown the activation of drug-responsive T-cells from the blood of patients with flucloxacillin-(5), amoxicillin-clavulanate- (6) and tuberculosis medication-induced liver injury (7). Moreover, several HLA allele associations have been identified in patients with DILI for a host of drugs including flucloxacillin (HLA-B\*57:01) (8), ximelagatran (HLA-DRB1\*07:01) (9) and lumiracoxib (HLA-DRB1:15:01) (10) and in patients with drug rash with eosinophilia and systemic symptoms (DRESS; e.g., vancomycin (11) and dapsone (12)) where liver injury develops as a component of a multifaceted disease. Although not always proven through T-cell studies with patient PBMC, the genetic associations suggest that drugs interact preferentially with the protein (or binding peptide) derived from the HLA of interest to stimulate T-cells that contribute towards the disease pathogenesis. HLA molecules displayed on the cell surface present drugs bound directly via a reversible pharmacological interaction (p-I concept) and covalently bound drug peptide adducts generated through protein processing (hapten concept).

The aim of this study was to elucidate whether atabecestat-responsive T-cells were detectable in atabecestat trial patients that developed elevations in liver enzymes and then generate T-cell clones to characterize phenotype and function of the drug-responsive cells along with pathways of T-cell activation and cross-reactivity with related structures. Patient PBMC were cultured with synthetic forms of atabecestat, diaminothiazine [DIAT], and atabecestat epoxide, each of which are susceptible to cytochrome P450-mediated metabolism liberating products with the potential to bind covalently to protein (Figure 1A). DIAT is formed from atabecestatvia the carboxylesterase-mediated hydrolysis of the amide bond resulting in loss of the cyano-pyridine ring structure. In contrast, the epoxide of atabecestat is formed on the amino-methyl-thiazinyl ring through cytochrome P450-3A mediated metabolism.

## Methods

**PBMC from human subjects**

PBMC were obtained from 14 study patients previously exposed to atabecestat (clinical studies 54861911ALZ2002, 54861911ALZ2003, and 54861911ALZ2004), of which 12 out of 14 had experienced liver ALT and AST elevations during therapy. The ALT increase in the selected patients ranged between 3.8 and 29.1x ULN. Table 1 summarizes the demographics of the patients and details of the adverse event. None of the patients showed skin symptoms or eosinophilia. PBMC from two healthy volunteers outside of the atabecestat studies were selected from the Liverpool cohort of 1200 genotyped healthy donors (13). Approval for the investigations was obtained from the Liverpool Research Ethics committee and each site’s respective Ethic Committee, and informed written consent was obtained from each donor. Chemagic magnetic separation (Chemagen, Baesweiler, Germany) was used to extract genomic DNA, and high-resolution sequence based HLA typing was performed by the Histogenetics laboratory (Histogenetics, Ossining, NY) at the following loci: HLA-A, -B, -C, -DRB1, -DQB1, and DQA1. PBMC isolated from blood using density centrifugation were cryopreserved on the day of isolation and stored in liquid nitrogen prior to conducting the experiments. Supplementary Table 1 shows results of HLA typing.

PBMC were subjected to *in vitro* diagnostic testing and cloning of individual T cells before characterization of cellular phenotype, TCR sequencing, function, antigen specificity and pathways of activation. Detailed methods are available in as supplementary material.

## Results

### Atabecestat and its metabolites stimulate low levels of proliferation and granzyme B secretion in certain patients with liver injury

The lymphocyte proliferation assay was conducted in 10 patient PBMCs while ELIspot for IFN-γ and granzyme B was conducted on 9/14 patients (this included one patient not presenting ALT elevations; Table 1) to assess the presence of drug-responsive circulating T-cells in patients with liver injury. PBMCs from 4 subjects were not viable upon defrosting of vials, while cellular yield was low in 1 subject, excluding them from ELIspot assays.

Low levels of proliferation (stimulation index [proliferation in compound treated wells/ proliferation in medium wells] 2-2.5) were induced by the DIAT and epoxide metabolites in patient 4; however, based on the low cpm values of the control condition and the size of the error bars, it is likely this was a false positive. All other subjects displayed no proliferation in the presence of any of the study compounds. Strong T-cell proliferation was induced with PBMC from all donors by the positive controls PHA and tetanus toxoid (data not shown).

Secretion of granzyme B was observed in response to the DIAT metabolite in patient 6, but no other compounds (Figure 1B). IFN-γ secretion was not detected. Furthermore, no secretion of IFN-γ or granzyme B was observed from PBMC of other patients with any of the study compounds. Strong secretion of IFN-γ and granzyme B was observed in response to PHA. Finally, no proliferation or secretion of IFN-γ or granzyme B was observed in healthy volunteers in response to atabecestat, DIAT and atabecestat epoxide.

**Atabecestat and its metabolites activate T-cell clones from patients with liver injury**

T-cell clones were generated from patients with liver injury to assess cellular phenotype, drug cross-reactivity and pathways of drug presentation. During initial proliferation testing of clones with medium and study compounds, atabecestat (10 μM), DIAT (25 μM), acetyl DIAT (25μM) and atabecestat epoxide (10μM), using duplicate cultures, responsive T-cell clones were generated to atabecestat from patients 4, 5 and 6, while DIAT responsive T-cell clones were generated from patients 1, 5, 6, 9 and 12 (Supplementary Table 2). Acetyl-DIAT responsive clones were only generated from patient 6, while weakly- atabecestat epoxide-responsive clones were observed in patients 4, 6 and 12.

Confirmation of the responsiveness of T-cell clones after the first testing stage was via a dose response study at compound concentrations of 0-100 µM (duplicate conditions per culture condition). Four, ten and three T-cell clones were stimulated to proliferate in a dose-dependent manner with atabecestat, DIAT and acetyl DIAT, respectively (Supplementary Figure 1A). The proliferative response of all clones generated from patient 6 is shown in Figure 1C. Treatment of the clones with atabecestat resulted in a reverse dose-response (lower proliferation at higher concentrations). This was not associated with atabecestat-induced T-cell toxicity; furthermore, concentrations of 10 µM and below did not lead to significant levels of proliferation (results not shown). No responsive T-cell clones were identified to atabecestat epoxide. The total number of study compound-responsive T-cell clones and percentage of responsive clones generated during initial testing and after dose-responsive assessment from each patient is summarised in Supplementary Table 2.

### Phenotype of study compound-responsive T-cell clones

With the exception of one CD8+ T-cell clone (patient 6; atabecestat-responsive) and one T-cell clone which was characterised as double positive (patient 5; DIAT-responsive) all responsive T-cell clones to atabecestat, DIAT and acetyl DIAT were phenotyped as CD4+ (Supplementary Figure 1B). Across the T-cell clones tested, a varied expression of TCR-Vβ was detected to the parent drug and the DIAT metabolite, with expression of rare TCRs (not detected by the antibodies in the kit) being predominantly expressed (Supplementary Figure 1C). Chemokine receptor expression was assessed using six atabecestat- or DIAT-responsive clones expressing a single TCR. All tested T-cell clones expressed CXCR3, while the early activation marker CD69 and CCR4 were expressed on 5/6 clones. E-cadherin was expressed across some of the clones (Supplementary Figure 1D). CXCR3 and CCR4 are involved in the recruitment of Th1 and Th2 cells (14, 15), respectively.

**TCR sequencing**

For sequencing analysis, all TCRα and TCRβ CDR3 sequences with a fraction >5% of the total reads were retrieved with a minimum total of >500 reads. Assuming these are indeed true single cell clones, we would expect only one functional TCRβ CDR3 sequence in combination with maximum 2 functional TCRα CDR3 sequences for each T cell clone. TCR sequencing performed on 6 atabecestat- or DIAT-responsive T-cell clones in total from 2 different patients revealed this was indeed the case for 4 clones (DIAT clones 1-3 and atabecestat clone 2). Two clones displaying 2 TCRβ CDR3 sequences were detected suggesting that the expansion was not originating from a single clone. Nevertheless, the data from all 6 clones clearly shows that atabecestat- and DIAT-responsive T-cells express different TCRβ clonotypes and TCRα and β pairings. An overview of the T-cell clones that were sequenced is given in Supplementary Table 3.

### Study compound-responsive T-cell clones exhibit distinct cytokine and cytolytic molecule secretion profiles

Secretion of cytokines and cytolytic molecules from one atabecestat- and five DIAT-responsive T-cell clones was assayed by ELIspot. Secretion of cytokines IFN-γ and IL-13 were detected from all clones. Three clones secreted IL-10 and IL-5, while low levels of IL-22 was also detected in three clones. IL-17 secretion was not detected. Secretion of granzyme B and perforin was detected following study compound treatment (Figure 2). Low levels of killing (2.4-10.8%) of [51Cr]-loaded autologous antigen presenting cells was observed with 3 study-compound treated clones (effector to target ratio 10:1; data not shown).

### Direct, reversible and HLA-DR-restricted activation of study compound-responsive CD4+ T-cell clones

To assess HLA dependency and class restriction of the CD4+ T-cell clone responses, HLA blocking studies were conducted. All CD4+ clones responsive to atabecestat and DIAT demonstrated HLA-DR dependency for T-cell activation (Figure 3). The pathway of atabecestat- and DIAT-specific T-cell activation was assessed using antigen presenting cell pulsing and glutaraldehyde fixation assays. T-cell clones exhibited no T-cell activation in the presence of antigen presenting cells pulsed with study compounds for 1 or 16 hours. The small increase in proliferation observed with antigen presenting cells pulsed with atabecestat for 1 h was not significant (P=0.17; student T-test). Conversely, T-cell activity was observed, in a reduced capacity, to atabecestat and DIAT in the presence of glutaraldehyde-fixed antigen presenting cells, but not in the absence of antigen presenting cells (Figure 4).

### Study compound-responsive T-cell clones display clear patterns of cross-reactivity

The cross-reactivity profiles of T-cell clones was measured via co-incubation with EBV-transformed B-cells and the 4 study compounds and assessment of proliferative responses or IFN-γ release. All T-cell clones generated to atabecestat yielded no crossreactivity with DIAT, while 3/4 clones displayed proliferative responses with acetyl DIAT. A weak proliferative responses was also observed with atabecestat epoxide in one clone. DIAT-responsive T-cell clones exhibited no cross-reactivity with atabecestat or atabecestat epoxide. However, 3/6 DIAT-responsive T-cell clones from various patients exhibited cross-reactivity towards acetyl DIAT. Figure 5 shows representative clones displaying the different cross-reactivity profiles (atabecestat-specific, DIAT-specific and cross-reactive).

Figure 5 also shows the preferred energy minimized 3-dimensional spatial arrangement of atabecestat and its metabolites. The structures of atabecestat, DIAT and acetyl DIAT, with the exception of the side-chain moieties, were almost identical. However, the 3-dimensional spatial arrangement of atoms within atabecestat epoxide differed significantly due to the introduction of the 3-membered ring.

All three acetyl DIAT-responsive clones exhibited strong cross-reactivity towards the DIAT metabolite. One of these clones also cross-reacted with atabecestat (Supplementary Figure 2).

## Discussion

Several forms of DILI are strongly associated with expression of one or a small number of HLA alleles (8-12, 16-18), suggesting that a derivative of the drug may interact with a degree of selectivity with the HLA protein encoded by the HLA risk allele and a peptide embedded within the antigen binding cleft of HLA to stimulate the T-cell response. Furthermore, drug-responsive T-cells have been detected in the circulation and inflamed liver of patients with DILI (5-7, 19, 20). Thus, it is becoming apparent that the adaptive immune system participates in the development of tissue pathology in genetically predisposed individuals. It must be emphasised that an HLA allele association only indicates a skewing in the preference for drug HLA-peptide binding. There are many cases of immune-mediated DILI in patients that do not express HLA risk alleles Furthermore, the majority of individuals that express HLA risk alleles do not go on to develop liver injury when exposed to culprit drugs. The best example of this is flucloxacillin-induced liver injury that preferentially develops in patients expressing HLA-B\*57:01; however, the iatrogenic disease is only observed in 1 in 1000 HLA-B\*57:01+ individuals exposed to the drug (8). All individuals are exposed to similar circulating concentrations of flucloxacillin and form the same protein adducts at levels that exceed the threshold for T-cell activation (5, 21-25). Hence, additional factors must impact on susceptibility.

In this study we focused on the therapeutic atabecestat, which functioned to reduce the synthesis of amyloid plaques, a significant feature of AD. Phase I studies were conducted to atabecestat in amyloid positive Caucasian and Japanese patients. Four-week treatment with atabecestat resulted in a mean reduction of 67% in cerebrospinal fluid-Aβ1-40 in both the Caucasian and Japanese patients, with a reduction of up to 90% in some patients (26). Following this, phase 2/3 studies were conducted, with a plan to enrol up to 1650 patients at risk of dementia with a primary endpoint being the slowing of cognitive decline in those patients receiving either 10 mg (decreased to 5mg after 11 participants received 10 mg to increase safety margin between low and high doses) or 25 mg of the drug compared to placebo. However, in May 2018 the atabecestat program was discontinued based on the detection of elevated liver enzymes. Of the 475 patients exposed to atabecestat, approximately 24% had ALT/AST of >1.5 x ULN, and 11% had ALT elevations >3x ULN. The evolution of these elevations was heterogeneous and some elevations were transient while patients remained on treatment. The patients with liver injury showed no signs of skin rash and/or eosinophilia. Interestingly, the dose of atabecestat (10-25 mg) is relatively low for a DILI causing drug and a genetic association with HLA has not been identified. Furthermore, the time to onset of raised liver enzymes (32-344 days; Table 1) is significantly longer than that associated with skin reactions such as maculopapular exanthema. Although the factors responsible for breaking tolerance after several months of drug therapy are not known, the expression and activity of immune co-inhibitory receptors such as PD-1 and CTLA-4 is an area of considerable interest. Through *in vitro* assessment of drug-specific T-cell priming, we have shown that the level of T-cell activation is significantly enhanced when immune regulation has been dysregulated with immune checkpoint inhibitors (27, 28). Clinical studies using a low molecular weight drug alongside an immune checkpoint inhibitor are also beginning to highlight the importance of immune dysregulation on susceptibility to DILI. For example, dacarbazine a relatively safe imidazole carboxamide chemotherapy drug, was not tolerable in a Japanese phase II study when combined with ipilimumab, a monoclonal antibody that targets CTLA-4. Twelve out of fifteen patients developed elevations in ALT and aspartate aminotransferase (29). Hence, it is possible that a natural imbalance in immune regulatory factors mediated by disease or environmental factors may be an important determinant of susceptibility. Alternatively, a low drug-responsive T-cell precursor frequency at the time of initial drug exposure might explain the delayed time to onset, but not the variable time to onset.

To investigate the involvement of the immune system in subjects who experienced elevated liver enzymes whilst receiving atabecestat, PBMC samples were collected, and T-cells were cloned from atabecestat- and atabecestat metabolite-exposed PBMC lines for phenotypic and functional assessment. HLA-genotyping was conducted on 4 of the patients PBMC that were subjected to T-cell cloning. Genotyping identified the common expression of HLA-DRB1\*04:01 and DRB1\*15:01 in 3 patients suggesting that if CD4+ T-cell responses were detected they may involve the drug interacting selectively with the proteins encoded by these alleles.

On the whole diagnostic assays conducted on the PBMCs of patients were negative by way of proliferation and secretion of cytokines and effector molecules. One exception was patient 6, which noted a subtle increase in secretion of granzyme B in response to the primary metabolite of atabecestat; DIAT. Patient 6 showed persistent liver enzyme elevations even one month after cessation of atabecestat treatment. Furthermore, a liver biopsy from this patient indicated a zone 3-predominant necroinflammatory process, with lymphocyte infiltration (3).

Eight patient PBMC were subjected to T-cell cloning. Study compound-responsive clones were generated from five of these patients with the greatest number stemming from patient 6. Generation of atabecestat-responsive T-cell clones was much more uncommon than seen with the DIAT metabolite, which may suggest a lower T-cell pre-cursor frequency. Furthermore, no confirmed T-cell clones were generated to the minor epoxide metabolite of atabecestat suggesting that if atabecestat epoxide-responsive clones exist, their precursor frequency must be very low. In total atabecestat-responsive clones were generated from two of the patients PBMC to undergo T-cell cloning. One of the atabecestat-responsive clones exhibited a CD8+ phenotype in contrast with all other generated clones which were CD4+. DIAT-responsive clones detected in 5 patients all displayed a CD4+ phenotype. Clones were activated with study compounds at concentrations of 10 µM and above. Steady-state human atabecestat plasma levels are reached by day 5 in line with the plasma t1/2 of 15-19h. At 25 mg qd atabecestat dosing a mean plasma Cmax of 0.7 µM was observed (26, 30). All circulating human metabolites showed a similar plasma half-life as atabecestat. The most abundant human plasma metabolite was the DIAT metabolite which reached a steady state Cmax of 0.05 µM. Although atabecestat drug-related material was not quantified in human liver samples, an atabecestat pharmacokinetic study was conducted in TK-NOG humanized chimeric liver mice showing 15-fold higher atabecestat drug-related material concentrations in humanized liver versus plasma. The DIAT metabolite concentrations were on average 12% of the atabecestat concentrations in liver extracts from this study. Collectively, this indicates that the concentration of atabecestat in plasma is lower than that needed to activate T-cells; however, T-cell stimulatory concentrations may be reached when the drug accumulates in liver. Atabecestat- and DIAT-responsive clones exhibited a varied expression of TCR-Vβ with the majority expressing a rare TCR not detected by the antibody repertoire. Given the differential reactivity profiles of the clones it is unlikely that TCR expression is conserved across the clones. For example, two DIAT responsive T-cell clones expressing rare TCR-Vβs exhibited cross-reactivity towards other study compounds, while two additional clones from the same patient exhibited no cross-reactivity.

Stimulation of clones with atabecestat or DIAT resulted in the secretion of the Th1 cytokine IFN-γ and Th2 cytokines IL-5 and IL-13. Furthermore, IL-10 and IL-22 were secreted from a more restricted number of study compound activated clones. Secretion of Th1 and Th2 cytokines, alongside IL-22 is a common feature for drug-responsive clones isolated from patients presenting with skin and liver conditions (6, 21, 31). IL-10 secretion is observed less frequently. IL-10 is predominantly immunoregulatory, ameliorating excessive and potentially pathogenic Th1 and CD8+ T-cell responses (32). The activated clones, which for the most part were CD4+, also secreted effector molecules perforin and granzyme B. These molecules act together, with perforin inserting pores into neighbouring cells to permit the delivery of granzyme B, which cleaves caspases triggering the apoptotic cascade.

All tested atabecestat- and DIAT-responsive CD4+ clones were activated in a manner restricted to a MHC Class II protein, namely HLA-DR. As discussed above, clones were generated from three patients that expressed the HLA-DR\*B alleles 04:01 and 15:01. However, genotyping conducted on patient 6, the patient yielding the largest number of study compound-responsive clones and exhibited the most serious clinical symptoms, indicated the expression of HLA-DR\*B alleles 12:01 and 13:02. This suggests that T-cell responses to atabecestat and DIAT in subjects are not restricted to the expression of one single HLA-allele. Future experiments are planned using antigen presenting cells from donors expressing different HLA alleles to define the HLA-DR molecules involved in presentation of atabecestat and its metabolites to T-cells. With this knowledge, antigen presenting cell pulsing experiments were conducted to characterise pathway(s) of atabecestat and DIAT-specific CD4+ T-cell activation. Antigen presenting cells were pulsed with study compounds for 1 or 16 hours prior to repeated washing to remove soluble and weakly associated compound prior to culturing the antigen presenting cells with T-cells. Antigen presenting cells pulsed with protein-reactive compounds (e.g., flucloxacillin, nitroso sulfamethoxazole (5, 33, 34)) or compounds that associate strongly with HLA (e.g., abacavir (35-37)) activate T-cell responses, while compounds that associate weakly with HLA via reversible binding interactions (e.g., carbamazepine (34, 38-40)) do not. All atabecestat- or DIAT-responsive clones were not activated with study compound-pulsed antigen presenting cells. This indicates that the clones are stimulated with atabecestat and DIAT via a non-covalent pharmacological binding interaction with HLA-DR molecules. Activation of T-cells in the presence of glutaraldehyde fixed antigen presenting cells was also observed, which is indicative of a direct interaction of atabecestat and DIAT with HLA-DR molecules in a manner independent of antigen processing. Fixation blocks protein processing and inhibits T-cell responses to protein antigens, but not antigenic peptides that bind directly to surface HLA molecules (38-43). Collectively, these findings are in agreement with the p-I concept proposed by Pichler (44). In essence, atabecestat and its metabolites have an undesirable (off-target) pharmacological interaction with immune receptors. The p-I concept also argues that certain drugs may bind initially to highly variable TCRs and that HLA proteins provide additional signals for T-cell activation. Identification of a drug-specific public TCR that drives severe carbamazepine-induced cutaneous reactions provides strong support for this concept. TCR sequencing was performed using 6 atabecestat or DIAT-responsive clones from 2 patients to determine whether a single rare TCR was identifiable. Although our dataset was somewhat limited, it clearly shows that the drug-responsive clones express different TCRβ clonotypes and TCRα and β pairings. Given the low circulating concentrations in human plasma and the absence of an association of liver injury with an HLA allele, atabecestat and its metabolites must bind with high affinity with multiple HLA proteins to activate the patient T-cells. Future research should attempt to determine whether there is a similar drug binding site which is common to the presenting HLA proteins.

Atabecestat- and DIAT-responsive and clones displayed either no cross-reactivity or cross-reacted with acetyl DIAT (Figure 4). Based on these findings, clones were generated from PBMC cultured with acetyl DIAT for further assessment of cross-reactivity. Interestingly, acetyl DIAT responsive clones displayed cross-reactivity with DIAT and in one clone atabecestat and DIAT (Supplementary Figure 2). Since all of the study compounds activate the T-cells through a non-covalent pharmacological binding interaction with HLA, with no requirement for protein processing, the similar spatial arrangement of atoms between atabecestat, DIAT and acetyl DIAT provides one explanation for the observed reactivity profiles of the clones. The acetyl-cyanopyridine structure that differentiates atabecestat from DIAT, must provide an essential HLA peptide binding interaction for the activation of atabecestat-responsive clones, whereas with DIAT-responsive clones sufficient binding energy must be provided by the remainder of the drug structure. Acetyl DIAT represents an intermediate structure, which explains its capacity to activate certain atabecestat- and DIAT-responsive clones. Very little reactivity was observed towards atabecestat epoxide, which exists in a very different confirmation (Figure 4).

We must consider the fact that these data are taken from a relatively small number of T-cell clones; nevertheless, our findings show that drug-responsive T‑cells with specificity to atabecestat and its DIAT metabolite are present in patients with atabecestat-induced liver injury. Thus, it is likely that the DILI reactions experienced by these patients while on treatment with atabecestat were mediated at least in part by the adaptive immune system, with CD4+ responses being the primary action. Furthermore, it is probable that such reactions were mediated to a greater degree by the DIAT metabolite of atabecestat. Future experiments profiling gene expression in drug-responsive T-cell clones and T-cell infiltrating liver of the same patients will provide direct evidence that circulating atabecestat- and/or DIAT-responsive T-cells are responsible for the tissue injury.

**Acknowledgements:** The authors would like to thank the patients and volunteers for their generous donations.

**Funding:** PJT is a post-doctoral researcher that received funds from Janssen R&D to support his studies. The project also received funding from the MRC Centre for Drug Safety Science (Grant number G0700654) and an MRC project grant (Grant number MR/R009635/1).

**Author contributions:** PJT, LK, and XM conducted the biological experiments. LL, PV and JS designed the clinical protocols and collected the patient samples. ADB, DDM and HW conducted the TCR sequencing experiments. DJN, JS and PV designed the research study and supervised the project. PJT, LK and DJN analysed the data and drafted the manuscript. All authors critically reviewed the manuscript.

**Conflicts of interest:** The authors declare no competing financial interest.

**List of abbreviations:** Alzheimer’s Disease, AD; upper limit of normal, ULN; alanine transaminase, ALT; drug-induced liver injury, DILI; peripheral blood mononuclear cells, PBMC; epstein barr virus, EBV; human leukocyte antigen, HLA.

**Figure Legends**

#### Figure 1. Study compound structures, granzyme B PBMC ELIspot and initial testing of T-cell clones from patient 6. (A) Structure of atabecestat and its metabolic conversion to DIAT and atabecestat epoxide. (B) PBMCs were incubated in a granzyme BELIspot plate with atabecestat, DIAT, acetyl DIAT and atabecestat epoxide for a period of 48 hours. The ELIspot plate was developed and counted using ELIspot AID reader. (C) T-cell clones from atabecestat-, DIAT-, acetyl DIAT- and atabecestat epoxide-derived T-cell lines (i.e., each figure panel shows separate clones)were co-incubated with autologous antigen presenting cells in the presence and absence of atabecestat (10 μM), DIAT (25 μM), acetyl DIAT (25μM) and atabecestat epoxide (10μM). Cells were incubated for 48 hours. [3H] thymidine was then added for an additional 16 hour incubation and T-cell proliferation was assessed using scintillation counting. Clones with a stimulation index of 1.5 or greater were selected and expanded for further analysis. Cross-reactivity was not possible at this early phase of the analysis.

**Figure 2. Analysis of cytokine and cytolytic molecule secretion from atabecestat- and DIAT-responsive T-cell clones.** T-cell clones were co-incubated with autologous EBV-transformed antigen presenting cells in the presence and absence of atabecestat (25µM) or DIAT (100µM) in an ELIspot plate coated with antibodies for IFN-γ, granzyme B (GrB), IL-5, IL-13, IL-10, IL-17, IL-22 and perforin for 48 hours. Following incubation, the ELIspot plate was developed for the respective cytokines.

**Figure 3. HLA-DR restriction of atabecestat and DIAT-responsive CD4+ T-cell clones.** T-cell clones were co-incubated with autologous EBV-transformed antigen presenting cells in the presence and absence of atabecestat (25µM) or DIAT (100µM) for 48 hours. Prior to addition of the drugs, blocking antibodies; Isotype I, Isotype II, HLA Class I and HLA Class II were co-incubated with antigen presenting cells and T-cell clones for 1 hour. The assay was also conducted using HLA Class II sub-class-specific antibodies HLA-DP, DQ and DR. T-cell responses were quantified via analysis of (A) proliferation using [3H] thymidine and scintillation counting or IFN-γ secretion by way of ELIspot.

**Figure 4. Direct activation of CD4+ T-cell clones with atabecestat and DIAT.** (A) T-cell clones were co-incubated with autologous EBV-transformed B-cells pre-pulsed with atabecestat (25µM) or DIAT (100µM) for 1 or 16 hours. The study compound-pulsed EBV-transformed B-cells were washed repeatedly to remove free compound prior to culturing with T-cells. (B) T-cell clones were incubated with atabecestat (25µM) or DIAT (100µM) in the presence or absence of autologous EBV-transformed B-cells (APC) fixed with glutaraldehyde to prevent protein processing. T-cell responses were measured either via analysis of proliferation through addition of [3H] thymidine and scintillation counting or through secretion of IFN-γ via ELIspot.

**Figure 5. Cross-reactivity of study compound-specific T-cell clones.** (A) Energy minimized 3D-structures of study compounds. (B) Proliferation of representative T-cell clones in the presence of study compounds. T-cell clones were co-incubated with autologous EBV-transformed B-cells in the presence and absence atabecestat, DIAT, atabecestat epoxide and acetyl DIAT for 48 hours. [3H] thymidine was added for an additional 16 hours and proliferation was measured via scintillation counting. Error bars denote ± SEM. (C) IFN-γ secretion from study compound stimulated T-cell clones. T-cell clones were co-incubated with autologous EBV-transformed B-cells in the presence and absence of atabecestat, DIAT, atabecestat epoxide and acetyl DIAT in a ELIspot plate for 48 hours. Following incubation, the ELIspot was developed for IFN-γ.

## Tables

**Table 1. Patient demographics and details of the adverse events**

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Patient id** | **Gender** | **Age** | **Country of origin** | **First study day ALT >3xULN** | **Maximum ALT increase (xULN)** | **Dose at time of reaction** | **Time to study blood sampling following 1st ALT > 3X ULN** | **PBMC proliferation assay** | **PBMC ELIspot** | |
| **IFN-γ** | **Granzyme B** |
| Patient 1 | F | 71 | Belgium | D 260 | 4.9 | 10 mg | 758 | -ve1 | -ve | -ve |
| Patient 2 | F | 66 | Belgium | / | / | 5 mg | / | -ve | -ve | -ve |
| Patient 3 | M | 68 | Belgium | D 168 | 4.5 | 10 mg | 1024 | -ve | n/a2 | n/a |
| Patient 4 | F | 75 | France | D 155 | 8.4 | 5 mg | 468 | -ve | -ve | -ve |
| Patient 5 | M | 63 | Sweden | D 288 | 7.5 | 50 mg | 691 | -ve | -ve | -ve |
| Patient 6 | F | 76 | UK | D 32 | 28.8 | 5 mg | 544 | -ve | -ve | **+ve** |
| Patient 7 | M | 73 | Spain | D 344 | 6.5 | 10 mg | 750 | -ve | -ve | -ve |
| Patient 8 | M | 75 | Belgium | / | / | 50 mg | / | n/a | n/a | n/a |
| Patient 9 | M | 57 | Germany | D 84 | 15.7 | 50 mg | 975 | -ve | -ve | -ve |
| Patient 10 | F | 68 | France | D 260 | 9.4 | 25 mg | 377 | n/a | n/a | n/a |
| Patient 11 | F | 67 | Australia | D 58 | 29.1 | 25 mg | 306 | n/a | n/a | n/a |
| Patient 12 | F | 64 | UK | D 115 | 8.6 | 25 mg | 152 | -ve | -ve | -ve |
| Patient 13 | F | 67 | UK | D 150 | 4.6 | 5 mg | 141 | -ve | -ve | -ve |
| Patient 14 | F | 76 | USA | D 84 | 3.8 | 25 mg | 84 | n/a | n/a | n/a |

1 result considered positive when stimulation index (response in drug treated/response in medium control) greater than 2.

2 N/A – Assay not conducted due to low cell number/viability

/ not applicable because no ALT elevation

**Supplementary Figure 1. Specificity of T-cell clones and characterisation of cellular surface phenotype.** (A) T-cell clones were co-incubated with autologous EBV-transformed B-cells in the presence of either atabecestat, DIAT, or acetyl DIAT 48 hours. [3H] thymidine was added for an additional 16 hours and proliferation was measured via scintillation counting. Error bars denote ± SEM. (B) CD4/8 phenotype of T-cell clones. T-cells were stained with antibodies CD4-FITC/APC and CD8-PE and analyzed for phenotype using flow cytometry. CD4/CD8/DP (double positive) clones expressed high levels of CD4 and CD8. (C) T-cells were stained with T-cell Vβ receptor antibodies. Cells were then washed and analysed for TCRVβ expression using flow cytometry. Rare refers to clones where V β receptor was not detected. (d) Cells were stained with antibodies CCR2, CXCR3, CCR1, CCR8, CCR9,CXCR1, CLA, CCR6, CXCR6, CD69, E-cadherin and CCR4. Cells were then washed and analysed for surface receptor expression using flow cytometry. Receptor was deemed to be expressed if mean staining intensity exceeded twice the isotype control.

**Supplementary Figure 2. IFN-γ release from an acetyl DIAT-responsive T-cell clone.** (a) The T-cell clone was co-incubated with autologous EBV-transformed antigen presenting cells in the presence and absence of atabecestat, DIAT, atabecestat epoxide and acetyl DIAT for 48 hours. Following incubation, the ELIspot was developed for IFN-γ.

## References

1. Sanabria-Castro A, Alvarado-Echeverria I, Monge-Bonilla C. Molecular Pathogenesis of Alzheimer's Disease: An Update. Ann Neurosci. 2017;24(1):46-54.

2. Henley D, Raghavan N, Sperling R, Aisen P, Raman R, Romano G. Preliminary Results of a Trial of Atabecestat in Preclinical Alzheimer's Disease. N Engl J Med. 2019;380(15):1483-5.

3. De Jonghe S, Weinstock D, Aligo J, Washington K,Naisbitt D.Biopsy pathology and immunohistochemistry of a case of immune-mediated drug-induced liver injury with atabecestat. Hepatology. 2020 [Epub ahead of print doi: 10.1002/hep.31403].

4. Maria VA, Victorino RM. Diagnostic value of specific T cell reactivity to drugs in 95 cases of drug induced liver injury. Gut. 1997;41(4):534-40.

5. Monshi MM, Faulkner L, Gibson A, Jenkins RE, Farrell J, Earnshaw CJ, Alfirevic A, Cederbrant K, Daly AK, French N, Pirmohamed M, Park BK, Naisbitt DJ. Human leukocyte antigen (HLA)-B\*57:01-restricted activation of drug-specific T cells provides the immunological basis for flucloxacillin-induced liver injury. Hepatology. 2013;57(2):727-39.

6. Kim SH, Saide K, Farrell J, Faulkner L, Tailor A, Ogese M, Daly AK, Pirmohamed M, Park BK, Naisbitt DJ. Characterization of amoxicillin- and clavulanic acid-specific T cells in patients with amoxicillin-clavulanate-induced liver injury. Hepatology. 2015;62(3):887-99.

7. Usui T, Meng X, Saide K, Farrell J, Thomson P, Whitaker P, Watson J, French NS, Kevin Park B, Naisbitt DJ. From the Cover: Characterization of Isoniazid-Specific T-Cell Clones in Patients with anti-Tuberculosis Drug-Related Liver and Skin Injury. Toxicol Sci. 2017;155(2):420-31.

8. Daly AK, Donaldson PT, Bhatnagar P, Shen Y, Pe'er I, Floratos A, Daly MJ, Goldstein DB, John S, Nelson MR, Graham J, Park BK, Dillon JF, Bernal W, Cordell HJ, Pirmohamed M, Aithal GP, Day CP. HLA-B\*5701 genotype is a major determinant of drug-induced liver injury due to flucloxacillin. Nat Genet. 2009;41(7):816-9.

9. Hirasawa M, Hagihara K, Abe K, Ando O, Hirayama N. In Silico and In Vitro Analysis of Interaction between Ximelagatran and Human Leukocyte Antigen (HLA)-DRB1\*07:01. Int J Mol Sci. 2017;18(4).

10. Singer JB, Lewitzky S, Leroy E, Yang F, Zhao X, Klickstein L, Wright TM, Meyer J, Paulding CA. A genome-wide study identifies HLA alleles associated with lumiracoxib-related liver injury. Nat Genet. 2010;42(8):711-4.

11. Konvinse KC, Trubiano JA, Pavlos R, James I, Shaffer CM, Bejan CA, Schutte RJ, Ostrov DA, Pilkinton MA, Rosenbach M, Zwerner JP, Williams KB, Bourke J, Martinez P, Rwandamuriye F, Chopra A, Watson M, Redwood AJ, White KD, Mallal SA,Phillips EJ. HLA-A\*32:01 is strongly associated with vancomycin-induced drug reaction with eosinophilia and systemic symptoms. J Allergy Clin Immunol. 2019;144(1):183-92.

12. Zhang FR, Liu H, Irwanto A, Fu XA, Li Y, Yu GQ, Yu YX, Chen MF, Low HQ, Li JH, Bao FF, Foo JN, Bei JX, Jia XM, Liu J, Liany H, Wang N, Niu GY, Wang ZZ, Shi BQ, Tian HQ, Liu HX, Ma SS, Zhou Y, You JB, Yang Q, Wang C, Chu TS, Liu DC, Yu XL, Sun YH, Ning Y, Wei ZH, Chen SL, Chen XC, Zhang ZX, Liu YX, Pulit SL, Wu WB, Zheng ZY, Yang RD, Long H, Liu ZS, Wang JQ, Li M, Zhang LH, Wang H, Wang LM, Xiao P, Li JL, Huang ZM, Huang JX, Li Z, Liu J, Xiong L, Yang J, Wang XD, Yu DB, Lu XM, Zhou GZ, Yan LB, Shen JP, Zhang GC, Zeng YX, de Bakker PI, Chen SM, Liu JJ.. HLA-B\*13:01 and the dapsone hypersensitivity syndrome. N Engl J Med. 2013;369(17):1620-8.

13. Faulkner L, Gibson A, Sullivan A, Tailor A, Usui T, Alfirevic A, Pirmohamed M, Naisbitt DJ, Kevin Park B.. Detection of Primary T Cell Responses to Drugs and Chemicals in HLA-Typed Volunteers: Implications for the Prediction of Drug Immunogenicity. Toxicol Sci. 2016;154(2):416-29.

14. Sebastiani S, Allavena P, Albanesi C, Nasorri F, Bianchi G, Traidl C, Sozzani S, Girolomoni G, Cavani A. Chemokine receptor expression and function in CD4+ T lymphocytes with regulatory activity. J Immunol. 2001;166(2):996-1002.

15. Bonecchi R, Bianchi G, Bordignon PP, D'Ambrosio D, Lang R, Borsatti A, Sozzani S, Allavena P, Gray PA, Mantovani A, Sinigaglia F. Differential expression of chemokine receptors and chemotactic responsiveness of type 1 T helper cells (Th1s) and Th2s. J Exp Med. 1998;187(1):129-34.

16. Nicoletti P, Aithal GP, Bjornsson ES, Andrade RJ, Sawle A, Arrese M, Barnhart HX, Bondon-Guitton E, Hayashi PH, Bessone F, Carvajal A, Cascorbi I, Cirulli ET, Chalasani N, Conforti A, Coulthard SA, Daly MJ, Day CP, Dillon JF, Fontana RJ, Grove JI, Hallberg P, Hernández N, Ibáñez L, Kullak-Ublick GA, Laitinen T, Larrey D, Lucena MI, Maitland-van der Zee AH, Martin JH, Molokhia M, Pirmohamed M, Powell EE, Qin S, Serrano J, Stephens C, Stolz A, Wadelius M, Watkins PB, Floratos A, Shen Y, Nelson MR, Urban TJ, Daly AK. Association of Liver Injury From Specific Drugs, or Groups of Drugs, With Polymorphisms in HLA and Other Genes in a Genome-Wide Association Study. Gastroenterology. 2017;152(5):1078-89.

17. Lucena MI, Molokhia M, Shen Y, Urban TJ, Aithal GP, Andrade RJ, Day CP, Ruiz-Cabello F, Donaldson PT, Stephens C, Pirmohamed M, Romero-Gomez M, Navarro JM, Fontana RJ, Miller M, Groome M, Bondon-Guitton E, Conforti A, Stricker BH, Carvajal A, Ibanez L, Yue QY, Eichelbaum M, Floratos A, Pe'er I, Daly MJ, Goldstein DB, Dillon JF, Nelson MR, Watkins PB, Daly AK. Susceptibility to amoxicillin-clavulanate-induced liver injury is influenced by multiple HLA class I and II alleles. Gastroenterology. 2011;141(1):338-47.

18. Urban TJ, Nicoletti P, Chalasani N, Serrano J, Stolz A, Daly AK, Aithal GP, Dillon J, Navarro V, Odin J, Barnhart H, Ostrov D, Long N, Cirulli ET, Watkins PB, Fontana RJ;. Minocycline hepatotoxicity: Clinical characterization and identification of HLA-B \*35:02 as a risk factor. J Hepatol. 2017;67(1):137-44.

19. Mennicke M, Zawodniak A, Keller M, Wilkens L, Yawalkar N, Stickel F, Keogh A, Inderbitzin D, Candinas D, Pichler WJ. Fulminant liver failure after vancomycin in a sulfasalazine-induced DRESS syndrome: fatal recurrence after liver transplantation. Am J Transplant. 2009;9(9):2197-202.

20. Wuillemin N, Terracciano L, Beltraminelli H, Schlapbach C, Fontana S, Krahenbuhl S, Pichler WJ, Yerly D. T Cells Infiltrate the Liver and Kill Hepatocytes in HLA-B(\*)57:01-Associated Floxacillin-Induced Liver Injury. Am J Pathol. 2014;184(6):1677-82.

21. Meng X, Al-Attar Z, Yaseen FS, Jenkins R, Earnshaw C, Whitaker P, Peckham D, French NS, Naisbitt DJ, Park BK. Definition of the Nature and Hapten Threshold of the beta-Lactam Antigen Required for T Cell Activation In Vitro and in Patients. J Immunol. 2017;198(11):4217-27.

22. Jenkins RE, Meng X, Elliott VL, Kitteringham NR, Pirmohamed M, Park BK. Characterisation of flucloxacillin and 5-hydroxymethyl flucloxacillin haptenated HSA in vitro and in vivo. Proteomics Clin Appl. 2009;3(6):720-9.

23. Bechara R, Maillere B, Joseph D, Weaver RJ, Pallardy M. Identification and characterization of a naive CD8+ T cell repertoire for benzylpenicillin. Clin Exp Allergy. 2019;49(5):636-43.

24. Azoury ME, Fili L, Bechara R, Scornet N, de Chaisemartin L, Weaver RJ, Claude N, Maillere B, Parronchi P, Joseph D, Pallardy M.. Identification of T-cell epitopes from benzylpenicillin conjugated to human serum albumin and implication in penicillin allergy. Allergy. 2018;73(8):1662-72.

25. Scornet N, Delarue-Cochin S, Azoury ME, Le Mignon M, Chemelle JA, Nony E, Maillère B, Terreux R, Pallardy M, Joseph D.. Bioinspired Design and Oriented Synthesis of Immunogenic Site-Specifically Penicilloylated Peptides. Bioconjug Chem. 2016;27(11):2629-45.

26. Timmers M, Streffer JR, Russu A, Tominaga Y, Shimizu H, Shiraishi A, Tatikola K, Smekens P, Börjesson-Hanson A, Andreasen N, Matias-Guiu J, Baquero M, Boada M, Tesseur I, Tritsmans L, Van Nueten L, Engelborghs S. Pharmacodynamics of atabecestat (JNJ-54861911), an oral BACE1 inhibitor in patients with early Alzheimer's disease: randomized, double-blind, placebo-controlled study. Alzheimers Res Ther. 2018;10(1):85.

27. Gibson A, Faulkner L, Lichtenfels M, Ogese M, Al-Attar Z, Alfirevic A, Alfirevic A, Esser PR, Martin SF, Pirmohamed M, Park BK, Naisbitt DJ. The Effect of Inhibitory Signals on the Priming of Drug Hapten-Specific T Cells That Express Distinct Vbeta Receptors. J Immunol. 2017;199(4):1223-37.

28. Naisbitt DJ, Olsson-Brown A, Gibson A, Meng X, Ogese MO, Tailor A, Thomson P. Immune dysregulation increases the incidence of delayed-type drug hypersensitivity reactions. Allergy. 2020 Apr;75(4):781-797.

# 29. Yamazaki N, Uhara H, Fukushima S, Uchi H, Shibagaki N, Kiyohara Y, Tsutsumida A, Namikawa K, Okuyama R, Otsuka Y, Tokudome T. Phase II Study of the Immune-Checkpoint Inhibitor Ipilimumab Plus Dacarbazine in Japanese Patients With Previously Untreated, Unresectable or Metastatic Melanoma. Cancer Chemother Pharmacol. 2015;76(5):969-75.

30. Novak G, Streffer JR, Timmers M, Henley D, Brashear HR, Bogert J, Russu A, Janssens L, Tesseur I, Tritsmans L, Van Nueten L, Engelborghs S. Long-term safety and tolerability of atabecestat (JNJ-54861911), an oral BACE1 inhibitor, in early Alzheimer's disease spectrum patients: a randomized, double-blind, placebo-controlled study and a two-period extension study.  Alzheimers Res Ther. 2020;12:58.

31. Zhao Q, Alhilali K, Alzahrani A, Almutairi M, Amjad J, Liu H, Sun Y, Sun L, Zhang H, Meng X, Gibson A, Ogese MO, Kevin Park B, Liu J, Ostrov DA, Zhang F, Naisbitt DJ. Dapsone- and nitroso dapsone-specific activation of T cells from hypersensitive patients expressing the risk allele HLA-B\*13:01. Allergy. 2019;74(8):1533-48.

32. Couper KN, Blount DG, Riley EM. IL-10: the master regulator of immunity to infection. J Immunol. 2008;180(9):5771-7.

33. Wuillemin N, Adam J, Fontana S, Krahenbuhl S, Pichler WJ, Yerly D. HLA haplotype determines hapten or p-i T cell reactivity to flucloxacillin. J Immunol. 2013;190(10):4956-64.

34. Castrejon JL, Berry N, El-Ghaiesh S, Gerber B, Pichler WJ, Park BK, Naisbitt DJ. Stimulation of human T cells with sulfonamides and sulfonamide metabolites. J Allergy Clin Immunol. 2010;125(2):411-8 e4.

35. Illing PT, Vivian JP, Dudek NL, Kostenko L, Chen Z, Bharadwaj M, Miles JJ, Kjer-Nielsen L, Gras S, Williamson NA, Burrows SR, Purcell AW, Rossjohn J, McCluskey J. Immune self-reactivity triggered by drug-modified HLA-peptide repertoire. Nature. 2012;486(7404):554-8.

36. Ostrov DA, Grant BJ, Pompeu YA, Sidney J, Harndahl M, Southwood S, Oseroff C, Lu S, Jakoncic J, de Oliveira CA, Yang L, Mei H, Shi L, Shabanowitz J, English AM, Wriston A, Lucas A, Phillips E, Mallal S, Grey HM, Sette A, Hunt DF, Buus S, Peters B. Drug hypersensitivity caused by alteration of the MHC-presented self-peptide repertoire. Proc Natl Acad Sci U S A. 2012;109(25):9959-64.

37. Adam J, Eriksson KK, Schnyder B, Fontana S, Pichler WJ, Yerly D. Avidity determines T-cell reactivity in abacavir hypersensitivity. Eur J Immunol. 2012;42(7):1706-16.

38. Schnyder B, Burkhart C, Schnyder-Frutig K, von Greyerz S, Naisbitt DJ, Pirmohamed M, Park BK, Pichler WJ. Recognition of sulfamethoxazole and its reactive metabolites by drug-specific CD4(+) T cells from allergic individuals. J Immunol. 2000;164(12):6647-54.

39. Wu Y, Sanderson JP, Farrell J, Drummond NS, Hanson A, Bowkett E, Berry N, Stachulski AV, Clarke SE, Pichler WJ, Pirmohamed M, Park BK, Naisbitt DJ. Activation of T cells by carbamazepine and carbamazepine metabolites. J Allergy Clin Immunol. 2006;118(1):233-41.

40. Burkhart C, von Greyerz S, Depta JP, Naisbitt DJ, Britschgi M, Park KB, Pichler WJ. Influence of reduced glutathione on the proliferative response of sulfamethoxazole-specific and sulfamethoxazole-metabolite-specific human CD4+ T-cells. Br J Pharmacol. 2001;132(3):623-30.

41. Nassif A, Bensussan A, Boumsell L, Deniaud A, Moslehi H, Wolkenstein P,  Bagot M, Roujeau JC. Toxic epidermal necrolysis: effector cells are drug-specific cytotoxic T cells. J Allergy Clin Immunol. 2004;114(5):1209-15.

42. Yaseen FS, Saide K, Kim SH, Monshi M, Tailor A, Wood S, Meng X, Jenkins R, Faulkner L, Daly AK, Pirmohamed M, Park BK, Naisbitt DJ. Promiscuous T-cell responses to drugs and drug-haptens. J Allergy Clin Immunol. 2015;136(2):474-6 e8.

43. Zanni MP, von Greyerz S, Schnyder B, Brander KA, Frutig K, Hari Y, Valitutti S, Pichler WJ. HLA-restricted, processing- and metabolism-independent pathway of drug recognition by human alpha beta T lymphocytes. J Clin Invest. 1998;102(8):1591-8.

44. [Pichler WJ. Immune pathomechanism and classification of drug hypersensitivity.](https://pubmed.ncbi.nlm.nih.gov/30843233/) Allergy. 2019;74(8):1457-71.

45. Pan RY, Chu MT, Wang CW, Lee YS, Lemonnier F, Michels AW, Schutte R, Ostrov DA, Chen CB, Phillips EJ, Mallal SA, Mockenhaupt M, Bellón T, Tassaneeyakul W, White KD, Roujeau JC, Chung WH, Hung SI. Identification of drug-specific public TCR driving severe cutaneous adverse reactions. Nat Commun. 2019;10(1):3569.